WATER-QUALITY VARIABILITY IN FOUR RESERVOIRS IN PHILLIPS AND VALLEY COUNTIES, MONTANA, MAY THROUGH AUGUST 1981 By Rodger F. Ferreira and John H. Lambing

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CONVERSION FACTORS

The following factors can be used to convert from the International System of Units (SI) in this report to the equivalent inch-pound units.

Multiply SI unit	By	To obtain inch-pound unit
hectare (ha)	2.471	acre
kilometer	0.6214	mile
liter (L)	33.82	ounce, fluid (oz)
meter (m)	3.281	foot
milliliter (mL)	0.0338	ounce (fluid)
millimeter (mm)	0.0394	inch
square kilometer	0.3861	square mile
square meter (m^2)	10.76	square foot

Temperatures in degrees Celsius (°C) can be converted to degrees Fahrenheit (°F) by the formula:

F = 9/5 (°C) + 32

WATER-QUALITY VARIABILITY IN FOUR RESERVOIRS

IN PHILLIPS AND VALLEY COUNTIES, MONTANA,

MAY THROUGH AUGUST 1981

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Rodger F. Ferreira and John H. Lambing

ABSTRACT

Four reservoirs in Phillips and Valley Counties were studied from May to August 1981 to describe: (1) the variation in water quality that occurs from late spring (May) to late summer (August) and during 24 hours in late summer (August), and (2) possible causes for water-quality variation during these periods. The reservoirs represent considerable ranges of water quality and reservoir age that exist for many similar reservoirs in eastern Montana. Although there are similarities in water quality of reservoirs from the same regional mineralogy, differences in water quality caused by local mineralogy and land use mask differences caused by reservoir age.

All the reservoirs had distinct thermal gradients but lacked true stratification as a result of circulation induced by the wind. Such mixing helps prevent oxygen depletion in bottom waters during the summer. Dissolved-oxygen gradients existed in all the reservoirs, with the smallest concentrations occurring in the near-bottom water of the reservoirs and during the night. Nighttime dissolved-oxygen concentrations in reservoirs 19 and 24 were less than in reservoirs 1 and 9. Similar trends occured with pH. The fluctuations in dissolved-oxygen concentration and pH most likely result from variation in phytoplankton and zooplankton concentrations detected in conjunction with decomposition of organic matter in the bottom sediments.

Most chemical constituents in the water generally became more concentrated from May to August because of cumulative water losses through evapotranspiration. However, this condition was most evident with the major dissolved constituents. Reservoir 19 had sodium bicarbonate sulfate water, reservoirs 1 and 24 had sodium bicarbonate water, and reservoir 9 had sodium sulfate water. Major dissolved constituents and nutrients had little change in concentration during diel sampling. Results of biological analyses were variable. All four reservoirs contained similar types of planktonic organisms but in different proportions. A common pattern for phytoplankton concentrations was a slight increase between the June and July sampling and a decrease from the early to late August sampling. The total number of benthic invertebrates were fewer in reservoir 9 than in reservoirs 1, 19, and 24. However, reservoir 9 had a more diverse benthic community than the other reservoirs. There were no consistent trends in bacterial or zooplankton concentrations among the study reservoirs.

Major water-quality changes that occurred in the reservoirs could affect proposed uses of the study reservoirs. August concentrations of lead and nighttime concentrations of dissolved oxygen in reservoirs 19 and 24 exceeded the criteria protective of fish. Late afternoon pH in all reservoirs approached or exceeded the maximum limits protective of fish propagation, waterfowl habitat, livestock watering, and recreational swimming.

INTRODUCTION

Twenty-four reservoirs (fig. 1) in Phillips and Valley Counties were sampled in 1978 and 1979 to characterize their water quality and to evaluate their suitability for fish propagation, waterfowl habitat, livestock watering, and recreational use. Most of the reservoirs are formed by earthen dams constructed by the U.S. Bureau of Land Management. Reservoir surface areas ranged from 0.2 to 146 ha and maximum reservoir depths ranged from 0.05 to 6.5 m.

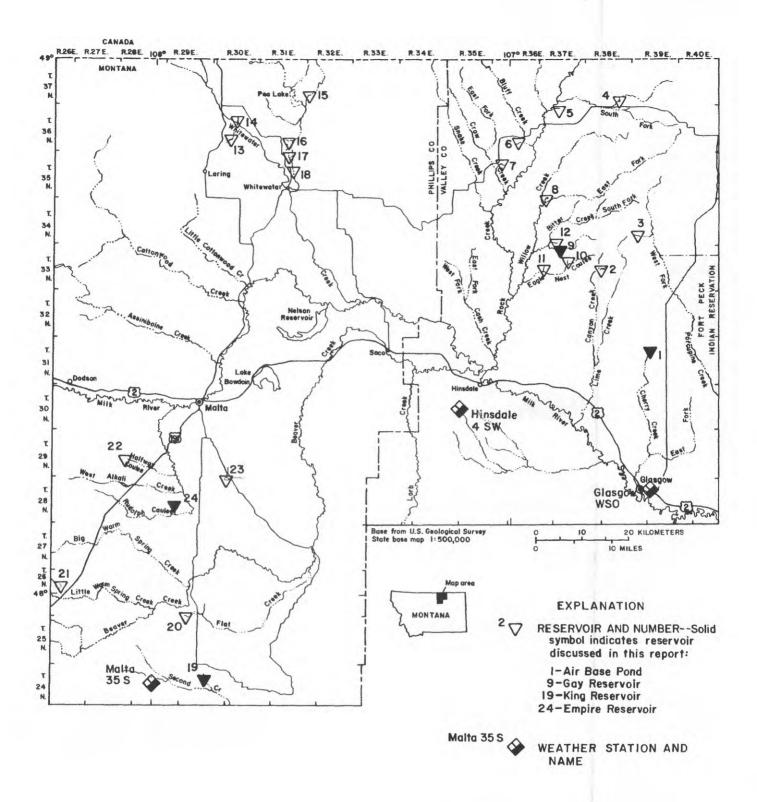
To evaluate each of the 24 reservoirs, water-quality data were collected once during each limnologically extreme period: late winter, late spring, late summer, and early autumn. The data then were compared to a set of criteria protective of each proposed use (Ferreira, 1983). Several of the reservoirs were suitable for all the proposed uses; a few of the reservoirs had water quality that indicated unsuitability for all uses. All the reservoirs generally were enriched with nutrients (eutrophic) and, therefore, had the potential for massive algal blooms and suppressed dissolved-oxygen concentrations (Ferreira, 1980; 1983). The resultant small dissolved-oxygen concentrations could be stressful to fish.

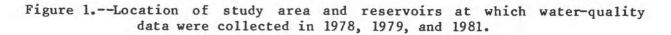
Compared to other times of the year, early spring and late summer commonly are periods of greatest phytoplankton production in lakes and reservoirs, mostly because of increased nutrient availability after spring snowmelt and subsequent warm-water temperatures and long days in late summer (Reid and Wood, 1976). During the summer, thermal stratification and succession of plant and animal populations can augment the variability in chemical water quality. In enriched lakes, water quality can vary considerably during 24 hours as a result of changes produced by photosynthesis, respiration, and decomposition.

To gain a better understanding of the variation, the U.S. Bureau of Land Management entered into a cooperative program with the U.S. Geological Survey in 1981 to investigate May through August and diel (24-hour) changes in water quality in selected reservoirs that had been sampled in 1978 and 1979. Data from this study supplement the quarterly information obtained in 1978 and 1979.

Purpose and scope

The purpose of this report is to describe for four selected reservoirs: (1) the variation in physical, chemical, and biological quality of water occurring approximately monthly from late spring (May) to late summer (August) and during 24 hours in late summer (August) of 1981, and (2) possible causes for water-quality variation during these periods. Although only a subset of the reservoirs sampled in 1978 and 1979 was sampled for this study, there is enough difference within the subset that observed changes in water quality would be representative of similar changes in the remaining reservoirs. The four study reservoirs were chosen pri-





marily on the basis of being average in size among the reservoirs sampled in 1978 and 1979 and of representing considerable ranges in water quality and reservoir age. In 1981, the study reservoirs ranged in age from 9 to 44 years. Reservoirs of different age could provide information that would be helpful in determining the possibility of managing each reservoir through a succession of uses associated with different stages of eutrophication. For example, young reservoirs might be suitable for cold-water fisheries and eventually progress through stages more suitable for warm-water fisheries, waterfowl habitat, and finally livestock watering. The May through August and diel variability in water quality described in this report is intended to be useful for development of long-term management strategies of similar U.S. Bureau of Land Management reservoirs.

Samples were collected monthly from May through July and twice in August; during sampling, depth profiles of temperature, dissolved oxygen, pH, and specific conductance were measured. Water-quality samples were analyzed for major dissolved constituents, selected plant nutrients, and selected trace elements. Biological samples were analyzed for phytoplankton, zooplankton, benthic invertebrates, and bacteria.

During 24 hours in late August, water temperature and dissolved oxygen were recorded continuously, and pH and specific conductance were measured 10 times at a depth of 1 m. Vertical profiles of water temperature, dissolved oxygen, pH, and specific conductance were measured and water samples for chemical and biological analyses were collected twice during the 24 hours.

Study area

The location of the study reservoirs is shown in figure 1. Topography generally is flat except for breaks along the larger streams. Grasses cover most of the landscape, and willow (*Salix* sp.) and cottonwood (*Populus* sp.) trees are abundant in localized areas where water is sufficient for growth. The area generally is used for livestock grazing.

The drainage of each reservoir transects the Bearpaw Shale of Late Cretaceous age (Ross and others, 1955). The Bearpaw Shale is a dark-gray and brown clay shale, which forms gumbo soil when wet. Although much of the drainage overlies the Bearpaw Shale, reservoir 1 also overlies the Flaxville Formation of late Tertiary age. This unit is a brown, yellow, and gray deposit of gravel, sand, and silt with local areas of marl and volcanic ash. A small part of the drainage area for reservoir 9 overlies the Judith River Formation of Late Cretaceous age, which is a light-colored sandstone near the top underlying the Bearpaw Shale and a dark-gray siltstone and sandy shale near the bottom.

The setting and general size of the reservoirs are similar to the many "prairie pothole" lakes that abound in the Upper Midwest (Barica, 1974; Eisenlohr and others, 1972). Generally, "prairie pothole" lakes are shallow (1 to 5 m deep), have no permanent outflow, have no strong thermal stratification (characterized by three regions of different temperatures), and are extremely eutrophic (Barica, 1974). The study reservoirs have surface areas less than 4.0 ha and depths less than 7.0 m. The two older reservoirs (1 and 19) are larger in surface area and deeper than the two newer reservoirs (9 and 24) (table 1).

