Organic Solutes in Ground Water at the Idaho National Engineering Laboratory

(U.S.) Geological Survey Idaho Falls, ID

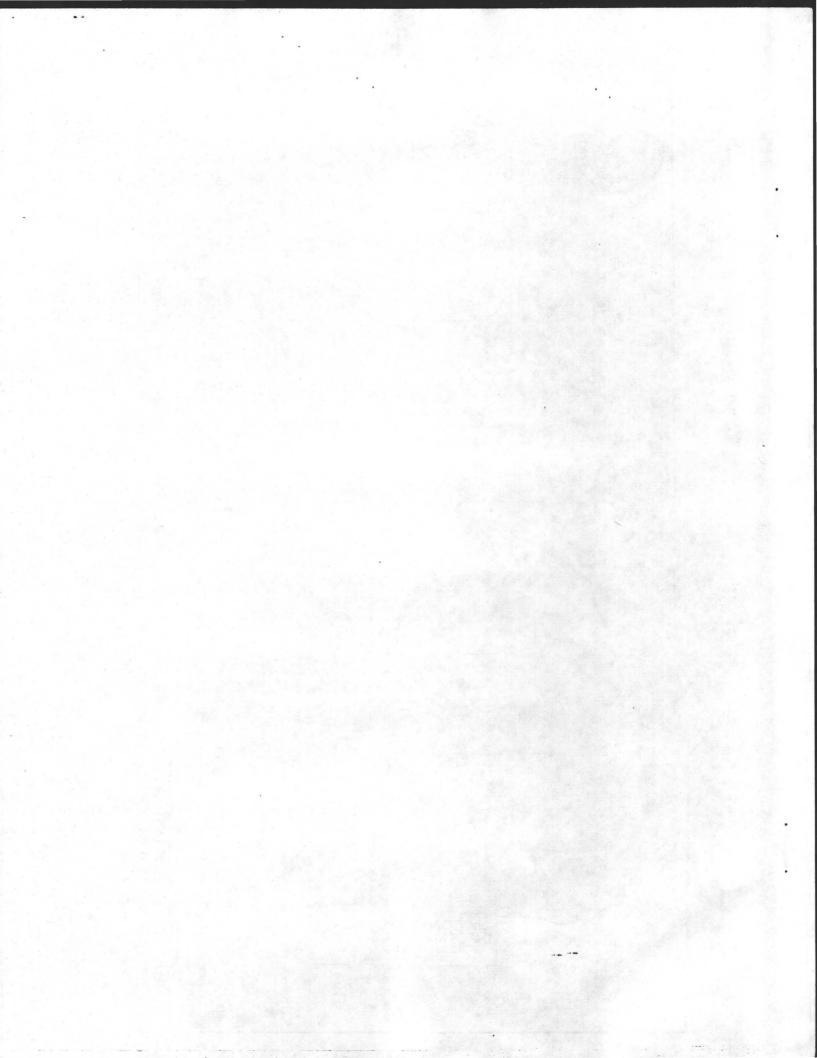
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herbicides. Fourteen wells and four potential organic sources were sampled for volatile and semivolatile organic compounds. The results of the DOC analyses indicate no high level organic contamination of INEL

ground water. The only detectable insecticide or herbicide was a DDT concentration of 10 parts per trillion (0.01 micrograms per liter in one observation well).

The volatile and semivolatile analyses do not indicate the presence of hazardous organic contaminants in significant amounts (>10 micrograms per liter in the samples taken).

Due to the lack of any significant organic ground-water contamination in this reconnaissance survey, the second phase of the study, which was to follow up the first phase by additional sampling of any contaminated wells, was cancelled.

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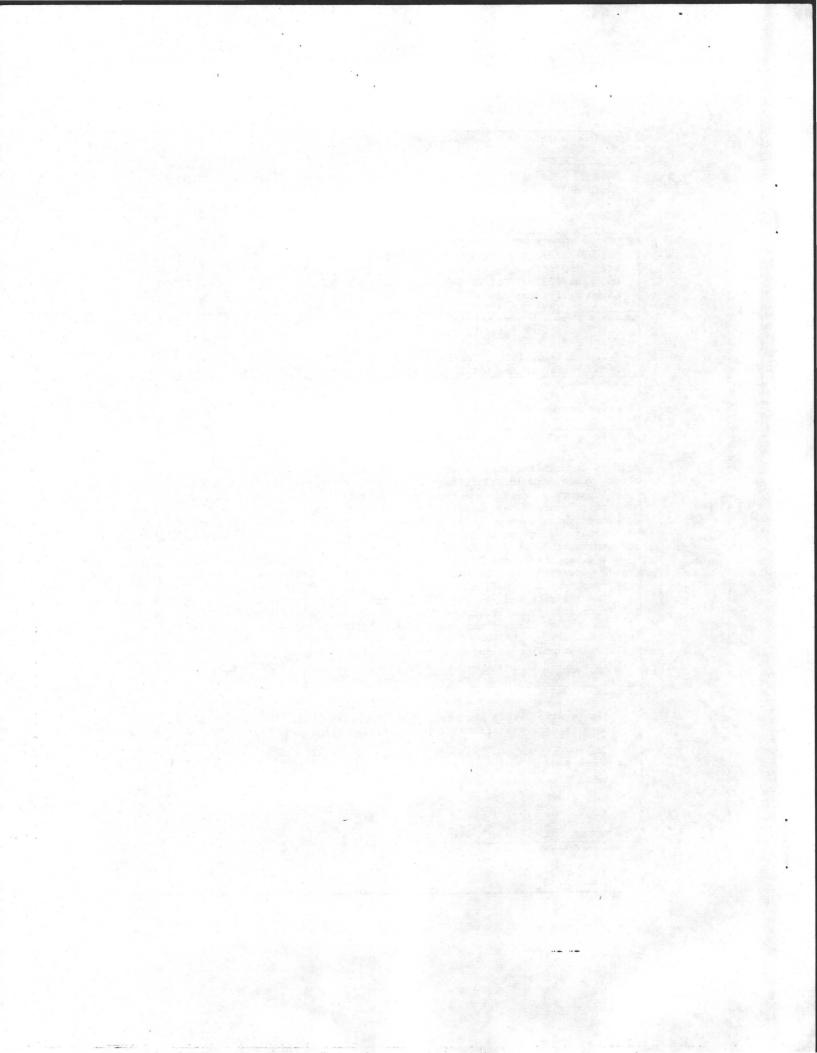
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ORGANIC SOLUTES IN GROUND WATER AT THE IDAHO NATIONAL ENGINEERING LABORATORY

By Jerry A. Leenheer and Jefferson C. Bagby

U.S. GEOLOGICAL SURVEY

Water-Resources Investigation 82-15

Prepared on behalf of the U.S. DEPARTMENT OF ENERGY



March 1982

I.a

UNITED STATES DEPARTMENT OF THE INTERIOR

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CONTENTS

Page

Abstract	•	•		•	•				•		•	•	1
Introduction	•	•				•	•	•	•				1
Purpose of study	•					•	•					•	2
Acknowledgments	•	•	•			•							4
Regional hydrology	•		•		•								4
Organic sampling program	•	•											5
Sampling site selection													
Field sampling techniques													
Dissolved organic carbon .													
Insecticides, herbicides, and													
Volatiles													
Analytical methods													
Results and discussion													
Dissolved organic carbon .													
Insecticides and herbicides													
Volatiles and semivolatiles .													
Cost	•												36
Summary and conclusions													
Selected references													37

ILLUSTRATIONS

Figure	1.	Relief map of Idaho showing the location of the INEL, Snake River Plain and generalized ground- water flow lines of the Snake River Plain
	2.	Distribution of dissolved organic carbon in ground-
		water, INEL vicinity, August 1980 6
	3.	Distribution of dissolved organic carbon in ground
		water, ICPP-TRA vicinity, August 1980 7
	4.	Photograph of bailer and thief sampler used for
		collecting water samples from wells without
		pump

TABLES

Table	1.	Organic sample collection parameters and dissolved	
		organic carbon results	8
	2.	Dissolved organic carbon concentrations in water	
		from background wells compared to method of	
		well sampling	L7
	3.	Gas chromatographic-mass spectrometric reconnaissance	
		survey of extractable and volatile organic	-
		solutes in ground water at the Idaho National	
		Engineering Laboratory	19

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The following factors can be used to convert inch-pound units published herein to the International System (SI) of units.