4

Table 1Location,	construction	date,	and	physical	characteristics	of	the	reservoirs
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 Num	Reservoir		Lo	cation	Year dam	Drain- age area (square - kilom- eters)	Full- pool sur- face area (hec- tares)	Measured depth at sampling location (meters)				
ber (fi 1)	g. Name	Latitude	Longitude	Land-line description	con- struc- ted			Mini- mum	Maxi- mum			
	Valley County											
1 9	Air Base Pond Gay Reservoir	48°26'40" 48°38'30"	106°35'30" 106°51'10"	T. 31 N., R. 39 E., sec. 18 T. 33 N., R. 37 E., sec. 2	1942 1972	2.15 .91	2.7 1.3	3.5 1.2	6.5 4.7			
				Phillips County								
19 24	King Reservoir Empire Reservoir	47°51'00" 48°09'50"	107°51'30" 107°58'00"	T. 24 N., R. 30 E., sec. 15 T. 28 N., R. 29 E., sec. 22	1937 1968	1.22 2.23	3.8 1.8	3.0 1.1	5.3 2.3			

Among the four study reservoirs, only reservoir 1 has extensive growths of woody plants, and these occur along its south and southeast shore. The rock-faced earthen dam of reservoir 19 is covered with a dense growth of wild-rose bushes. However, most of the perimeters of reservoirs in Phillips and Valley Counties are covered with grasses. Compared to reservoirs 1, 19, and 24, reservoir 9 is located in a more deeply incised drainage. Reservoirs 1 and 19 are easily accessible and have been developed as recreational sites. Along the north shore of reservoir 1 are recreational facilities. Portable toilet facilities are provided at reservoir 19 during the summer.

The climate of the study area can be described as continental with hot summers and cold winters. Annual mean air temperatures at weather stations in the study area range from 2.4 to 5.2° C. The warmest month generally is July, with monthly mean temperatures ranging from 18.1 to 21.3° C. January, which generally is the coldest month, has monthly mean temperatures that range from -15.2 to -13.3° C (U.S. Department of Commerce, 1965). Six to 12 cold waves, commonly accompanied by strong winds and blowing snow, usually move through the study area each winter.

The summers are typified by frequent rainshowers and numerous windy periods (Cordell, 1971). Mean annual precipitation at weather stations within the study area ranges from 289 to 340 mm. Generally one-half of this precipitation falls during May, June, and July. Snowfall normally occurs between November and March.

Daily precipitation that occurred from May to August 1981 at weather stations (fig. 1) near each study reservoir is shown in figure 2. Included in the illustration is the cumulative precipitation that occurred between sampling dates. Monthly departures from normal (30-year average) precipitation amounts were available only for the Glasgow WSO weather station near reservoir 1. Generally, precipitation was about 25.4 mm less than normal from May through August of 1981, and for the 4 months prior to the study. By the end of the study, reservoirs 19 and 24 had received the most precipitation among the reservoirs.

Water gains to prairie reservoirs occur principally by overland runoff and ephemeral flow from precipitation and by ground-water inflow. Water losses from the reservoirs occur by seepage through and around each dam, evaporation from the water surface, and transpiration by aquatic and riparian vegetation. As a measure

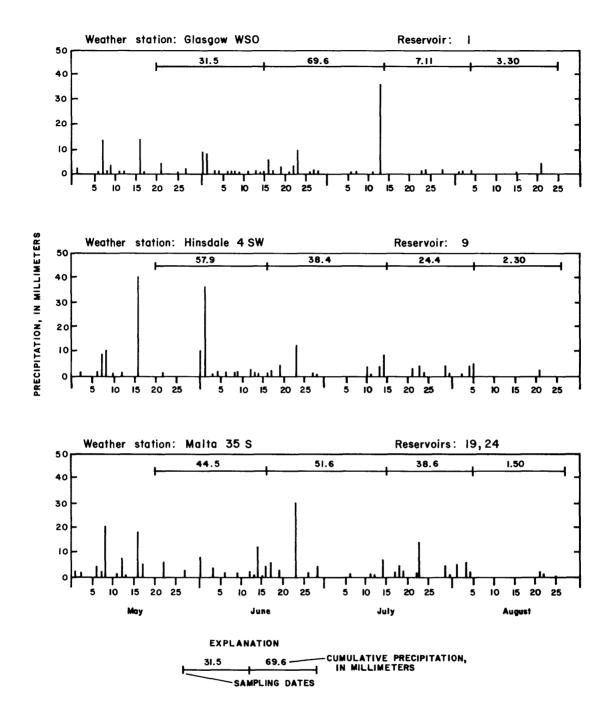


Figure 2.--Daily precipitation at three weather stations from May to August 1981 and cumulative precipitation between five sampling dates.

of the overall effect of water gains and losses, changes in water-surface elevation from the May sampling were recorded for each reservoir (fig. 3). The water-surface elevation loss in reservoir 9 was the largest among the reservoirs. Because of various surface- and ground-water inflow rates, water losses, and drainage area sizes, declines in the water surface do not correlate with total precipitation that occurred among the study reservoirs.

SAMPLING METHODS AND ANALYSIS

Each of the four reservoirs was sampled approximately monthly, starting in May 1981 and ending with two samplings in August 1981. During each of the sampling periods, vertical profiles of selected water-quality variables were made and samples for chemical and biological analyses were collected. Because the largest differences in water quality between night and day commonly occur in late summer, diel sampling was conducted during the sampling period in late August.

Vertical profiles of water temperature, dissolved oxygen, pH, and specific conductance were made using a multiparameter instrument. In each reservoir, measurements were made near the dam, which was estimated to be the deepest part of the original stream channel.

At the same location as the profiles, water samples for chemical analyses were collected with an acrylic Kemmerer¹ water sampler. Where reservoir depths exceeded 2 m, near-surface (1.0 m) and near-bottom samples were collected to represent the water quality of the reservoir. In shallow reservoirs of 2 m or less, one chemical sample was collected at middepth (1.0 m) to represent the water quality of the reservoir. Care was taken not to disturb the reservoir sediment when sampling near the bottom. All samples were pretreated onsite according to methods of the U.S. Geological Survey (Friedman, 1979). Chemical constituents in water samples were analyzed at the U.S. Geological Survey laboratory in Denver, Colo., using methods described by Skougstad and others (1979).

Depth of light penetration was estimated with a Secchi disk. The depth of light penetration was considered to be the average depth of disappearance and reappearance of a black and white disk 200 mm in diameter (Hutchinson, 1967). Light penetration as a percentage of surface light also was profiled with depth, using a relative irradiance meter.

Samples for analysis of phytoplankton and zooplankton were collected from the same location as the chemical analyses. Phytoplankton and zooplankton analyses included enumeration and species identification. Phytoplankton were analyzed by Susswasser Laboratory, Paso Robles, Calif., and zooplankton were analyzed by Harner-White Ecological Consultants, Inc., Littleton, Colo.

Benthic-invertebrate samples were collected during late August in each reservoir at three locations that included deep, intermediate, and shallow water depths.

¹ The use of named products in this report is for identification only and does not constitute endorsement by the U.S. Geological Survey.

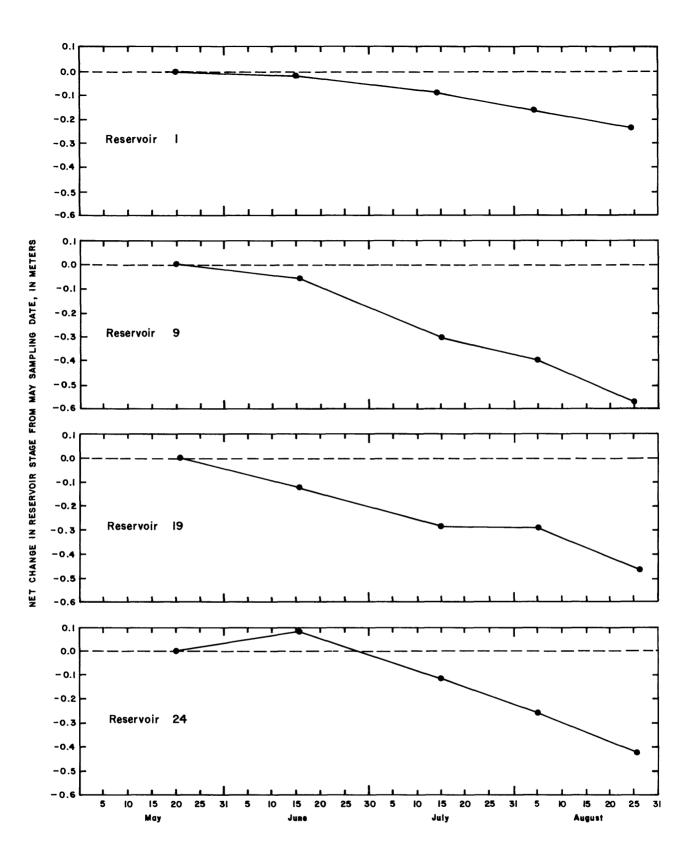


Figure 3.--Net changes in stage of reservoirs 1, 9, 19, and 24 between five sampling dates.

The sampling device used was an Eckman grab sampler having jaw dimensions of 152 x 152 mm. Benthic-invertebrate samples were analyzed by Michael Fillinger, Helena, Mont. The samples were preserved and the organisms identified to species using procedures described by Greeson and others (1977).

Water samples for total coliform, fecal coliform, and fecal streptococcal bacterial analyses were collected at the chemical sampling site and along the shore of easiest access for livestock at each reservoir. Bacteria were analyzed according to procedures described by Greeson and others (1977).

Diel sampling during late August, at the same location as the regular sampling, consisted of continuous recording of temperature and dissolved oxygen at a depth of 1.0 m. The probe of a continuous recording instrument for measuring temperature and dissolved oxygen was suspended along with multiparameter probes by floats tethered from an anchor (fig. 4). Specific conductance and pH were measured about every 3 hours during the same 24 hours as the continuous measurements. Vertical profiles of temperature, dissolved oxygen, pH, and specific conductance were obtained twice daily--about 1.5 hours before sunset and 1.5 hours before sunrise.