Multiply inch-pound units	By	To obtain metric units
feet (ft) inches (in.)	0.3048	meters (m) millimeter (mm)
miles (mi) square miles (mi ²) gallons (gal)	1.609 2.590 3.785 3.785x10-3 3.785x103 1.233x103	kilometers (km) square kilometers (km ²) liters (L) cubic meters (m ³) cubic meters (m ³) cubic meters (m ³)

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By

Jerry A. Leenheer and Jefferson C. Bagby

ABSTRACT

In August 1980, the U.S. Geological Survey (USGS) started a reconnaissance survey of organic solutes in drinking water sources, groundwater monitoring wells, perched water-table monitoring wells, and in select waste streams at the Idaho National Engineering Laboratory (INEL).

The survey was to be a two-phase program. In the first phase, 77 wells and four potential point sources were sampled for dissolved organic carbon (DOC). Four wells and several potential point sources of insecticides and herbicides were sampled for insecticides and herbicides. Fourteen wells and four potential organic sources were sampled for volatile and semivolatile organic compounds.

The results of the DOC analyses indicate no high level (>20 mg/L DOC) organic contamination of ground water. The only detectable insecticide or herbicide was a dichlorodiphenyltrichloroethane (DDT) concentration of 10 parts per trillion (0.01 μ g/L) in one observation well.

The volatile and semivolatile analyses do not indicate the presence of hazardous organic contaminants in significant amounts (>10 $\mu\,g/L$) in the samples taken.

Due to the lack of any significant organic ground-water contamination in this reconnaissance survey, the second phase of the study, which was to follow up the first phase by additional sampling of any contaminated wells, was canceled.

INTRODUCTION

The Idaho National Engineering Laboratory (INEL) formerly the National Reactor Testing Station (NRTS), was established in 1949 by the United States Atomic Energy Commission (AEC) (later reorganized as Energy Research and Development Administration (ERDA), and now as the Department of Energy (DOE)) to build, operate, and test various types of nuclear reactors. The reactors are built primarily to develop peacetime uses of atomic energy. Fifty-two reactors have been constructed to date, of which 17 are still operable. The INEL site covers about 890 square miles on the eastern Snake River Plain (fig. 1) and has an average altitude of 4,900 feet above sea level. This plain is underlain by the Snake River Plain aquifer, the major aquifer in Idaho. The INEL obtains its entire water supply from this aquifer. Aqueous chemical and radioactive wastes are discharged to shallow ponds and to shallow or deep wells. The pond and shallow-well wastes infiltrate the ground, form perched bodies of water, and then percolate toward the Snake River Plain aquifer.

For the past 30 years, the U.S. Geological Survey (USGS), Water Resources Division (WRD), has maintained a research project office at the INEL. One of the primary functions of this office has been to investigate the geochemical conditions of the Snake River Plain aquifer (Barraclough and others, 1967; Robertson, 1974; Barraclough and others, 1976; Robertson, 1977; Barraclough and others, 1981).

Purpose of Study

In January 1980, the U.S. Environmental Protection Agency (EPA) recommended that a ground-water monitoring program for organic constituents be started at the INEL. In response to this recommendation, DOE requested the U.S. Geological Survey to undertake such a program. The purpose of the study was to determine the distribution and extent of organic constituents in the ground water beneath the INEL.

The U.S. Geological Survey proposed a two-phase organic sampling The first phase would be an organic reconnaissance survey of program. the ground water beneath the INEL. This would entail a large scale dissolved organic carbon (DOC) sampling program of INEL production and ground-water observation wells to identify areas containing higher levels of organic constituents (>20 mg/L DOC). Many of the samples would be collected around the Idaho Chemical Processing Plant (ICPP) and the Test Reactor Area (TRA). The ICPP and the TRA dispose of cooling tower, chemical, and radioactive wastes through the use of disposal wells and/or waste-seepage ponds and thus are potential sources of organic contamination of ground water. Samples for specific organic analysis of pesticides, volatile, and solvent-extractable semivolatile compounds would also be collected near potential contamination sources to detect organic chemicals for which EPA has established maximum contaminant levels (U.S. Environmental Protection Agency, 1976).

If the first phase organic reconnaissance survey had established any organic contamination problem areas, the second phase of the program would have begun. In this phase, ground waters with high DOC values would have been characterized in greater detail by DOC fractionation analysis (Leenheer and Huffman, 1979), in order to ascertain the nature and origin of the DOC ground-water contamination. Additional qualitative and quantitative organic analyses for specific compounds would also be performed on whole waters and fractions from the DOC fractionation procedure.

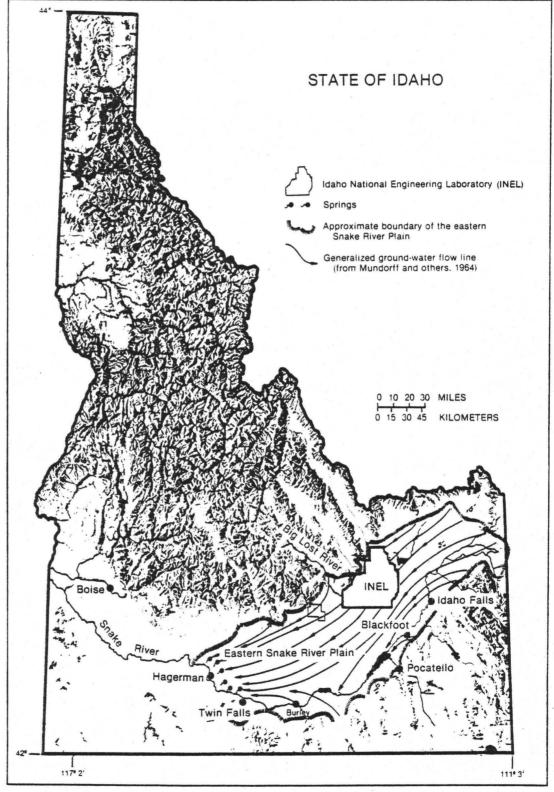


Figure 1. Relief map of Idaho showing the location of the INEL, Snake River Plain and generalized ground water flow lines of the Snake River Plain aquifer.

In August 1980, the USGS collected water samples for phase one. This report contains a brief explanation of how the sampling locations were selected, how the samples were collected, and how the samples were analyzed. It also contains a complete presentation of the data collected, the interpretation thereof, and a suggestion concerning future organic monitoring at the INEL.

Acknowledgments

The U.S. Geological Survey wishes to express appreciation to the DOE, EG&G Idaho, Inc., EXXON Nuclear Idaho Company, Inc., and Westinghouse Electric Corporation employees who helped, through their cooperation, to complete phase one of this study. Special thanks go to the employees of the National Water Quality Laboratory (NWQL) in Denver for their prompt analysis of a large number of samples and their aid in the interpretation of the results.

Regional Hydrology

The eastern Snake River Plain is a large structural basin 12,000 square miles in area (fig. 1). It has been filled to its present level with 2,000 to 10,000 feet of thin basaltic lava flows, rhyolitic deposits, and interbedded sediments. A more detailed description of the geology is found in Robertson, Schoen, and Barraclough (1974). Nearly all the eastern Snake River Plain is underlain by a vast ground-water reservoir known as the Snake River Plain aquifer, which contains an estimated 1 billion acre-feet of water. The flow of ground water in the aquifer is principally to the south-southwest (fig. 1) at relatively high velocities, generally 5 to 25 feet per day (Robertson, Schoen, and Barraclough, 1974, p. 13). Transmissivity of the aquifer is high, generally ranging from 1 million to 100 million gallons per day per foot or 134,000 to 13,400,000 feet squared per day (Robertson, Schoen, and Barraclough, 1974, p. 12).

The basaltic volcanic rocks and interbedded sediments composing the aquifer are all included in the Snake River Group of Quaternary age. The basement rocks are probably composed of older volcanic and sedimentary rocks. The basalt is the principal aquifer. The water-bearing openings in the basalt are distributed throughout the rock system in the form of intercrystalline and intergranular porespace, fractures, cavities, interstitial voids, interflow zones, and lava tubes. The variety and degree of interconnection of these openings complicates the direction of ground-water movement locally throughout the aquifer.

Ground-water recharge to the INEL is derived primarily as underflow from the northeastern part of the Plain and also from adjacent drainages on the west and north. Most of the ground water underlying the INEL enters the ground in the uplands to the north, northeast, and northwest; moves south or southwestward through the aquifer; and discharges through springs along the valley of the Snake River near Hagerman (fig. 1). Lesser amounts of recharge are derived from direct precipitation on the Plain. Some of the precipitation evaporates, and some infiltrates the ground surface and percolates through the unsaturated subsurface to the regional water table.