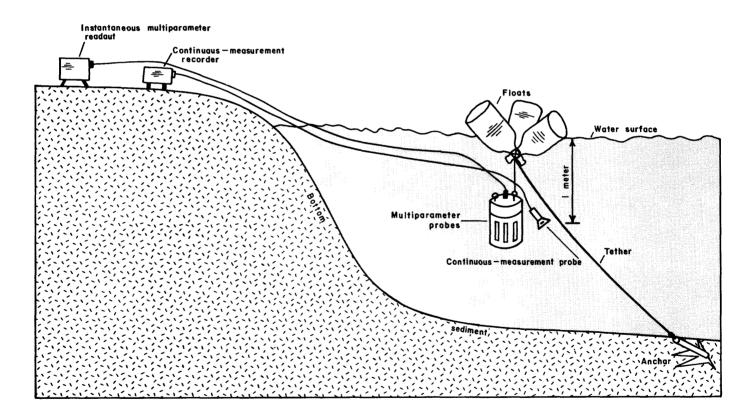


Figure 4.--Probe suspension for measurement of temperature, dissolved oxygen, pH, and specific conductance at a depth of 1 meter during the late August diel sampling.

Diel samples for major dissolved constituents, nutrients, trace elements, and chlorophyll a and b were collected at the same times as the profiles (1800 and 0500

hours). Chlorophyll a and b samples were analyzed by the U.S. Geological Survey laboratory in Denver, Colo., following methods described by Greeson and others (1977).

MAY THROUGH AUGUST TRENDS

Onsite measurements

Depth profiles of temperature, dissolved oxygen, pH, and specific conductance are described for the five sampling dates in this section of the report. Waterquality data collected during the nighttime phase of the diel sampling in late August are described in the section titled "Diel trends."

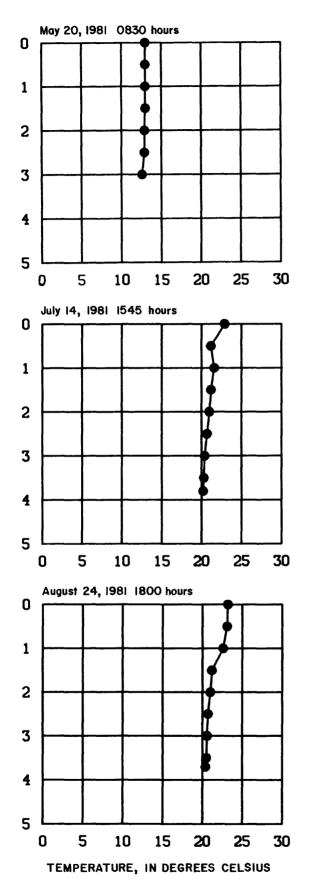
Temperature

Water temperatures during the five samplings (figs. 5-8) ranged from 12.6°C in the near-bottom water of reservoir 1 in May to 24.0°C in the near-surface water of reservoir 9 in late August. Water temperatures in the reservoirs generally followed the trend of air temperature, with the minimum temperatures occurring in May and the maximum temperatures occurring in late August. However, because of unseasonably warm temperatures during sampling in May, temperature gradients with depth generally were larger in May than in June.

Deep reservoirs have a larger heat capacity than shallow reservoirs, which allows large temperature differences to develop between the near-surface and nearbottom water as summer progresses. These large temperature differences can result in distinct density layers that resist mixing and become thermally stratified into three temperature regions: epilimnion, metalimnion, and hypolimnion. In shallow reservoirs having less heat capacity, only small temperature differences develop between near-surface and near-bottom water as summer progresses. With small temperature differences, there is less resistance to wind-induced mixing and the water does not become thermally stratified. The slight temperature gradient that occurs from the surface to the bottom in shallow reservoirs when distinct water layers do not form is termed "diminutive thermal stratification" (Wetzel, 1975).

None of the reservoirs had thermal stratification consisting of an epilimnion, metalimnion, and hypolimnion; however, at times they all had diminutive thermal stratification (figs. 5-8). Instead of increasing from May to August, the degree of temperature difference between the near-surface and near-bottom water was variable from month to month, which indicates that periods of complete mixing probably occur throughout the summer. In fact, the largest temperature differences existed in reservoirs 9 (5.2 C°) and 24 (6.0 C°) during the May sampling, which were followed by homogeneous temperature profiles during the June sampling. In addition to heat capacity, time of day probably is a determining factor in measured temperature differences between near-surface and near-bottom water. During the early morning after night-long cooling, temperature gradients may be small enough to allow total mixing by the wind. By late afternoon, the near-surface water may warm enough to resist mixing with the cooler near-bottom water.





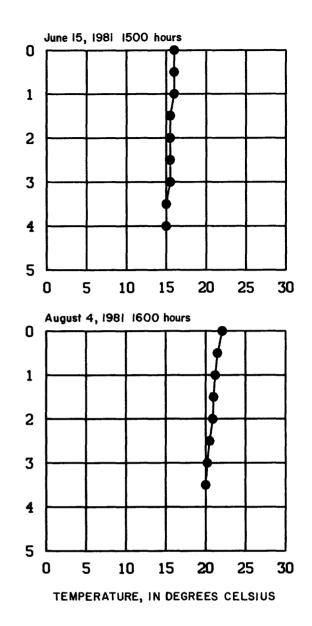
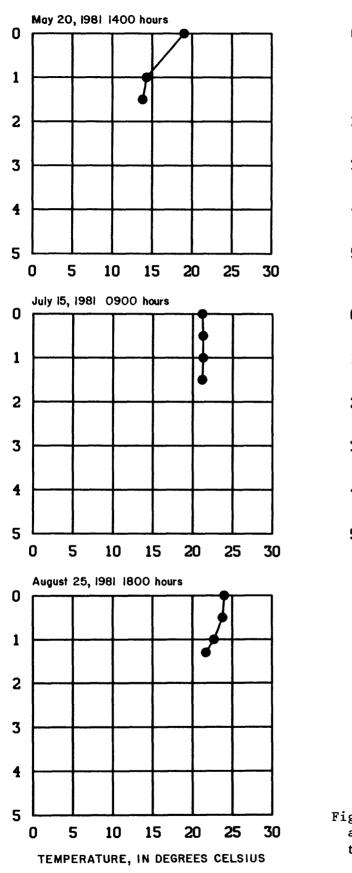


Figure 5.--Depth profiles of temperature on five sampling dates (May through August) in reservoir 1.





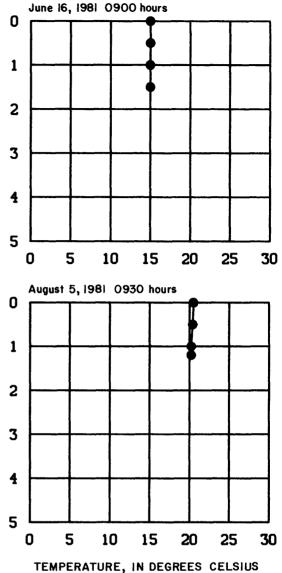
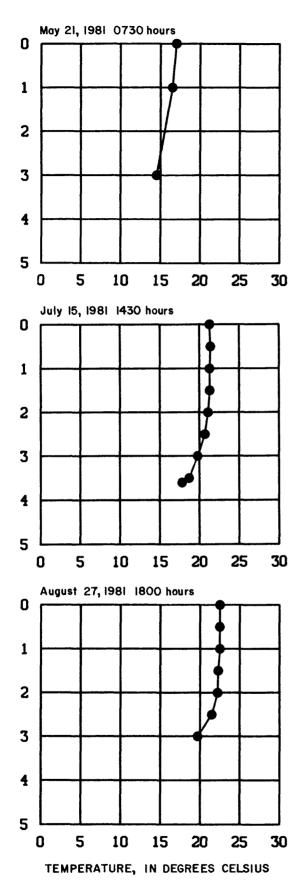


Figure 6.--Depth profiles of temperature on five sampling dates (May through August) in reservoir 9.





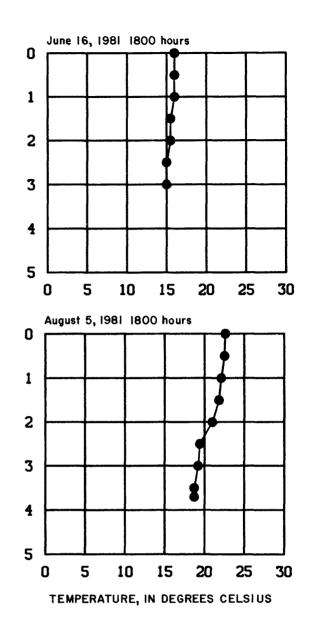
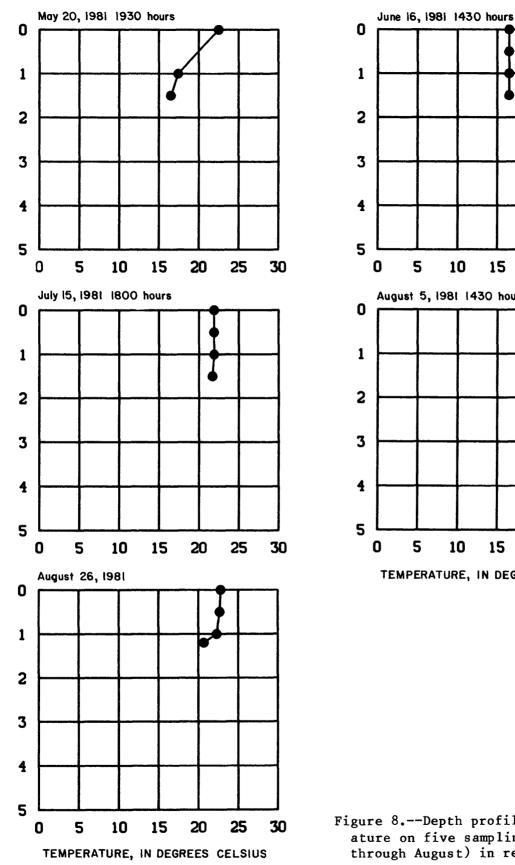


Figure 7.--Depth profiles of temperature on five sampling dates (May through August) in reservoir 19.





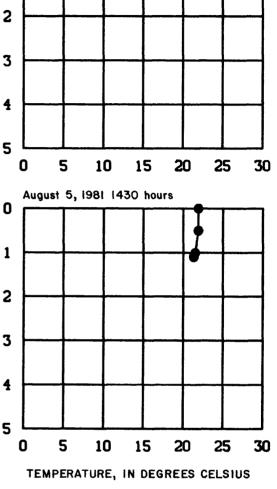


Figure 8.--Depth profiles of temperature on five sampling dates (May through August) in reservoir 24.

Dissolved oxygen

Dissolved-oxygen concentrations measured during daylight (figs. 9-12) ranged from 0.3 mg/L (milligram per liter) in the near-bottom water of reservoir 19 during late August to 13.4 mg/L in the near-surface water of reservoir 19 during early August. With the exception of reservoir 9, the largest dissolved-oxygen concentration in the near-surface water of each reservoir was measured during either early or late August. In reservoir 9, dissolved-oxygen concentrations were largest in May and June possibly because of increased production by a well-established macrophyte population in response to the influx of nutrients from spring runoff.

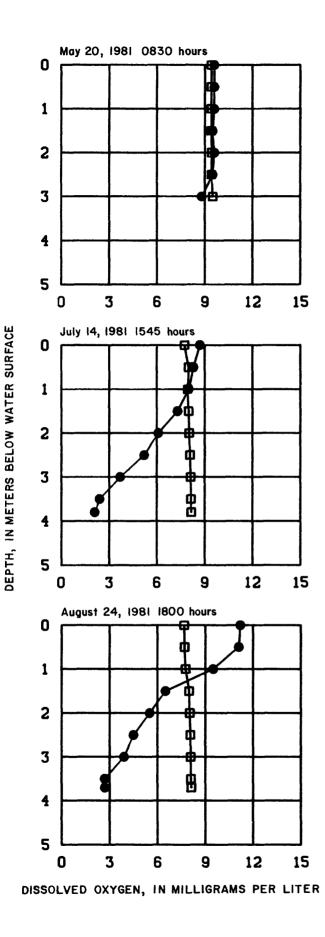
Dissolved-oxygen gradients existed in all reservoirs; the dissolved-oxygen concentrations generally were maximum and larger than saturation near the surface, and minimum and smaller than saturation near the bottom. The largest differences in dissolved-oxygen concentrations between near-surface and near-bottom water in each reservoir generally occurred during either early or late August sampling.

Reservoir 9 generally had the smallest dissolved-oxygen concentration differential during each sampling and maintained larger dissolved-oxygen concentrations in the near-bottom water than reservoirs 1, 19, and 24. The large oxygen concentrations in the near-bottom water of reservoir 9 probably resulted in part from photosynthesis by macrophytes and attached algae that were growing across the entire reservoir bottom. In contrast, reservoirs 1 and 19 generally had the largest differences in dissolved-oxygen concentration from near-surface to near-bottom water and the smallest dissolved-oxygen concentration in the near-bottom water. The decomposition of black organic muds observed in the bottom sediments of reservoirs 1 and 19 most likely contributed to decreased concentrations of dissolved oxygen.

The dissolved-oxygen gradient between the near-surface and near-bottom water mainly is a result of photosynthesis by aquatic plants in the euphotic zone and respiration and decomposition of aquatic plants and animals in the sediment. Because of dissolved-oxygen consumption by decomposition and respiration in the near-bottom water coupled with little replacement by photosynthesis, near-bottom water generally has less dissolved oxygen than near-surface water.

The shallowness of the reservoirs helps prevent the development of anoxic conditions that would limit their usefulness. Periodic mixing during the summer replenishes dissolved oxygen that is consumed by decomposition in the bottom water. If stratification occurred in these reservoirs at higher reservoir stages during wet years, the near-bottom water could possibly become anaerobic by late summer. The possibility of this condition is indicated by an increase in the difference of dissolved-oxygen concentrations between near-surface and near-bottom water from May to August. By preventing anaerobic conditions in the near-bottom water, mixing could moderate water-quality changes in other constituents such as trace elements, that otherwise would occur as uncirculating near-bottom water formed a reducing environment capable of bringing trace elements into solution. Conversely, mixing throughout the entire depth of the reservoir may recirculate nutrients deposited in the bottom sediments. Circulation could increase plant production and lead to potential dieoffs, with subsequent oxygen consumption. These contrasting physical processes could lead to fluctuating dissolved-oxygen concentrations throughout the summer.

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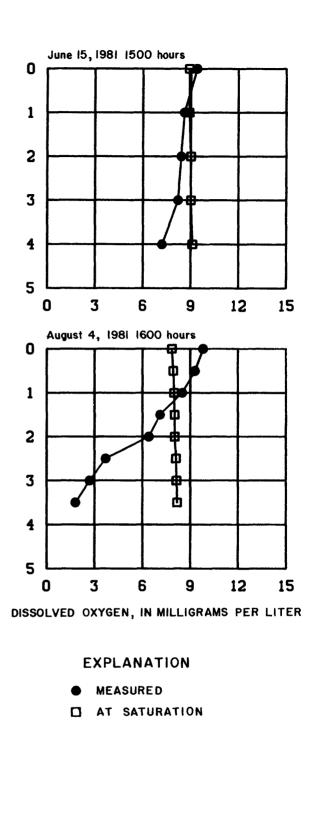
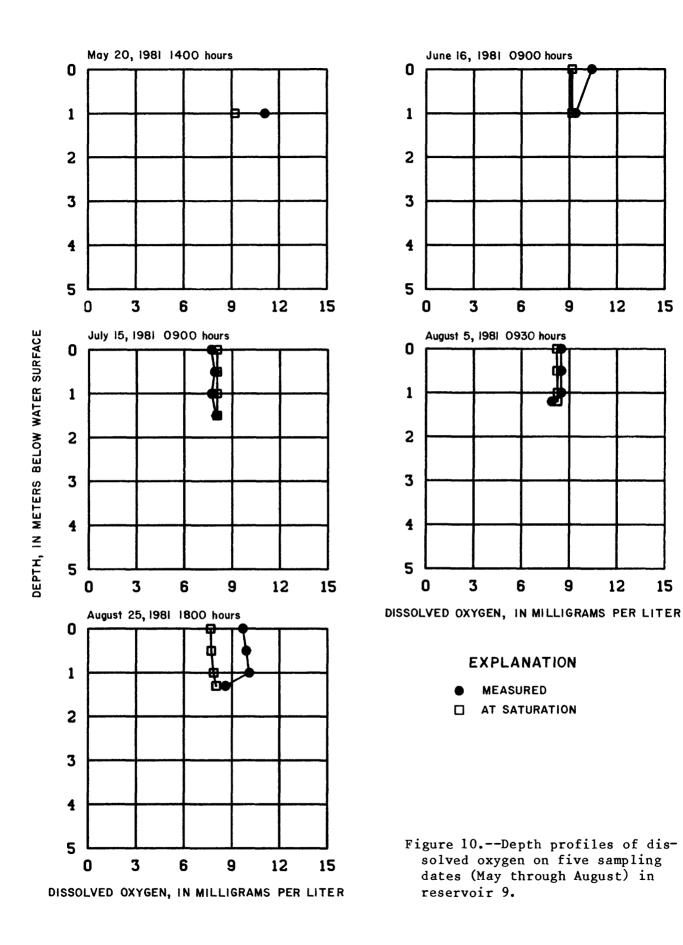
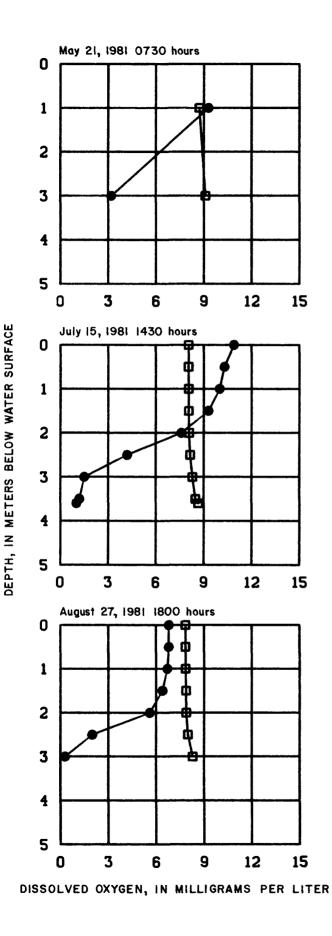


Figure 9.--Depth profiles of dissolved oxygen on five sampling dates (May through August) in reservoir 1.





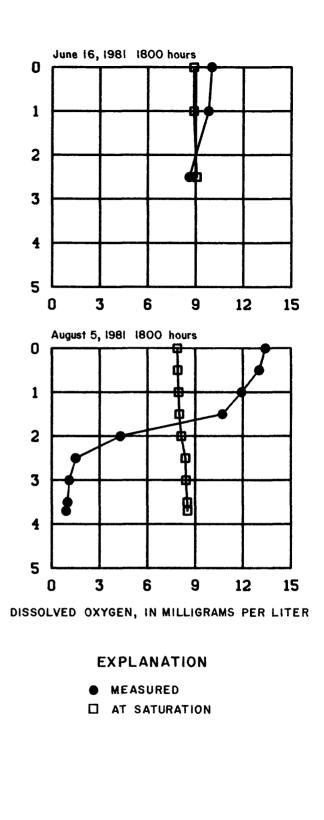
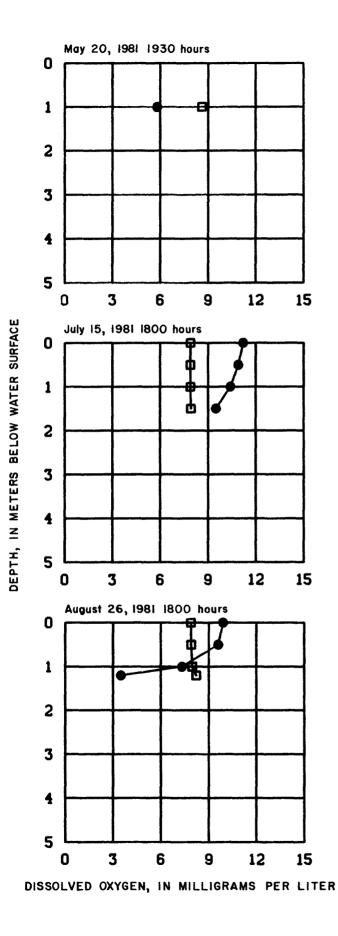


Figure 11.--Depth profiles of dissolved oxygen on five sampling dates (May through August) in reservoir 19.



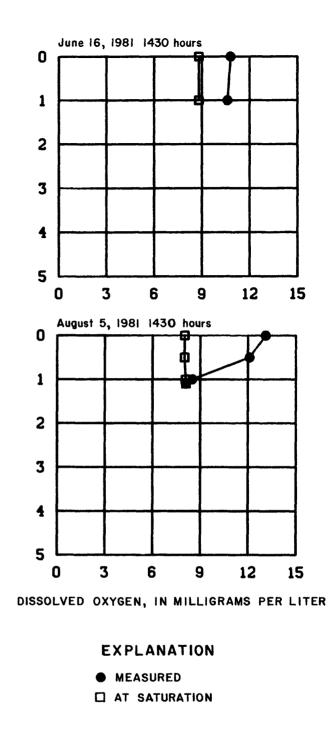


Figure 12.--Depth profiles of dissolved oxygen on five sampling dates (May through August) in reservoir 24.

Values of pH in the near-surface water of the study reservoirs (figs. 13-16) ranged from 8.1 in reservoir 24 in June to 9.8 in reservoir 9 in late August. Reservoirs 1 and 9 generally had larger pH values than reservoirs 19 and 24. The smallest pH, 7.9, occurred in the near-bottom water of reservoir 19 in May.