ORGANIC SAMPLING PROGRAM

Sampling Site Selection

The sites selected for DOC sampling were placed into two separate groups based on two criteria. The first group, consisting of ten wells, was chosen as background wells for ground-water DOC baseline data. These background wells were chosen because they are remote from any possible INEL organic input and include the Atomic City well, Fire Station #2, Highway #3, P&W-2, Blaine Larsen's well, Site 14, and USGS observation wells 4, 19, 26, and 27 (fig. 2). Five of the wells selected are equipped with pumps. The other five wells are not, and required a thief sampling to obtain a comparison of the two sampling methods.

The second group of wells was chosen because they are used for drinking water or because of their close proximity to a potential source of organic ground-water contamination. For the purpose of this study, any INEL facility was considered to be a potential source of organic ground-water contamination.

A total of seventy-seven DOC samples was collected from seventy-four surface- and ground-water sources across the INEL. These samples provided the data base for the phase one study. Two water samples each were collected from the Site 14 well, well 65, and well 47, to check sampling repeatability.

The locations of all wells sampled are indicated on the maps in figures 2 and 3. Additional information on each well is given in table 1.

Samples for insecticide and herbicide analyses were taken from the ICPP east side waste effluent and the TRA cooling tower effluent. Both of these effluent streams are discharged directly into the Snake River Plain aquifer and thus could be point sources for organic ground-water contamination. Samples for insecticide and herbicide analysis were also taken from wells 40, 43, 47, and 67. These wells are all near the ICPP disposal well which has had low concentrations of insecticides and herbicides discharged into it in the past. If insecticides or herbicides could be detected in the ground water, these wells would be the most likely to show them (as was shown by the results).

Volatile and semivolatile organic samples were collected for GC-MS analysis from the following possible organic contaminant sources: (1) ICPP east side waste effluent (disposal well), (2) TRA chemical waste pond, (3) TRA cooling tower waste effluent (disposal well), and (4) TRA radioactive waste pond. Samples were also collected from observation wells 40, 43, 46, 47, 53, 54, 55, 56, 57, 58, 65, 67, 68, and 70.

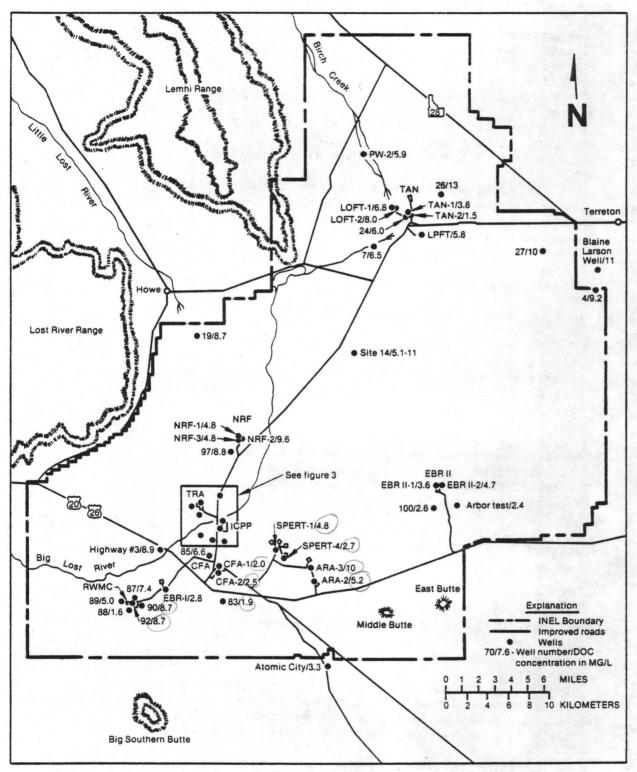
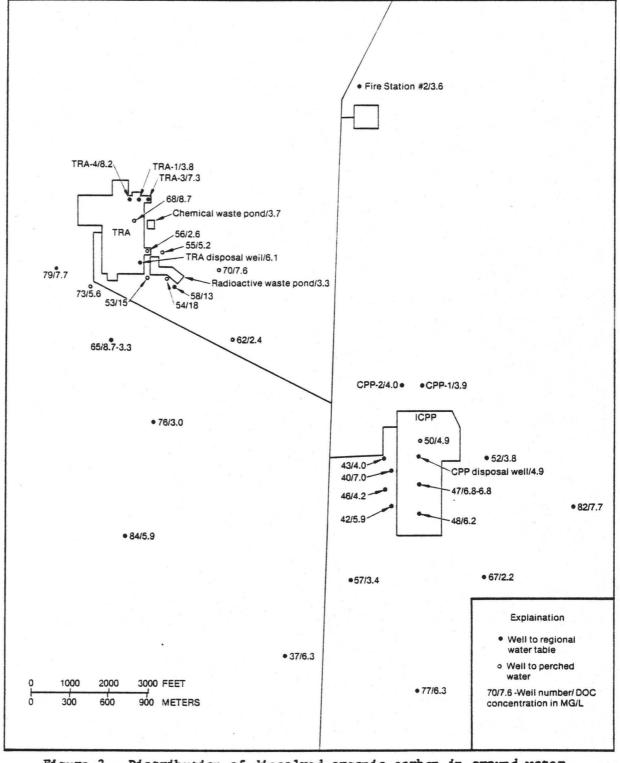


Figure 2. Distribution of dissolved organic carbon in ground water, INEL vicinity, August 1980.



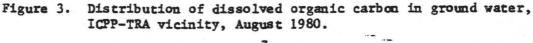


Table 1.--Data table for organic sample collection parameters and dissolved organic carbon results

IP.	Production wel	l in	operation,	pumping	more	than	500	gpm:	NA.	not	available]	1
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Well Number	Station Identification Number	Collection Date - Time	Filtration Date - Time	Temperature at collection (°C)	Method of collection	Sample depth or pump time (min)/ rate (gal/min)	DOC (mg/L)
ARA-2	433106112492101	8/12/80 - 1000	8/12/80 - 1300	14.0	Pump	P	5.2
ARA-3	433156112494401	8/12/80 - 1015	8/12/80 - 1315	15.5	Pump	Р	10
ARBOR test	433508112384801	8/14/80 - 0936	8/14/80 - 1030	13.0	Thief	710 feet	2.4
Atomic City	432638112484101	8/13/80 - 1115	8/13/80 - 1220	15.0	Pump	NA	3.3
Blaine Larsen Well	434715112282701	8/12/80 - 1220	8/13/80 - 0943	NA	Pump	P	11
CFA-1	433204112562001	8/12/80 - 1105	8/12/80 - 1215	11.5	Pump	Р	2
CFA-2	433144112563501	8/12/80 - 1120	8/12/80 - 1225	12.0	Pump	P	2.5
CPP-1	433433112560201	8/15/80 - 1037	8/15/80 - 1055	15.5	Pump	Р	3.9
CPP-2	433432112561001	8/13/80 - 1538	8/14/80 - 0840	16.0	Pump	Р	4
CPP eastside waste	433413112560401	8/18/80 - 0955	8/18/80 - NA	22.0	Surface	Stream	4.9
EBR-1	433051113002601	8/12/80 - 0920	8/12/80 - 0950	NA	Pump	P	2.8
EBR-II-1	433546112391601	8/14/80 - 0830	8/14/80 - 1055	13.0	Pump	Р	3.6
EBR-II-2	433544112391301	8/14/80 - 0842	8/14/80 - 1054	13.5	Pump	Р	4.7
Fire Station #2	433548112562301	8/11/80 - 1522	8/12/80 - 0925	15.0	Pump	P	3.6
Highway #3	433256113002501	8/11/80 - 1350	8/11/80 - 1450	NA	Pump	NA	8.9
LOFT-1	435120112432101	8/12/80 - 1448	8/13/80 - 0935	10.0	Pump	1/1000	6.8
LOFT-2	435119112431801	8/12/80 - 1420	8/13/80 - 0900	10.0	Pump	1/1000	8