In May, June, and July, pH values generally were homogeneous with depth in reservoirs 1, 9, and 24, whereas values decreased with depth in reservoir 19. In these months, the pH differences between the near-surface and near-bottom waters in reservoir 19 ranged from 0.2 to 0.8 standard unit. In each reservoir the largest pH gradients usually occurred in August. At this time the pH differences between the near-surface and near-bottom water for reservoir 19 were the largest among the reservoirs, ranging from 0.7 to 0.9 standard unit.

A byproduct of respiration and decomposition is CO_2 (carbon dioxide), which on entering the water decreases the pH. During photosynthesis CO_2 dissolved in water is utilized, thereby increasing the pH during the day. Differences in pH between the near-surface and near-bottom water in the study reservoirs most likely result from photosynthesis, respiration, and decomposition in a manner similar to that of dissolved oxygen.

Of the four proposed reservoir uses, recreational swimming would be limited in all the study reservoirs because of measured pH values greater than the recommended maximum of 8.3 (National Technical Advisory Committee to the Secretary of the Interior, 1968). During August in reservoir 9, fish propagation, waterfowl habitat, and livestock watering might be affected adversely by pH at or near recommended values-9.0 for fish and livestock (U.S. Environmental Protection Agency, 1978) and 9.2 for waterfowl (National Technical Advisory Committee to the Secretary of the Interior, 1968).

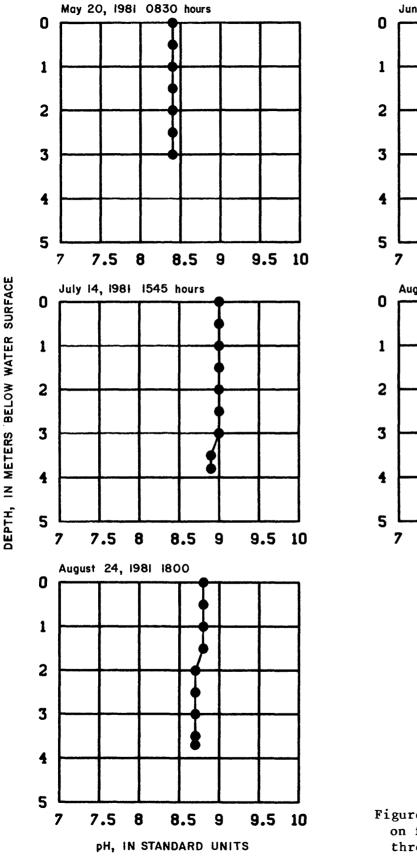
Specific conductance

The reservoirs represent a considerable range of specific conductance (figs. 17-20). Reservoir 24 had the smallest specific-conductance values, which ranged from 192 to 308 μ S/cm (microsiemens per centimeter at 25° Celsius). Reservoir 9 had the largest specific-conductance values, which ranged from 3,500 to 4,610 μ S/cm.

Minimal specific-conductance gradients consisting of a slight increase near the bottom were determined in each reservoir. However, middepth maximum specificconductance values were measured in reservoir 1 during the July and August sampling and in reservoir 9 during the May sampling.

The differences in specific conductance among the study reservoirs are related to differences in local mineralogy and the effect of varying hydrologic factors. Mineralogy differences are discussed in the section of this report titled "Chemical analyses, major dissolved constituents."

Ground-water flow, surface-water flow, evaporation, transpiration, and precipitation control the net volume of water in each reservoir. Surface-water inflow and evaporation are major factors that cause seasonal changes in specific conductance by changing the concentration of dissolved solids. Probably because of water



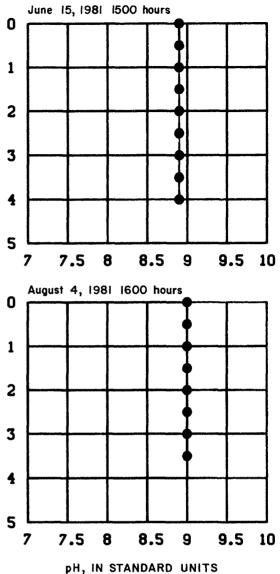
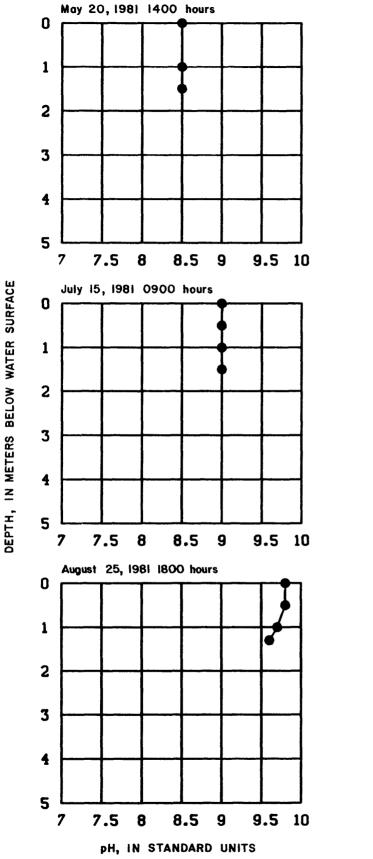


Figure 13.--Depth profiles of pH on five sampling dates (May through August) in reservoir 1.



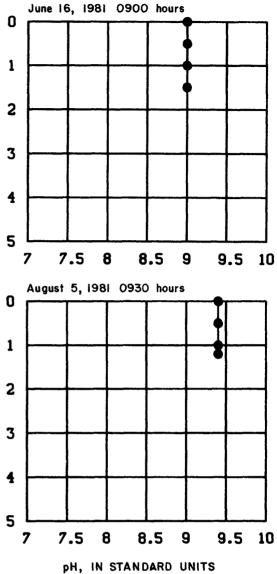
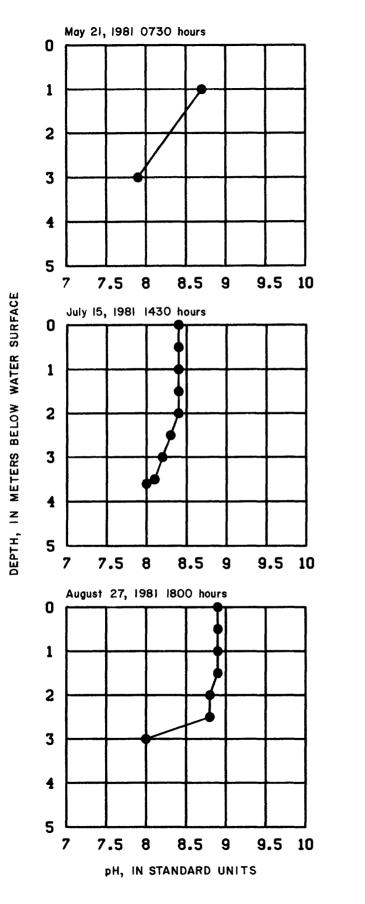


Figure 14.--Depth profiles of pH on five sampling dates (May through August) in reservoir 9.



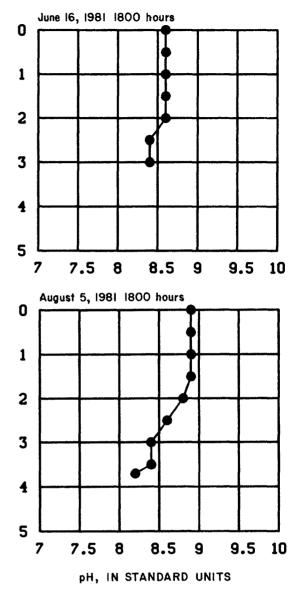
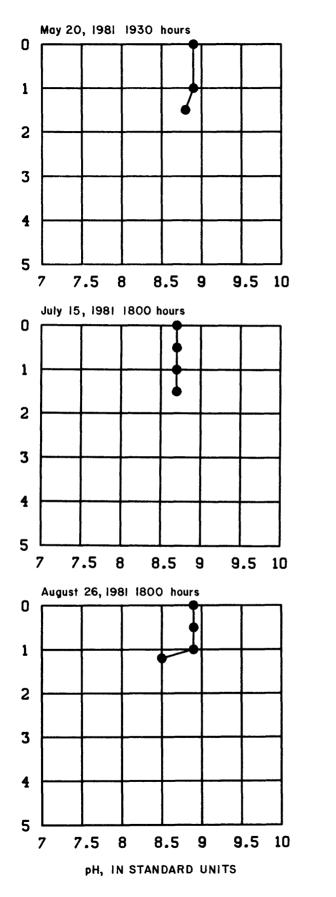


Figure 15.--Depth profiles of pH on five sampling dates (May through August) in reservoir 19.





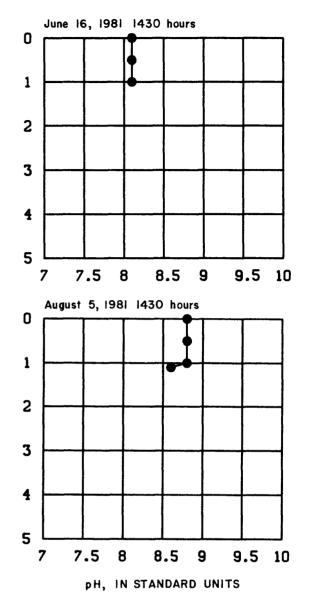
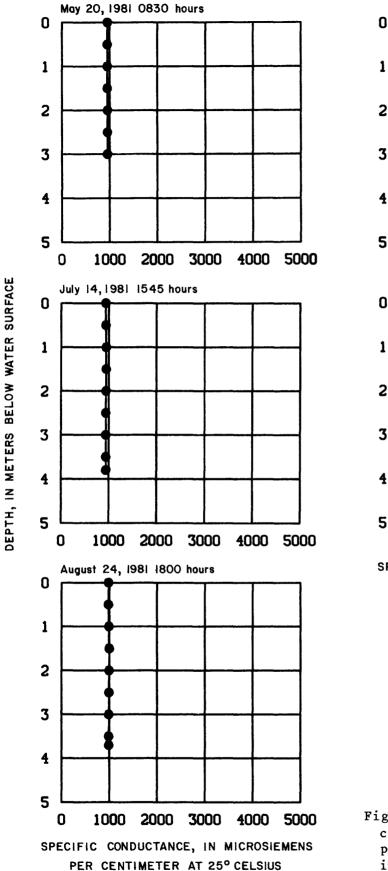
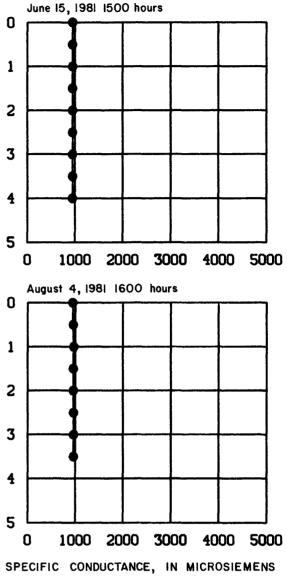


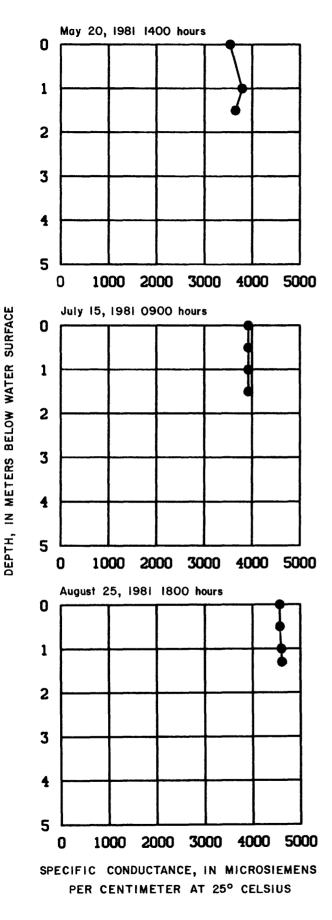
Figure 16.--Depth profiles of pH on five sampling dates (May through August) in reservoir 24.

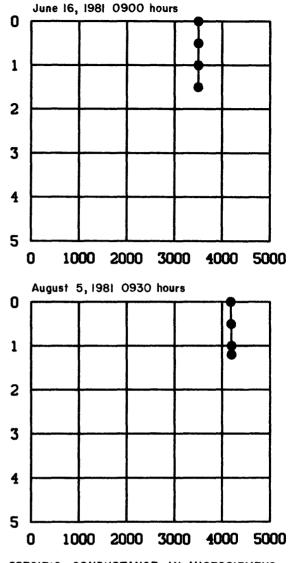




PER CENTIMETER AT 25° CELSIUS

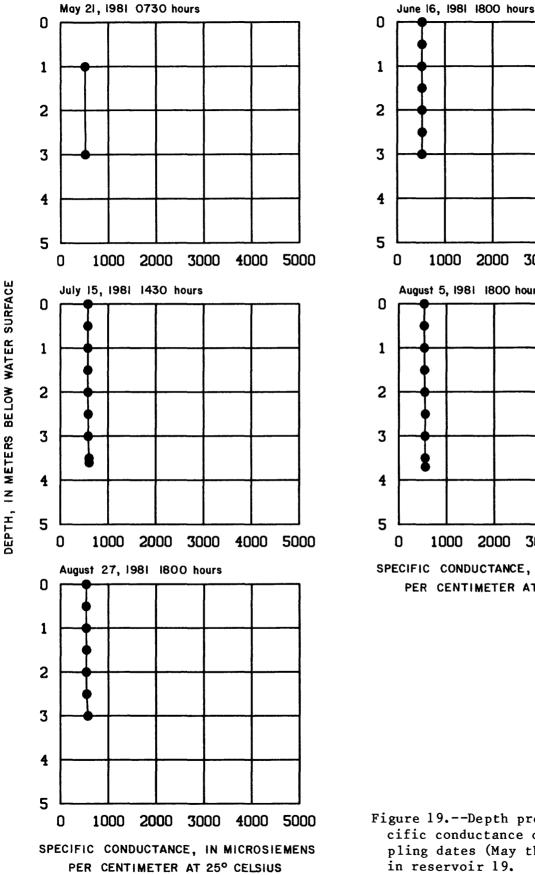
Figure 17.--Depth profiles of specific conductance on five sampling dates (May through August) in reservoir 1.





SPECIFIC CONDUCTANCE, IN MICROSIEMENS PER CENTIMETER AT 25° CELSIUS

Figure 18.--Depth profiles of specific conductance on five sampling dates (May through August) in reservoir 9.



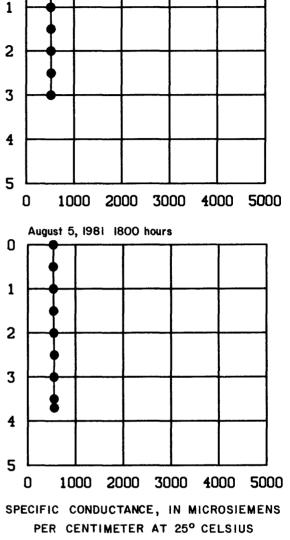
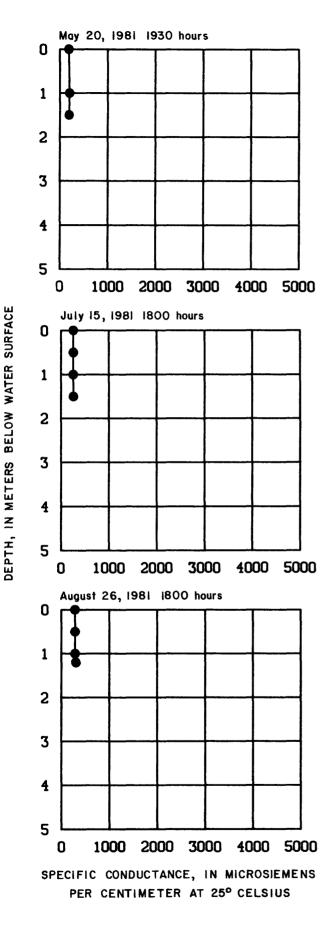
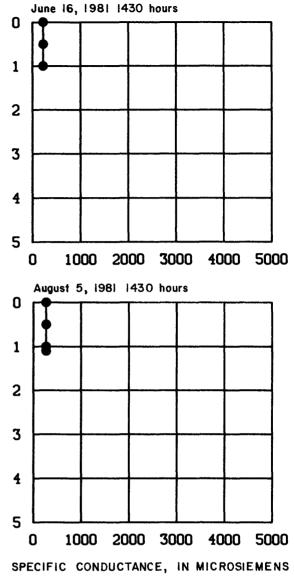


Figure 19.--Depth profiles of specific conductance on five sampling dates (May through August) in reservoir 19.





PER CENTIMETER AT 25° CELSIUS

Figure 20.--Depth profiles of specific conductance on five sampling dates (May through August) in reservoir 24.

loss with evaporation and transpiration, dissolved-solids concentrations generally increase from May to August. Among the reservoirs, the occurrence of minimum specific conductances during times other than May can be related to the occurrence of summer precipitation. Given a relatively large drainage area, summer precipitation could provide sufficient surface-water inflow to dilute the dissolved-solids concentration. About 38.1 mm of precipitation a few days prior to sampling reservoir 9 in June and reservoir 1 in July most likely was the cause of minimum specific conductances after May.

Light penetration

The small percentage of surface light penetration and the shallow Secchi-disk depth both indicate that reservoirs 19 and 24 are more turbid than reservoirs 1 and 9 (fig. 21). The greatest light penetration was observed in reservoir 1 where two Secchi-disk depths were greater than 1.5 m and light sufficient for photosynthesis and probably net primary production (greater than 1 percent of the incident light) occurred near the bottom. Although reservoir 9 did not have the clarity of reservoir 1, sufficient light for photosynthesis also reached the bottom because of the shallow depths of the reservoir. Reservoir 9 had macrophytes (aquatic plants) rooted across the lake bottom.

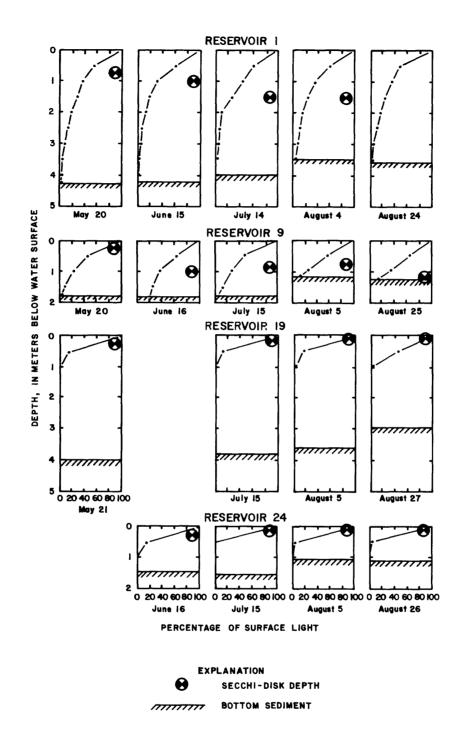
Reservoir 1 showed a gradual increase in percentage of light transmission and Secchi-disk depth from May to July. In reservoir 9 there was an abrupt increase in percentage of light transmission and Secchi-disk depth from May to June. Compared to May, June, and July, the August percentage of surface light in reservoir 9 showed a more linear decrease with depth. However, the Secchi-disk depths in reservoir 9 were variable during the study. Light penetration and Secchi-disk depths in reservoirs 19 and 24 remained about the same during the study.

Seasonal changes in light penetration among the reservoirs are affected by a combination of factors, including concentration of suspended materials transported by spring runoff, seasonal phytoplankton succession, shoreline disturbance by cattle, and wind-induced mixing of the reservoir, which resuspends bottom sediments. The greater light penetration in reservoirs 1 and 9 could be the result of wind protection afforded by adjacent hills. Reservoirs 19 and 24 are in relatively open areas and, therefore, are subject to more frequent wind mixing. Reservoirs 1 and 9 also contained smaller phytoplankton concentrations than reservoirs 19 and 24, which would allow greater light penetration (see Biological analyses, Phytoplankton).

Chemical analyses

Major dissolved constituents

Relative concentrations of the major dissolved constituents indicate that reservoirs 1 and 24 have sodium bicarbonate water, reservoir 9 has sodium sulfate water, and reservoir 19 has sodium bicarbonate sulfate water (table 4; Supplemental Information section at back of report). Reservoir 9 had the largest dissolvedsolids concentrations (2,900-3,790 mg/L) and reservoir 24 had the smallest (123-162 mg/L). No consistent monthly trend occurred at each reservoir for each major dissolved constituent; however, the dissolved-solids concentration (summation of all constituents with a correction for CO_2 loss) increased slightly from May to August.



. Figure 21.--Relationship of depth to percentage of surface light and Secchi-disk depth in reservoirs 1, 9, 19, and 24 on five or less sampling dates (May through August).