Well Number	Station Identification Number	Collection Date - Time	Filtration Date - Time	Temperature at collection (^O C)	Method of collection	Sample depth or pump time (min)/ rate (gal/min)	DOC (mg/L)
LPFT	434946112412401	8/12/80 - 1531	8/13/80 - 0930	13.0	Pump	Р	5.8
NRF-1	433859112545401	8/13/80 - 1055	8/13/80 - 1335	13.5	Pump	Р	4.8
NRF-2	433854112545401	8/13/80 - 1108	8/13/80 - 1337	11.5	Pump	Р	9.6
NRF-3	433858112545501	8/13/80 - 1047	8/13/80 - 1417	15.0	Pump	Р	4-8
PW-2	435419112453101	8/12/80 - 1625	8/12/80 - 2020	13.0	Thief	350 feet	5.9
Site 14 (1)	434334112463101	8/13/80 - 1500	8/13/80 - 1550	16.0	Pump	30/8	5.1
Site 14 (2)	434334112463101	8/13/80 - 1505	8/13/80 - 1552	16.0	Pump	35/8	11
SPERT-1	433252112520301	8/12/80 - 1042	8/12/80 - 1240	11.0	Pump	Р	4.8
SPERT-4	433247112515201	8/12/80 - 1035	8/12/80 - 1255	12.0	Pump	Р	2.7
TAN-1	435056112420001	8/12/80 - 1510	8/13/80 - 0912	11.0	Pump	Р	3.8
TAN-2	435100112420701	8/12/80 - 1517	8/13/80 - 0855	11.0	Pump	Р	1.5
TRA-1	433521112573801	8/13/80 - 1255	8/13/80 - 1415	17.0	Pump	Р	3.8
TRA-3	433521112573501	8/13/80 - 1301	8/13/80 - 1350	18.0	Pump	Р	7.3
TRA-4	433521112574201	8/13/80 - 1247	8/13/80 - 1352	18.0	Pump	Р	8.2
TRA chemical wast	te pond NA	8/18/80 - 1310	8/18/80 - 1445	NA.	Surface	Pond	3.7
TRA cooling tower	r disposal NA	8/18/80 - 1234	8/18/80 - 1446	17.0	Surface	Stream	6.1
TRA radioactive w	waste pond NA	8/18/80 - 1330	8/18/80 - 1400	NA	Surface	Pond	3.3

Table 1.--Data table for organic sample collection parameters and dissolved organic carbon results (continued)

(P, Production well in operation, pumping more than 500 gpm; NA, not available)

Table 1.---Data table for organic sample collection parameters and dissolved organic carbon results (continued)

Well Number	Station Identification Number	Collection Date - Time	Filtration Date - Time	Temperature at collection (°C)	Method of collection	Sample depth or pump time (min)/ rate (gal/min)	DOC (mg/L)
4	434657112282201	8/12/80 - 1200	8/12/80 - 2000	NA	Thief	300 feet	9.2
7	434926112444201	8/12/80 - 1350	8/12/80 - 2030	17.0	Thief	250 feet	6.5
19	434430112575901	8/12/80 - 1750	8/12/80 - 1955	18.0	Thief	295 feet	8.7
24	435053112420801	8/12/80 - 1456	8/12/80 - 1940	14.0	Thief	300 feet	6
26	435215112394201	8/12/80 - 1341	8/12/80 - 1940	18.0	Thief	260 feet	13
27	434854112322101	8/12/80 - 1258	8/12/80 - 2015	18.0	Thief	255 feet	10
37	433326112564801	8/11/80 - 1550	8/12/80 - 0930	12.0	Pump	20/5	6.3
40	433413112561201	8/18/80 - NA	8/18/80 - 1143	21.2	Pump	20/7	7
42	433403112561201	8/13/80 - 1604	8/14/80 - 0857	17.5	Thief	500 feet	5.9
43	433415112561501	8/18/80 - 0945	8/18/80 - 1205	16.1	Pump	20/7	4
46	433407112561501	8/18/80 - 1008	8/18/80 - 1040	14.0	Thief	500 feet	4.2
47(1)	433407112560301	8/18/80 - 1140	8/18/80 - 1246	15.7	Pump	20/7	6.8
47(2)	433407112560301	8/18/80 - 1145	8/18/80 - NA	15.7	Pump	20/7	6.8
48	433401112560301	8/14/80 - 1206	8/14/80 - 1414	14.0	Thief	520 feet	6.2
50	433419112560201	8/14/80 - 1223	8/14/80 - 1412	19.5	Bailer	379 feet	4.9
52	433415112554401	8/13/80 - 1713	8/14/80 - 0837	16.0	Thief	500 feet	3.8
53	433503112573401	8/18/80 - 1438	8/18/80 - 1515	NA	Bailer	78 feet	15

[P, Production well in operation, pumping more than 500 gpm; NA, not available]

Table 1.---Data table for organic sample collection parameters and dissolved organic carbon results (continued)

Well Number	Station Identification Number	Collection Date - Time	Filtration Date - Time	Temperature at collection ([°] C)	Method of collection	Sample depth or pump time (min)/ rate (gal/min)	DOC (mg/L)
54	433503112572801	8/18/80 - 1426	8/18/80 - 1459	NA	Thief	85 feet	18
55	433508112573001	8/18/80 - 1354	8/18/80 - 1515	NA	Thief	78 feet	5.2
56	433509112573501	8/18/80 - 1302	8/18/80 - 1408	12.0	Thief	79 feet	2.6
57	433344112562601	8/18/80 - 0937	8/18/80 - 1041	NA	Thief	550 feet	3.4
58	433500112572501	8/18/80 - 1141	8/18/80 - 1429	12.0	Thief	495 feet	13
62	433446112570601	8/14/80 - 1312	8/14/80 - 1350	13.0	Thief	152 feet	2.4
65(1)	433447112574701	8/18/80 - 1225	8/18/80 - 1244	14.5	Pump	20/7	3.3
65(2)	433447112574701	8/18/80 - 1220	8/18/80 - NA	14.5	Pump	20/7	8.7
67	433344112554101	8/18/80 - 1104	8/18/80 - 1145	13.5	Pump	25/7	2.2
68	433516112573901	8/18/80 - 1211	8/18/80 - 1427	14.0	Thief	120 feet	8.7
70	433504112571201	8/18/80 - 1335	8/18/80 - 1500	10.5	Thief	95 feet	7.6
73	433502112575401	8/14/80 - 1255	8/18/80 - 1427	15.5	Thief	120 feet	5.6
75	433425112573201	8/13/80 - 1732	8/14/80 - 0821	14.0	Thief	500 feet	3.0
	433315112560501	8/13/80 - 1305	8/13/80 - 1435	14.0	Thief	540 feet	6.3
77	433505112581901	8/13/80 - 1801	8/14/80 - 0858	14.0	Thief	550 feet	7.7
79	433401112551001	8/13/80 - 1216	8/13/80 - 1500	14.0	Thief	500 feet	7.7
82		8/13/80 - 1620	8/14/80 - 0815	11.5	Pump	45/7	1.9
83	433023112561501	6/13/00 - 1020	0/14/00 0015				

[P, Production well in operation, pumping more than 500 gpm; NA, not available]

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Table 1.--Data table for organic sample collection parameters and dissolved organic carbon results (continued)

Well umber	Station Identification Number	Collection Date - Time	Filtration Date - Time	Temperature at collection ([°] C)	Method of collection	Sample depth or pump time (min)/ rate (gal/min)	DOC (mg/L)
84	433357112574201	8/13/80 - 1516	8/13/80 - 1845	NA	Bailer	479 feet	5.9
85	433246112571201	8/13/80 - 1431	8/13/80 - 1835	14.0	Thief	570 feet	6.6
87	433013113024301	8/11/80 - 1330	8/11/80 - 1450	14.0	Pump	45/2	7.4
88	432940113030201	8/11/80 - 1315	8/11/80 - 1430	13.0	Pump	20/5	1.6
89	433005113032901	8/11/80 - 1330	8/11/80 - 1430	13.0	Pump	20/5	5.0
90	432954113020501	8/11/80 - 1245	8/11/80 - 1505	13.0	Pump	20/4	8.7
92	433000113025301	8/14/80 - 1120	8/14/80 - 1352	11.5	Baller	209 feet	8.7
97	433807112551501	8/13/80 - 1131	8/13/80 - 1437	13.0	Thief	440 feet	8.8
100	433503112400701	8/14/80 - 0906	8/14/80 - 1032	13.0	Thief	710 feet	2.6

[P, Production well in operation, pumping more than 500 gpm; NA, not available]

It should be noted that some of the observation wells sampled for various types of organic compounds are perched water wells (fig. 3) and do not reflect the organic concentrations in the Snake River Plain aquifer (Robertson, 1977). Perched water wells 50, 53, 54, 55, 56, 68, 70, and 92 were sampled to study the leaching tendencies of surface disposed organic compounds at various INEL facilities.

FIELD SAMPLING TECHNIQUES

Special sampling techniques were employed in the organic sampling program. Special collection methods are required to assure that the sample analyzed in the laboratory is in the same organic state as it was when originally collected. Samples were collected in special detergent-washed, acid-rinsed, sterilized bottles. Immediately after collection, all samples were placed on ice and kept at $1-4^{\circ}C$ until time of analysis.

Wells equipped with pumps were pumped until the water was representative of the water in the aquifer. Then a water sample was collected. For wells without pumps, a thief sampler (fig. 4) was used to collect samples from the well at a given depth. Four water samples from perchedwater wells were collected using a bailer (fig. 4). The bailer and thief sampler were washed with detergent, rinsed with distilled water, washed with acetone, and again rinsed with distilled water before sampling began each day. Between samples, the bailer and thief sampler were rinsed with distilled water.