Although several hydrologic factors function in combination with local mineralogy to produce the major dissolved constituents present during any given time, local mineralogy of the drainages for each reservoir is indicated to be a principal factor affecting the composition of constituents. The reservoirs are regionally close to one another, but only drainages of reservoirs 19 and 24 are located entirely within the area of the Bearpaw Shale. Both of these reservoirs had small dissolved-solids concentrations compared to reservoirs 1 and 9. However, the differences in water quality as a result of local mineralogy is indicated by the smaller dissolved-solids concentration (smaller specific conductance) and the sodium bicarbonate type water in reservoir 24 compared to the larger dissolved-solids concentration (larger specific conductance) and the sodium bicarbonate sulfate water in reservoir 19.

Continued leaching of major dissolved constituents from the drainage and subsequent concentrating of major dissolved constituents with evaporation are expected to increase dissolved-solids concentrations as reservoirs age. Yet, the results of samples from the reservoirs do not indicate that age is a principal factor in determining the relative concentration of dissolved constituents. For example, reservoir 9 had the largest dissolved-solids concentration even though it is the newest reservoir and reservoir 19 had the second smallest dissolved-solids concentration even though it is the oldest reservoir. Within each reservoir, dissolved-solids concentration could increase with age as indicated by the smaller dissolved-solids concentrations that were measured in 1978 and 1979 (Ferreira, 1980; 1983). However, these differences are affected by the volume of water gain or loss and commensurate diluting or concentrating that occurred prior to sampling.

The percentage increase in dissolved constituents from May to August due to evaporative water losses is affected by the ratio of the reservoir's volume to surface area. For a given volume, those reservoirs having larger surface area (shallow) would have a larger percentage loss of total volume of water for a given evaporation rate. Reservoirs 9 and 24, being relatively shallow compared to reservoirs 1 and 19, had a larger percentage increase in dissolved-solids concentration. The greater depths of reservoirs 1 and 19 resulted in a smaller percentage of water loss and, consequently, a smaller percentage increase in dissolved-solids concentration. Percentage increases in dissolved-solids concentration from May to August ranged from about 1 percent in reservoir 1 to 29 percent in reservoir 9.

Reservoirs 1 and 19 each were sampled at two depths. Both reservoirs showed slight differences in dissolved-solids concentration between the near-surface and near-bottom water samples. The slight increases in dissolved-solids concentration determined in most samples of near-bottom water of the reservoirs could originate from dissolution of material in the bottom sediments.

Nutrients

Total nitrogen concentrations ranged from 1.4 mg/L in reservoirs 1 and 9 to 7.5 mg/L in reservoir 24 (table 5; Supplemental Information section at back of report). In reservoirs 19 and 24, total nitrogen concentrations were smaller in May and June than in July and August. In reservoirs 1 and 9, total nitrogen was variable from May to August, although both had maximum concentrations in early August. In reservoirs 1 and 19, differences in total nitrogen concentrations between the near-surface and near-bottom water were slight. Because total nitrogen is composed mostly of total organic nitrogen in these reservoirs, observed trends are a reflection

of trends in total organic nitrogen. Ammonia also showed trends similar to total nitrogen.

Increases of nitrogen concentrations in the reservoirs from May to August could result from the concentrating effect of evaporative water loss, continued input of nitrogen from decomposition in the bottom sediments, and external inputs from activities close to each reservoir such as grazing livestock. In addition, increases could result from nitrogen fixation by blue-green algae. Decreases can occur because of nitrogen uptake by aquatic plants or the settling of sediment to which nitrogen has sorbed. The variation in total nitrogen probably is due to a combination of all the above factors, of which the relative concentration of each can be different for each reservoir.

Reservoir 9 had the smallest total phosphorus concentrations (<0.01-0.10 mg/L, table 5). Reservoir 24 had the largest total phosphorus concentrations (<0.01-0.34 mg/L). Dissolved orthophosphorus concentrations were similar among all the reservoirs, except for larger concentrations in reservoir 1 during August.

Phosphorus can be derived from many of the same sources as nitrogen, but trends in phosphorus concentrations were not always similar to those of nitrogen. Only reservoir 1 had an increase of phosphorus from May to August. Reservoir 9 had the largest total phosphorus concentration occurring in May, presumably from spring runoff. Measured concentrations of nitrogen (total nitrogen greater than 1.1 mg/L N) and phosphorus (total phosphorus greater than 0.03 mg/L P) indicate that the reservoirs are enriched with nutrients (Ferreira, 1983).

Whether nitrogen or phosphorus is limiting phytoplankton growth can be indicated by comparing the concentrations of inorganic nitrogen $(NO_2 + NO_3 plus ammonia)$ to orthophosphorus. If inorganic nitrogen is more than 10 times the orthophosphorus concentration, then phosphorus generally is limiting. If inorganic nitrogen is less than five times the orthophosphorus concentration, then nitrogen commonly is limiting (Zison and others, 1977).

Generally, nutrient ratios are calculated using dissolved concentrations, because these forms are readily available for plant uptake. Because dissolved ammonia concentrations were not available, the ratios in this study were calculated with total concentrations of inorganic nitrogen and dissolved concentrations of orthophosphorus to indicate what the limiting nutrients might be. The dissolved and total concentrations of NO₂ + NO₃ were similar in samples collected during this study. However, nutrient samples collected in previous studies (Ferreira, 1980, 1983) indicate that the total ammonia concentration is more variable, ranging from being equal to being greater than the dissolved ammonia concentration. Therefore, when the ratio indicates nitrogen is the limiting nutrient (N/P < 5), using dissolved values of nitrogen will not change the interpretation. However, when the ratio indicates phosphorus is the limiting nutrient (N/P > 10), the interpretation may be questionable owing to the possibility of total ammonia concentration being considerably larger than that of dissolved ammonia. But, because nitrogen is not as strongly sorbed to sediment as phosphorus (total forms), total ammonia can be considered available for plant uptake. Therefore, the ratios calculated from total nitrogen concentrations are useful as a general indication of nutrient availability in the study reservoirs.

Based on nitrogen and phosphorus ratios, all the reservoirs were phosphorus limited in May but became nitrogen limited in June. In July all the reservoirs

remained nitrogen limited except reservoir 19, which again became phosphorus limited. In August only reservoir 1 remained nitrogen limited; the other reservoirs were phosphorus limited. Changes in the role of limiting nutrients probably occur because of the uptake and release of nutrients by fluctuating phytoplankton populations and sorption and desorption by sediments resuspended during mixing.

Relative differences among the reservoirs in total organic-carbon concentrations are similar to the relative differences for total phosphorus (table 5). Because the production of organic matter is in part dependent upon the availability of phosphorus, and the supply of carbon from atmospheric CO_2 is unlimited, relative differences in organic carbon among the reservoirs would be the same as phosphorus in the absence of major external sources. In general, the quantity of total organic carbon in the reservoirs is proportional to the number of planktonic organisms in each reservoir.

Reservoirs tend to progress through stages of increased nutrient enrichment with age. As productivity rates increase with nutrient supply, the suitability of a reservoir for different uses can change. Although nutrient concentrations in each reservoir probably are increasing with reservoir age, the variable concentrations of nutrients observed among the reservoirs of different ages do not show this relationship. Therefore, nutrient concentrations are more affected by local differences in mineralogy and land use than by aging of the study reservoirs.

Trace elements

Of all the trace elements analyzed for, manganese was present in the largest concentrations (table 6; Supplemental Information section at back of report); reservoirs 19 and 24 generally had the largest concentrations of total recoverable manganese among the study reservoirs. Total recoverable lead occurred in large concentrations in reservoirs 19 and 24, but only in late August. Dissolved iron had the second largest concentrations among the trace elements; the largest concentratrations occurred in reservoirs 9 and 24. Differences in trace-element concentrations are most likely caused by differences in local mineralogy.

In reservoirs 1, 19, and 24 the concentration of total recoverable manganese generally increased from May to August. In reservoir 9 total recoverable and dissolved manganese decreased from May to August. The only other consistent patterns among the trace elements were decreases in dissolved iron and manganese in reservoir 24 from May to August.

During each sampling, the near-bottom water in reservoirs 1 and 19 generally had larger trace-element concentrations than the near-surface water. Seasonal changes in the concentrations of trace elements are related to the concentrating effect of water loss through evaporation and changes in trace-element equilibria caused by biological processes. In late August, total recoverable lead in reservoirs 19 and 24 exceeded by the greatest degree the concentration recommended by the U.S. Environmental Protection Agency (1980) for the protection of fish (9.9 μ g/L at a hardness as calcium carbonate of 150 mg/L).

Biological analyses

Phytoplankton

Except for reservoir 9 in late August and reservoir 24 in May, Chlorophyta (green algae) was either dominant or codominant (equal to or greater than 15 percent) in all reservoirs during each sampling (table 7; Supplemental Information section at back of report). Cyanophyta (blue-green algae) generally was either dominant or codominant in May and August in all the reservoirs except reservoir 1. Concentrations of Cyanophyta were not detected as being dominant in reservoir 1 during the study. Cryptophyta (cryptomonads) was codominant in July or August in reservoirs 1, 9, and 19 and codominant in May in reservior 24. Bacillariophyta (includes diatoms) was dominant or codominant in late August in reservoir 1, in July in reservoir 9, in June in reservoir 19 and in May, June, and late August in reservoir 24. Euglenophyta (euglenoids) were dominant or codominant in May in reservoir 9 and in late August in reservoir 24.

The numbers and types of phytoplankton differ from one body of water to another; no truly "typical" community exists (Reid and Wood, 1976). The community composition of phytoplankton involves complex interactions by many factors such as nutrient loading, reservoir mixing, water type, temperature change, light availability, physiology and reproductive rate of the phytoplankton, and harvest rate by zooplankton. Therefore, similar types of phytoplankton occurring in different proportions in each of the study reservoirs is expected. Green algae, blue-green algae, and flagellates (which include cryptomonads) collected from the study reservoirs are characteristically abundant in warm, nutrient-enriched water, as opposed to cold, nutrient-deficient water in which diatoms generally are most abundant.

Because of the ability of blue-green algae to fix elemental nitrogen, they typically have late summer growth that exceeds other groups of algae as inorganic nitrogen ($NO_2 + NO_3$ plus ammonia) becomes depleted. This late summer growth could account, in part, for the dominance or codominance of blue-green algae in the study reservoirs. However, other factors probably are interacting with phytoplankton, because blue-green algae were not always dominant or codominant during June and July when nitrogen is indicated to be limiting. In reservoir 1, which was indicated to be nitrogen limited all months except May, blue-green algae were never detected as being dominant.

Phytoplankton cell concentrations (fig. 22) at a depth of 1 m were largest in reservoir 24 and smallest in reservoirs 1 and 9. The large phytoplankton cell concentrations in reservoir 24 correspond to large nutrient concentrations (tables 5 and 7). The smaller phytoplankton cell concentrations in reservoirs 1 and 9 correspond to smaller nutrient concentrations in these reservoirs. Although reservoirs 1 and 9 are indicated to be mesotrophic by their phytoplankton cell concentrations (between 1,000 and 15,000 cells per mL; Taylor and others, 1980), they both support large growths of macrophytes. Sampling during other years or at other times might indicate larger concentrations of phytoplankton than determined during this study.