Dissolved Organic Carbon

Samples for DOC analyses were collected between August 11 and 18 by either pump, thief sampler, or bailer. Immediately after collection, the sample bottles were put on ice and transported to the Radiological and Environmental Sciences Laboratory (RESL) at the INEL where they were filtered and refrigerated prior to their shipment to the NWQL in Denver, Colorado.

The preservation of samples for DOC analyses is accomplished by the combined effects of filtration, chilling, and contact with silver (Ag). Most micro-organisms will not pass through a 0.45-micron Ag filter. During filtration, approximately $l\mu g$ of Ag is dissolved from the filter per milliliter of water filtered. Silver in solution or as a colloid exhibits bactericidal properties (Chambers and others, 1962). In addition to the filtering and bactericidal effects of the Ag filter, the samples are sealed in a glass water-tight container and kept chilled on ice until analysis. Over a four week period of testing on a variety of samples (at the NWQL), no losses of DOC or contamination have been observed when using this method.

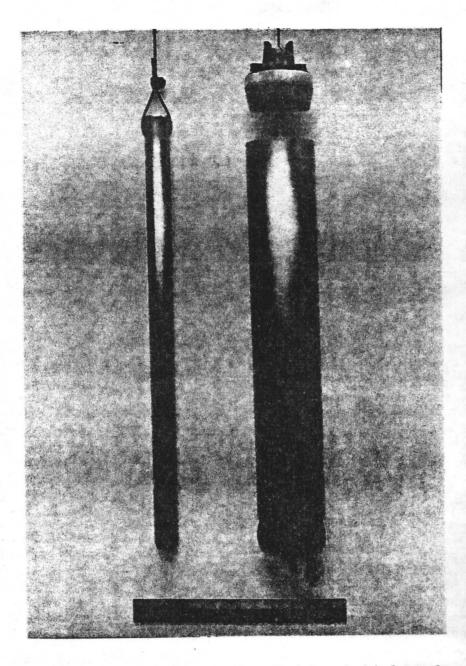


Figure 4. Photograph of bailer (left) and thief sampler (right) used for collecting water samples from wells without pump.

Insecticides, Herbicides, Semivolatiles

Insecticide, herbicide, and semivolatile samples were collected on August 18, 1980 by thief and pump sampling methods. One liter of raw unfiltered water was collected for each analysis. Immediately after collection, all samples were chilled and held at between $1-4^{\circ}C$ until analyses were done at the NWQL.

Volatiles

Samples for volatile organics were collected on August 18, 1980 by thief, bailer, or pump methods.

The Volatile Class of EPA priority pollutants includes those compounds that can be sparged from a water sample at room temperature. The fact that these low-boiling compounds are quite easily removed from the sample necessitates special care in the collection, shipping and storage of volatile samples. Bottles used for volatile sample collection are 250 milliliter, screw cap, septum-bottles, made of non-actinic (brown) glass. The septa are teflon coated.

Samples are collected in quadruplicate because once the water is subsampled during analysis, headspace is created in the bottle. This precludes further subsampling as the very volatile organic compounds will evaporate into the headspace and will no longer be present in the water sample at the original concentration.

During actual sample collection, the sample is collected in such a manner as to insure that no air bubbles pass through the bottle while it is being filled. The bottle is filled to overflowing and the cap is immediately replaced. Then the bottle is turned upside down to check for air bubbles. If bubbles are present, the bottle must be emptied and another sample taken. This process is repeated until no air bubbles are present in the sample. The sample is then kept chilled to $1-4^{\circ}C$ until it is analyzed.

For best results, analysis for volatile organics must be made within 36 hours of sample collection. For this reason, after sample collection was completed on August 18, 1980, the organic samples were removed from the storage refrigerator at the INEL, packed in ice, and transported to the NWQL in Denver where volatile organic analysis began on August 19, 1980.

Analytical Methods

Dissolved organic carbon was determined by the sealed ampoule, wetoxidation procedure described by Malcolm and others, 1973. DOC results were reported in milligrams per liter + 1 mg/L. Insecticide and PCB analyses were performed by various specific gas chromatographic procedures (Goerlitz and Brown, 1973) and covered the following compounds:

Aldrin - Total (Water) Chlordane - Total (Water) DDD - Total (Water) DDE - Total (Water) DDT - Total (Water) Dieldrin - Total (Water) Endosulfan I - Total (Water) Heptachlor Epoxide - Total (Water) Heptachlor - Total (Water) Lindane - Total (Water) Methoxychlor - Total (Water) Mirex - Total (Water) PCB - Total (Water) PCN - Total (Water) Perthane - Total Silvex - Total (Water) 2,4-D - Total (Water) 2,4-DP - Total (Water) 2,4,5-T - Total (Water)

Insecticide results were reported in μ g/L x 10⁻² with the exception of the results for chlordane, PCB, and Toxaphene which were reported in μ g/L x 10⁻¹.

Volatile analyses were performed by sparging a five-milliliter aliquot of sample for ten minutes with helium, and trapping volatile organic compounds on a column of Tenax GC. Trapped compounds were thermally desorbed and flushed onto the head of a Carbowax 20M column for GC-MS analysis as described by Pereira and Hughes, 1980.

Semivolatile compounds from one-liter samples were solvent extracted with methylene chloride first at pH 12 to isolate base-neutral semivolatile compounds, followed by a second extraction at pH-2 to isolate acid compounds. Methyl esters of organic acids were prepared by derivatization with diazomethane. The GC-MS analyses of each fraction were performed on a 2 mm ID x 6 feet glass column packed with a 3 percent SP 2100. This column was temperature-programmed with 50° to 260° C at 6° /minute after a 5 minute hold at 50° C and was held at 260° C for 10 minutes at the end of the program. Mass spectral scans were run from 60 to 450 amu. Water and reagent blanks were run, and semi-quantitative results were calculated by comparison of unknown chromatographic peaks with a $10 \pm g/L$ internal standard.

Mass spectral identifications of volatile and semivolatile compounds were performed by computer comparison with the National Bureau of Standards mass spectral data base. Only those identifications which have met vigorous comparison tests have been reported as present in the sample. Briefly, the criteria for identification are Fit \geq 800 and Purity \geq 400. Purity measures the resemblance of the unknown spectrum to a specified library entry. A Rurity value of 1000 signifies identical mass spectra. Fit is also a measure of spectral resemblance; however, peaks present in the unknown but absent in the library spectrum are not considered in the Fit algorithm. A Fit of 1000 would indicate that the unknown spectrum contains the library spectrum but possibly as an unresolved mixture. Those compounds which could not be identified with a high degree of certainty or which were present in the reagent blank have not been reported.

RESULTS AND DISCUSSION

In considering the results of any organic reconnaissance programs, it is important to remember several facts (U.S. Environmental Protection Agency, 1976):

- Organic compounds, both synthetic and natural, are present in all drinking waters to some extent. In addition, some halogenated organic compounds are produced during the disinfection of drinking water with chlorine.
- The major portion of organic compounds in most waters is of natural origin.
- Most of the specific organic compounds in drinking water have not been identified, and analysis for many of them is very difficult.
- Most of the identified organic compounds in drinking water have not been bioassayed.
- 5. The effect on humans of long-term ingestion of very low levels (ng/L to mg/L) of organic chemicals in drinking water is not known, and the portion of human exposure from drinking water versus the total exposure from all sources (food, air) is seldom known. However, the drinking water portion is usually considered to be small.

Dissolved Organic Carbon

The DOC results were more reflective of the sampling procedure (thief vs. pump sampling) than of the presence of organic contamination This relationship is shown in table 2.

	Pump sampled,	Thief sampled,
Well designation	dissolved organic carbon (milligrams per liter)	dissolved organic carbon (milligrams per liter)
Atomic City	3.3	
Blain Larson	11	
Fire Station #2	3.6	
Highway #3	8.9	
P&W #2		5.9
Site 14	5.1, 11	
4		9.2
19		8.7
26		13
27		10
Mean	7.1	9.3

Table 2.--Dependence of dissolved organic carbon oncentration in background wells on method of well sampling

Thief or bailer sampling does not flush the well, and relatively high DOC values can result from stagnant water and organic contamination remaining from drilling fluids and well casing. Pump sampling, on the other hand, does exchange the water in the well bore for new water from the aquifer, and therefore gives a more representative sample of actual DOC concentrations in the ground water.

In order to indicate any significant organic contamination, the DOC value for any given well should exceed twice the average DOC background value for the particular sampling method used. The average DOC value for background wells sampled with a thief sampler was 9.3 mg/L and for background wells sampled with a pump was 7.1 mg/L (table 2). Therefore, a well sampled with a thief sampler should have a DOC value of about 20 mg/L and a pumped well should have a DOC value of about 15 mg/L before any significant amount of organic contamination would be indicated.

Generally, the INEL production wells that are pumped regularly had the lowest DOC concentrations (table 1). The other pumped wells also had low concentrations. Wells sampled with a thief sampler had higher concentrations. The highest DOC concentrations were measured in the perched water wells around the TRA chemical and radioactive-waste ponds. However, none of the DOC samples taken from the INEL indicated any high level organic contamination of ground water.

Insecticides and Herbicides

No insecticides or herbicides were detected in any samples except for observation well 40, where dichlorodiphenyltrichloroethane (DDT) was found to be present at 10 parts per trillion $(0.01 \ \mu g/L)$. This low level (coupled with the absence of detectable insecticides and herbicides in the other samples) does not indicate significant contamination in INEL ground water by insecticides and herbicides.

Volatiles and Semivolatiles

Of the eighteen water samples sent to the NWQL, seventeen were analyzed for extractable and volatile organic constituents. The eighteenth sample was from the TRA radioactive waste disposal pond and was not analyzed due to its' radioactivity. The results are presented in table 3.

The compounds identified as being in the analyzed sample can be broken down into four categories based upon the judgment of the analyst.

- Occurs in Blank These compounds are present in the blank (distilled water) sample run prior to analysis of the other samples, and therefore probably do not occur in the original sample.
- 2. <u>Unlikely to Occur</u> These compounds probably result from variable sample contaminants which may or may not occur during the

I. ICPP Waste Effluent

A. Base-neutral extractable fraction

	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
	Ethylbenzene	Chloroform	Methyld1hydrofuran	None of compounds reported
		Trichloroethene	Trichloroethene	
			Dihydromethylpyran	
	•		Trimethyldodecatrienol	
в.	Acid extractable fract	tion		
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 $\mu g/L$)
	Ethylbenzene	Trichloroethene	Methylpentanoic acid	None of compounds reported
	Heneicosanoic acid	Heptane	Pentanoic acid	
		Methylhexane	Benzoic acid	
		Dimethylhexane	Methylhexanoic acid isomers	
		Ethylmethylhexane	Methylnonanoic acid isomers	
		Xylene isomers	Benzenedioic acid isomers	
		Tetrachloroethane	Methyldecanoic acid isomers	
		Dichloropentane	Tridecynoic acid	
		Tetramethylpentane	Tetradecynoic acid	
		Bromohexane isomers	Hexadecynoic acid	
		Methyloctane	Triacontanoic acid	
		Chlorooctane	Undecanoic acid	
		Diethylphthalate	Dimethylhexanone nonanoic ac	14
		Oxybis-octane	Dimetal inclusione nonunoite ac	
		Octanone		
		Hexanal		
		Methylhexanone		
		Nitromethylbenzene		
c.	Volatiles	MILIOWelnyIbenzene		
0.	Volaciies			
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 $\mu g/L$)
			Cyclohexane	None of compounds reported
			Dimethyloxetane	

II. Well-70

0

A. Base-neutral extractable fraction

ccurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
etrachloroethene thylbenzene			None of compounds reported
cid extractable frac	tion		
ccurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 µg/L)
eneicosanoic acid	Trichloroethene Methylhexane Xylene isomers Dichloropentane Propylbenzene Methoxypropanediol Bromohexane isomers Dimethylheptane isomers	Ethanedioic acid Methylpentanoic acid isomers Benzoic acid Decanoic acid Undecanoic acid Methyldecanoic acid isomers Tridecanoic acid Tridecynoic acid	None of compounds reported
olatiles	Diethylphthalate Oxybis-octane	Hexadecenoic acid Tetradececynoic acid Octadecenoic acid Triacontanoic acid Tetradecanoic acid	
ccurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 µg/L)
	Methylene chloride	Propanone Dimethyloxetane	None of compounds reported
	etrachloroethene chylbenzene cid extractable frac cours in Blank eneicosanoic acid	etrachloroethene thylbenzene cid extractable fraction cours in Blank Unlikely Occurrence eneicosanoic acid Trichloroethene Methylhexane Xylene isomers Dichloropentane Propylbenzene Methoxypropanediol Bromohexane isomers Dimethylheptane isomers Diethylphthalate Oxybis-octane	etrachloroethene thylbenzene cid extractable fraction ccurs in Blank Unlikely Occurrence Tentatively Identified eneicosanoic acid Trichloroethene Ethanedioic acid isomers Xylene isomers Benzoic acid isomers Xylene isomers Benzoic acid Dichloropentane Decanoic acid isomers Bromohexane isomers Tridecanoic acid isomers Bromohexane isomers Tridecanoic acid Diethylphthalate Hexadecenoic acid Oxybis-octane Tetradecerynoic acid Triacontanoic acid Tetradecenoic acid

Methylbutanone

. III. Well-67

21

A. Base-neutral extractable fraction

	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 µg/L)
		Diethylphthalate Dipropylphathalate	Tridecadiene Undecanoic acid methyl ester Tetramethylhexadecatirienol	None of compounds reported
B	Acid extractable fractio	n		
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 $_{\rm Hg}/L$)
	Acetic acid ethyl ester Heneicosanoic acid	Trichloroethene Heptane Trichloroethane nonanediol Bromohexane isomers Ethylmethylhexane Diisooctylphthalate Dimethylhexanal	Benzoic acid Methylhexanoic acid Methylnonanoic acid Decanoic acid Undecanoic acid Tetradecynoic acid Tetradecanoic acid Hexadecenoic acid Triacontanoic acid Cyclopentaneundecanoic acid	None of compounds reported
С	Volatiles			
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 pg/L)
		Methylene chloride		None of compounds reported

4

IV. Well-65

A. Base-neutral extractable fraction

	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
	Ethylbenzene	Diisooctylphthalate Trimethylpentene	Hexylpentyl ether Trimethyltetracontane Trimethyldodecatrienol Hexatriaceontane	None of compounds reported
B.	Acid extractable fractio	n		
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
	Acetic acid ethyl ester Heneicosanoic acid	Methylhexane Heptane Methylcyclohexane Tetrachloroethane Bromohexane isomers Dimethylhexane Oxybis-octane Diisooctylphthalate Methylpentanol Decenone	Methylnonanoic acid Decanoic acid Tetradecanoic acid Methyltetradecanoic acid Hexadecynoic acid Pentadecynoic acid Cyclopentaneundecanoic acid	None of compounds reported
c.	Volatiles			
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
		Methylene chloride	Trimethylpentane Dimethyldisulfide	Trimethylpentane Dimethyldisulfide

22

V. Well-58

A. Base-neutral extractable fraction

Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)	
		Methylnonanoic acid methyl ester Methylethyldioxetane	None of compounds reported	

B. Acid extractable fraction

	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ_B/L)
	Acetic acid ethyl ester	Cyclohexane Trichloroethene Heptane Methylcyclohexane Trichloroethane Xylene isomers Bromohexane isomers Methyltetrahydrofuranol Methylpentanol Hexanal Deceñone	Benzoic acid Methylhexanoic acid isomers Nonanoic acid Methylnonanoic acid isomers Methyldecanoic acid isomers Undecanoic acid Tridecynoic acid Octadecadiynoic acid Dimethyloctahydrophenanthren carboxylic acid Cyclopentaneundecanoic acid Octadecadecynoic acid	None of compounds reported
c.	Volatiles			
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
		Methylene chloride	Cyclohexane Dimethyloxetane Methylpropane	None of compounds reported

VI. Well-57

A. Base-neutral extractable fraction

Occurs in Blank

Unlikely Occurrence Diethylphthalate

Tentatively Identified

Significant Concentration (>10 µg/L)

Decylhydroxylamine Dimethylphthalate

None of compounds reported

Pheny1methy1sydnone Propylheptanol Methylnonanoic acid methyl ester

Acid extractable fraction в.

Occurs in Blank

Tentatively Identified Unlikely Occurrence

Significant Concentration (>10 µg/L)

Acetic acid ethyl ester Trichloroethene Heneicosanoic acid

Heptane Methylcyclohexane Dimethylhexane Dimethyloxirane Xylene isomers Tetrachloroethane Dichloropentane Dimethylnonane Bromohexane isomers Diethylphthalate Oxybis-octane Diisooctylphthalate Heptenone Benzeneacetaldehyde Octanedione Tetramethylhexatrienol Nitromethylbenzene Nitropropylbenzene

Methyl pentanoic acid isomers None of compounds reported Benzoic acid Octanoic acid Cyclopentaneundecanoic acid Decanoic acid Methyldecanoic acid Oxononanoic acid Benzene dicarboxylic acid isomers Triacontanoic acid Undecanoic acid Tridecynoic acid Tetradecanoic acid Pentadecynoic acid Methyltetradecanoic acid isomers Hexadecenoic acid isomers Octadecadiynoic acid Dimethyloctahydrophenanthrene Carboxylic acid

C. Volatiles

Unlikely Occurrence Occurs in Blank

Tentatively Identified

Significant Concentration (>10 µg/L)

None of compounds reported

Methylene chloride

Cyclohexane Dimethylpentane Methylethylcyclopropane

VII. Well-56

A. Base-neutral extractable fraction

Acetic acid ethyl ester Trichloroethene

Occurs in Blank

Occurs in Blank

Heneicosanoic acid

Ethylbenzene

Unlikely Occurrence

Tentatively Identified

Significant Concentration (>10 µg/L)

Diethylphthalate Butyl 2-methylpropylphthalate Propionamide I Propylethylhexanol Hexylpentyl ether Hexatriacontane Trimethyltetracontane Tetratetracontane Tetradecynoic acid methyl ester

None of compounds reported

B. Acid extractable fraction

Acetic acid pentyl ester

Bromocyclohexane isomers

Unlikely Occurrence

Methylcyclohexane

Methylhexene

Dimethylheptane

Diethylthalate

Ethynyloxirane

Dimethylhexanal Bezaldehyde

Diisooctylphthalate

Oxybis-octane

Tentatively Identified

Cyclopentaneundecanoic acid

Methyldecanoic acid isomers

Benzenedicarboxylic acid isomers

Methylhexanoic acid

Benzoic acid

Heptanoic acid

Undecanoic acid

Docosenoic acid

Tetradecanoic acid

Heptadecatriynoic acid

Significant Concentration (>10 µg/L)

None of compounds reported

C. Volatiles

Occurs in Blank

Unlikely Occurrence

Tentatively Identified

Significant Concentration (>10 µg/L)

Methylene chloride

Cyclohexane Methylethylcyclopropane None of compounds reported

25

VIII. Well-55

A. Base-neutral extractable fraction

Occurs in Blank

Unlikely Occurrence

Tentatively Identified

Significant Concentration (>10 µg/L)

Trichloroethene Dimethylpentene

Unlikely Occurrence

Heptanol Methylethyldioxolane Trimethylcyclopentane Trimethyldodecatrienol

Tentatively Identified

None of compounds reported

B. Acid extractable fraction

Occurs in Blank

Acetic acid ethyl ester Heneicosanoic acid

Trichloroethene Cyclohexane Bromohexane isomers Bromocyclohexane Tetrahydro-dimethylpyranone Bezaldehyde

Benzoic acid Hexanoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers Nonanoic acid isomers Tridecanoic acid Hexadecanoic acid Tetradecynoic acid Cyclopentaneundecanoic acid

Significant Concentration (>10 µg/L)

None of compounds reported

C. Volatiles - none detected

IX. Well-54

27

A. Base-neutral extractable fraction

Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
Ethylbenzene	Ethylmethylcyclopentane	Nonanal	None of compounds reported
	Tetrachloroethane	Dimethylethoxyl methylbenzene	
	Xylene isomers	Hexylpentyl ether	
	Dipropylphthalate	Hexatriacontane	
	Trimethylhexane	High-moleculor-weight hydroca	rbons
Acid extractable	fraction		
Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 µg/L)
Heneicosanoic acid	Cyclohexane	Hexanoic acid	None of compounds reported
	Methylcyclohexane	Methylnonanoic acid	
	Chloroform	Decanaoic acid	
	Bromohexane isomers	Undecanoic acid	
	Oxybis-octane	Tetradecanoic acid	
		Tridecynoic acid	
C. Volatiles			
Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
	Methylene chloride	Cyclohexane Methylethylcyclopropane	None of compounds reported

X. Well-53

28

A. Base-neutral extractable fraction

	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
		Diethylphthalate Diisooctylphthalate Butyl 2-methylpropyl- phthalate Xylene isomers	Trimethyldodecatrienol High-moleculor-weight hydrocarbons	None of compounds reported
в.	Acid extractable fra	ction		
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
		Dimethylpentane Methylcyclohexane Tetrachloroethane	Methylnonanoic acid isomers Undecanoic acid isomers Methyldecanoic acid	None of compounds reported
c.	Volatiles			
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
			Methylethylcyclopropane Dimethyloxetane	None of compounds reported

\$

XI. Well-47

A. Base-neutral extractable fraction

Occurs in Blank Ethylbenzene

Xylene isomers Butyl 2-methylpropylphthalate

Unlikely Occurrence

Tentatively Identified Methylphenylhydrazine

Hexylpentyl ether

Significant Concentration (>10 µg/L)

None of compounds reported

Methoxyhexene Nonanediol Tridecadiene Dimethyldodecatrienol Methylpentanol Methylnonanoic acid methylester isomers Cyctopentaneundecanoic acid Methylester

B. Acid extractable fraction

Unlikely Occurrence Tentatively Identified Significant Concentration (>10 μ g/L) Occurs in Blank Ethylbenzene Trichloroethene Benzoic acid None of compounds reported Acetic acid ethyl ester Heptane Benzene acetic acid Cyclohexane Methylnonanoic acid isomers Dichloromethylpropane Methyldecanoic acid isomers Tridecynoic acid Xylene isomers Tetrachloroethane Tetradecanoic acid Dichloropentane Cyclopentaneundecanoic acid Tetramethylhexane Bromohexane isomers Dimethylheptane Methyloctane **Ethylbenzene** Chlorooctane Decenone Dimethylheptanol Benzaldehyde Trimethylpentenol Octadeceno1 C. Volatiles Occurs in Blank Unlikely Occurrence Tentatively Identified Significant Concentration (>10 µg/L) None of compounds reported Methylene chloride

XII. Well-43

A. Base-neutral extractable fraction

	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
	Ethylbenzene	Diethylphthalate	Methoxyhexene	Methylpentadecanoic acid methyl ester
	nt ny i benzene	Diisooctylphthalate	Nonaned 101	Hexadecenoic acid methyl ester
		Trimethylhexane	Methylnonane	
		Xylene isomers	Nonanoic acid methyl ester	
		Ny route results	Dodecanone	
			Methylnonanoic acid methyl es	ster
			Phenylphenyl hydrazine	•
			Dodecadiene	
			Cyclopentaneundecanoic acid m	nethyl ester
			Trimethyldodecatrienol	
			Methylpentadecanoic acid meth	nyl ester
			Hexadecenoic acid methyl este	
в.			Tentatively Identified	Significant Concentration (>10 $\mu g/L$)
	Occurs in Blank	Unlikely Occurrence	Tentativery Identified	DIBITICANT CONCONCILIANT FREEZE
		Methylcyclohexane Trimethylpentenol	Methylnonanoic acid isomers Undecanoic acid	None of compounds reported
c.	Volatiles			
	Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
		Methylene chloride		None of compounds reported

XIII. TRA disposal well

. 11

A. Base-neutral extractable fraction

Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 µg/L)
Ethylbenzene	Diethylphthalate	Trimethyldodecatrienol	Trimethyldodecatrienol
	Trichloroethene	Phenyl propanone	
	Xylene isomer	Methylethylcyclopentane isomer	9
	Dimethylpentene	Dimethylethyl benzene	
	Ethylmethylpentane	Methylphenylethanone	
		Tetramethylbenzene	
		Dihydro-methyl-1(N)indene	
		Naphthalene	
		Dimethyl-methylethylbenzene	
		Methylene-bis-pyrrolidine	
		Hexylpentyl ether	
		Butylbenzene	
Acid extractable fracti	on		
Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 µg/L)
Occurs in Blank	Unlikely Occurrence		
<u>Occurs in Blank</u> Acetic acid ethyl ester	Unlikely Occurrence Methylcyclohexane	Benzoic acid	Significant Concentration (>10 µg/1.) None of compounds reported
Occurs in Blank	Unlikely Occurrence Methylcyclohexane Dichloropentane	Benzoic acid Methylhexanoic acid isomers	
<u>Occurs in Blank</u> Acetic acid ethyl ester	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers	
<u>Occurs in Blank</u> Acetic acid ethyl ester	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers Dimethylheptane	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers	
<u>Occurs in Blank</u> Acetic acid ethyl ester	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers Dimethylheptane Methylpentanol	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers Tętradecanoic acid	
<u>Occurs in Blank</u> Acetic acid ethyl ester	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers Dimethylheptane Methylpentanol Hexenone	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers	
<u>Occurs in Blank</u> Acetic acid ethyl ester	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers Dimethylheptane Methylpentanol Hexenone Methylbutynol	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers Tętradecanoic acid	
<u>Occurs in Blank</u> Acetic acid ethyl ester	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers Dimethylheptane Methylpentanol Hexenone	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers Tętradecanoic acid	Significant Concentration (>10 µg/1.) None of compounds reported
<u>Occurs in Blank</u> Acetic acid ethyl ester Ethylbenzene	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers Dimethylheptane Methylpentanol Hexenone Methylbutynol	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers Tętradecanoic acid	
<u>Occurs in Blank</u> Acetic acid ethyl ester Ethylbenzene	Unlikely Occurrence Methylcyclohexane Dichloropentane Bromohexane isomers Dimethylheptane Methylpentanol Hexenone Methylbutynol	Benzoic acid Methylhexanoic acid isomers Methylnonanoic acid isomers Methyldecanoic acid isomers Tętradecanoic acid	

XIV. We11-40

A. Base-neutral extractable fraction

Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
	Xylene	Dihydromethylpyrone	None of compounds reported
	Diethylphthalate	Propenylcyclopentane	
	Di(methylethyl)-dimethyl-	Trimethylcyclobutanone	
	aziridine	Ethylpropylhexanol	
	Tetrachloroethane	Hexylpentylether	
	Dimethylhexene	Hexatriacontane	
	Propylcyclopentane	Trimethyldodecatrienol	
	Trimethylhexane	Methylnonanoic acid methyl ester	
		Cyclopentaneundecanoic acid methyl ester	
		Tetradecynoic acid methyl ester	
B. Acid extractable fra	iction		
Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
			None of compounds reported
	DifeooctyInhthalate	Methylnonanoic acid	None of compounds reported
	Diisooctylphthalate	Methylnonanoic acid Methyldecanoic acid	None of compounds reported
	Diethylphthalate	Methyldecanoic acid	wone of compounds reported
	Diethylphthalate Octadecenal	Methyldecanoic acid Undecanoic acid	wone of compounds reported
	Diethylphthalate Octadecenal Dimethyloctanol	Methyldecanoic acid Undecanoic acid Octadecenol	
	Diethylphthalate Octadecenal Dimethyloctanol Tetramethylhexadecatrienol	Methyldecanoic acid Undecanoic acid Octadecenol Tetradecanoic acid	
	Diethylphthalate Octadecenal Dimethyloctanol	Methyldecanoic acid Undecanoic acid Octadecenol	wone of compounds reported

C. Volatiles

Occurs in Blank

Unlikely Occurrence

Tentatively Identified

Significant Concentration (>10 µg/L)

None of compounds reported

Methylene chloride

32

XV. TRA chemical waste pond

H

A. Base-neutral extractable fraction

Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 µg/l.)
	Xylene isomers	Ethylmethylcyclohexane	Ethylmethylcyclohexane
	Diisooctylphthalate	Heptane	
	Trichloroethene	Methyldihydropyran	
	Diethylphthalate	Dimethylcyclohexane	
	Methylbutanoic acid	Ethylmethylcyclohexane	
		Propylcyclohexane	
		Bicyclotrimethyloctanone	
		(Dimethylethoxy) methyl benze	ne
Acid extractable frac	tion		
Occurs in Blank	Unlikely Occurrence	Tentatively Identified	Significant Concentration (>10 μ g/L)
<u>Occurs in Blank</u> Heneicosanoic acid	Unlikely Occurrence Methylcyclohexane	Tentatively Identified Methyldecanoic acid isomers	Significant Concentration (>10 µg/L) None of compounds reported
	Methylcyclohexane	Methyldecanoic acid isomers	
	Methylcyclohexane Oxybls-octane	Methyldecanoic acid isomers Undecanoic acid isomers	
	Methylcyclohexane Oxybis-octane Diisooctylphthalate	Methyldecanoic acid isomers Undecanoic acid isomers Tetradecanoic acid isomers	
Heneicosanoic acid	Methylcyclohexane Oxybis-octane Diisooctylphthalate	Methyldecanoic acid isomers Undecanoic acid isomers Tetradecanoic acid isomers	

XVI. Well-46

34

A. Base-neutral extractable fraction

Occurs in Blank

Unlikely Occurrence

Tentatively Identified

Significant Concentration (>10 µg/L)

Xylene Diethylphthalate Diisooctylphthalate

.

N-(hydroxy methyl) propenamide Methylphenyl-dloxolane None of compounds reported

B. Acid extractable fraction - lost prior to analysis

C. Volatiles

Occurs in Blank

Unlikely Occurrence

Tentatively Identified

Significant Concentration (>10 µg/L)

Aziridine

Ethylmethylcyclopropane

None of compounds reported

XVII. Well-68

A. Base-neutral extractable fraction

Occurs in Blank

Ethylbenzene

Trichloroethene Xylene isomer Diethylphthalate Butyl 2-methylpropylphthalate

Unlikely Occurrence

Tentatively Identified

Significant Concentration (>10 µg/L)

None of compounds reported

Methylpentenol Dimethylpentene Ethylmethylcyclopentane Ethylcyclopentane Dimethylhexane Trimethylcyclopentane isomers Butenediamide Ethylcyclohexane Propylcyclopentane Ethyldodecene Trimethyldodecatrienol

B. Acid extractable fraction

Occurs in Blank Unlikely Occurrence Tentatively Identified Significant Concentration (>10 µg/L) Ethylbenzene Trichloroethane Hydroxypropanoic acid None of compounds reported Heneicosanoic acid Bromocyclopentane Benzoic acid Bromohexane Methylhexanoic acid isomers Dimethylheptane Methylnonanoic acid isomers Diisooctylphthalate Decanoic acid Dimethylhexanal Methyldecanoic acid Benzaldehyde Undecanoic acid Nitromethylbenzene Tetradecanoic acid Tetramethy1-hexadecatrienol Tetradecynoic acid Hexadecanoic acid Pentadecynoic acid C. Volatiles Occurs in Blank Unlikely Occurrence Tentatively Identified Significant Concentration (>10 $\mu g/L$) Methylene chloride Chloroform Chloroform Ethylmethylcyclopropane

analytical procedure. A "clean lab" capability is required to avoid these contaminants and the NWQL does not have this capability.

- 3. <u>Tentatively Identified</u> These compounds were probably in the water samples when they were received at the NWQL. Keep in mind that these analyses are unconfirmed by standards and their source may be due to (a) organics introduced by well drilling, (b) organic compounds introduced by sampling, (c) organic compounds introduced by well casing, (d) organic compounds which naturally occur in the area ground water, and (e) organic contaminants introduced into the ground water through present waste disposal practices.
- 4. Significant Concentration All compounds which occur at levels near or above the internal standard (>10 μ g/L) were grouped at this level. The analysis was qualitative in nature and the specific concentration of the compounds in this category cannot be estimated except to say that they were near or above the internal standard.

Only those compounds from the "Tentatively Identified" categories which were also found in "Significant Concentrations" will be discussed here. The other compounds are either unlikely to occur or else they occur in such low concentrations as to be negligible.

<u>Well 65</u> - Trimethylpentane and dimethyldisulfide were tentatively identified in the volatiles fraction. These compounds may have migrated from the TRA disposal well or may result, especially dimethyldisulfide, from microbiological transformations of natural organic substrates. The first compound is used as a solvent and in making fuels.

<u>Well 43</u> - Methylpentadecanoic acid methyl ester and hexadecenoic acid methyl ester were tentatively identified in the base-neutral extractable fraction. These compounds probably occur naturally in ground ater.

TRA disposal well - Trimethyldodecatrienol was tentatively identified in the base-neutral fraction. This is an alcohol, possibly used as detergent somewhere in the cooling tower system.

TRA chemical waste pond - Ethylmethylcyclohexane was tentatively identified in the base-neutral fraction. This compound may originate from solvents, greases, or fuels used at the TRA.

<u>Well 68</u> - Chloroform has been tentatively identified in the volatiles fraction. This is a perched water well inside the TRA compound where chloroform may occur from chlorinated water leaching into the well r it may be a result of the analytical process.

None of these compounds reported at significant concentrations are reported to be hazardous except chloroform, and chloroform was an unconfirmed analysis which occurred in only one sample. The cost for the 1980 Organic Reconnaissance Study at the INEL, including planning, sample collection, sample analysis, laboratory time, manpower, equipment, sample transportation, result interpretation, and initial report publication, has amounted to about \$20,000.

SUMMARY AND CONCLUSION

Results of the 1980 reconnaissance survey of organic solutes in ground water at the INEL have indicated no significant areas of organic contamination of ground water. Because of this, the second phase of this study was canceled. It is suggested that general organic reconnaissance studies be conducted at the INEL in the future on a periodic basis.

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