Throughout the season, the abundance of major groups of phytoplankton varies in response to environmental changes. Generally for temperate lakes, two pulses of phytoplankton growths occur during the year: one in early spring in response to an influx of nutrients with runoff, and one in late summer as the water becomes warmer and metabolic rates of the plants increase (Reid and Wood, 1976). A spring pulse in phytoplankton concentration in the study reservoirs was not evident. For the

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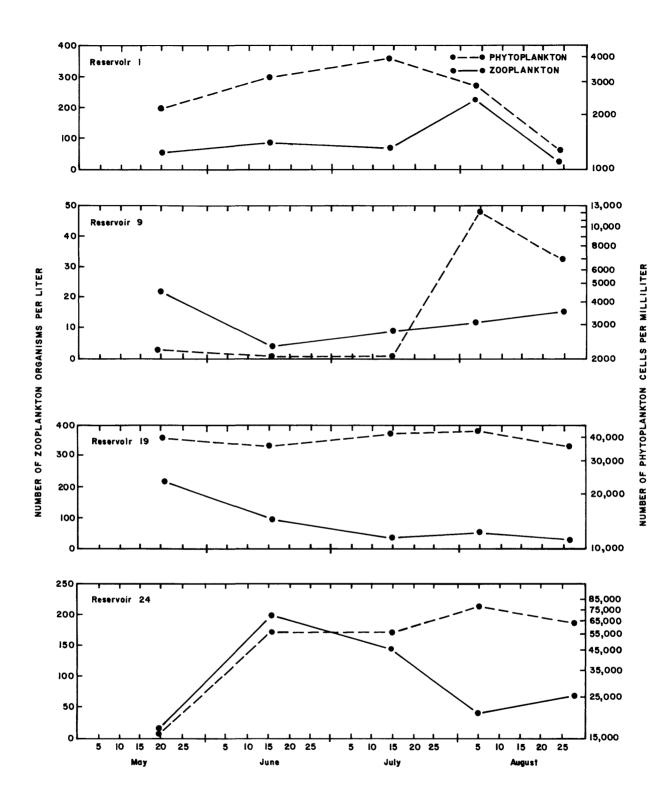


Figure 22.--Variation in the number of zooplankton organisms and phytoplankton cells at a depth of 1 meter in reservoirs 1, 9, 19, and 24 on five sampling dates (May through August).

sampling dates in this study, reservoirs 9, 19, and 24 had peak phytoplankton concentrations in early August (fig. 22). In reservoir 1 the peak cell concentration occurred in July.

Because phytoplankton cell concentrations are generally variable during spring and summer, lines connecting consecutive sampling points in figure 22 may not be representative of actual phytoplankton cell concentrations. However, among all the reservoirs, a common pattern for phytoplankton concentrations was a slight increase between the June and July sampling and a decrease from the early to late August sampling. The overall increase in phytoplankton concentration coincides with the seasonal increase in water temperatures. Increasing day length also probably contributes to increased phytoplankton growth until summer solstice. The decrease in day length after summer solstice probably contributes, in part, to the decrease in phytoplankton cell concentrations. Although there was no consistent trend in nutrient concentrations between the two August samplings, other factors causing a decrease in phytoplankton concentrations might include complexing reactions that render nutrients unavailable for uptake or zooplankton grazing.

Detected differences in phytoplankton concentrations among the study reservoirs coincide with the relative differences detected in dissolved oxygen and pH. Reservoir 9, which had the smallest dissolved oxygen and pH differences between the near-surface and near-bottom water and the largest dissolved-oxygen concentrations and pH values in near-bottom water, had relatively small phytoplankton concentrations. Reservoir 19, which had the largest differences in pH and dissolved oxygen between the near-surface and near-bottom water and the smallest dissolved-oxygen concentrations and pH values in the near-bottom water, had relatively large phytoplankton concentrations. These relative differences demonstrate the effect that phytoplankton can have on water quality. Large concentrations of phytoplankton can greatly increase the dissolved-oxygen concentration and pH in the near-surface water, but with increased production there is an attending large accumulation of organic matter in the bottom sediment that can create a large oxygen demand.

Zooplankton

Zooplankton concentrations in samples collected from the reservoirs are presented in table 8 (Supplemental Information section at back of report). Six zooplankton taxa were detected, with the largest percentages occurring in the orders Cladocera and Copepoda of the phylum Arthropoda. These orders generally are the most abundant zooplanktors in lakes (Reid and Wood, 1976). The May and June samples in reservoir 1 and the June sample in reservoir 24 were the only samples containing organisms in the order Ploima of the phylum Rotifera. Except for the occurrence of Daphnia ambigua in the June sample, zooplankton in reservoir 9 consisted of a single species--Cyclops bicuspidatus thomasi. The reason for the occurrence of a single species in reservoir 9 is not known. However, Cyclops bicuspidatus thomasi was present in every reservoir during each sampling. The Copepoda Diaptomus sp. was collected only in reservoirs 19 and 24. The species of zooplankton that exist in each reservoir could persist year after year as indicated in other studies (Reid and Wood, 1976).

Changes in total number of zooplankton organisms per liter (fig. 22) varied among the reservoirs. Reservoir 9 generally contained the smallest number of zooplankton organisms. In reservoirs 1, 19, and 24 an inverse relationship between number of phytoplankton and number of zooplankton is indicated (fig. 22). This relationship is strong from June to early August in reservoir 1, weak during the same period in reservoir 19, and strong from June to late August in reservoir 24. From May to June in all the reservoirs, zooplankton and phytoplankton indicate a direct relationship; however, total numbers of both plankton are increasing in reservoirs 1 and 24 and decreasing in reservoirs 9 and 19. These data indicate that the presence of phytoplankton as food does not constitute the sole limiting factor Several studies show this same lack of consistent correlation for zooplankton. between zooplankton and phytoplankton (Reid and Wood, 1976). Generally, zooplankton population cycles in the summer are variable, probably affected by a combination of changing food supply; shifts in the quality of food; and predation by zooplankton, fish, and other organisms such as insects (Wetzel, 1975). In addition. the sampling in this study might not have been frequent enough to show the true relationship that exists between phytoplankton and zooplankton. The relative numbers of each group may be different depending on whether the samples were collected during an increasing or decreasing phase of population change.

Benthic invertebrates

The most numerous classes of benthic invertebrates collected from the study reservoirs in late August were Oligochaeta (aquatic worms) and Insecta (insects) (table 9; Supplemental Information section at back of report). Organisms from the class Crustacea, particularly *Hyallela azteca* (scuds), were numerous in the shore sample from reservoir 24. Mainly because of the large number of Oligochaeta and Crustacea, reservoir 24 was the most productive in terms of total number of organisms. However, reservoir 24 was also the only study reservoir from which Mollusca was not collected. The most numerous organisms in reservoirs 1, 9, and 19 were Diptera (flies), of which Chironomus sp. generally comprised the largest percentage.

Except for reservoir 9, a large percentage of the bottom sediments was void of plant life except for epipelic algae. These areas are inhabited in large numbers by Oligochaeta and *Chironomus* sp., because of the abundant food supply (sediment containing organic matter of internal and external origins and colonized with bacteria and other microorganisms) and freedom from other competing benthic organisms. In contrast to most other benthic invertebrates, both of these organisms can tolerate small dissolved-oxygen concentrations and anaerobic conditions for short periods of time.

Hyallela azteca is a common freshwater species and an omnivorous substrate feeder that consumes bacteria, algae, and particulate detritus. Hyallela azteca reaches maximum production rates at temperatures between 20° and 25°C, which was similar to the range of temperatures in the reservoirs during sampling. It is more numerous along the shores because of the large quantities of available food and larger dissolved-oxygen concentrations of near-surface water compared to concentrations in the deeper locations of the reservoirs. Hyallela azteca can be a main source of food for fish populations.

Reservoirs 1, 19, and 24 had similar total numbers of benchic invertebrate taxa collected from all three sampling locations. Reservoir 9 had almost twice as many benchic invertebrate taxa as the other reservoirs. Because of the greater number of taxa, even though total numbers of organisms were fewer, reservoir 9 had a larger diversity index (table 2) and therefore a more stable community than reservoirs 1, 19, and 24. Part of this diversity is the result of a more diverse environment provided by the extensive growths of macrophytes along the reservoir

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bottom. Disregarding other factors, stable benthic invertebrate communities are more able to sustain fish populations than unstable communities.

Table 2.--Diversity index of benthic invertebrates collected from each reservoir during August 1981

[Calculations based on combined numbers of individuals and taxa from all three sampling locations in each reservoir]

	Divers	ity index
Reservoir	Brillouin ¹	Shannon-Weaver ²
1	1.13	1.65
9	2.13	3.09
19	1.30	1.89
24	1.22	1.77

¹Diversity index (from Brillouin, 1962): $H = \frac{1}{N} \log \frac{N!}{N_1!N_2!\cdots N_s!}$

²Diversity index (from Wilhm and Dorris, 1968): $H = -\sum_{i=1}^{s} \frac{N_i}{N} \log \frac{N_i}{N}$

where in both equations H is Diversity Index (information per individual), N is the total number of individuals, s is the total number of taxa, and N_i (i=1,2,...s) is the number of individuals in the ith species.

Fecal bacteria

Reservoirs 19 and 24 generally had larger fecal coliform and fecal streptococcal bacteria concentrations than reservoirs 1 and 9 (table 10; Supplemental Information section at back of report). There were no consistent trends in bacterial concentrations among the study reservoirs, nor were there consistent differences between shore and midpoint bacterial concentrations. The ratio between the fecal coliform and fecal streptococcal concentrations generally was less than 1.0 in reservoirs 1 and 9 and greater than 1.0 in reservoirs 19 and 24. However, interpretation of the data is difficult because of nonideal colony counts. Most of the bacteria presumably originated from livestock and waterfowl waste, which can be a variable nonpoint source of contamination. Although cattle were observed only at reservoir 9, cattle waste was present in drainages of the other reservoirs. Studies indicate that 100-day-old cattle wastes can release fecal coliform in concentrations that exceed recreational water-quality criteria (Kress and Gifford, 1984). In addition, waterfowl were observed at least once in each of the reservoirs.

DIEL TRENDS

Onsite measurements

At measurement depth of 1 meter

Water temperature, measured dissolved-oxygen concentrations, and calculated dissolved-oxygen concentrations at saturation are plotted in figure 23. Values of pH and specific conductance are plotted in figure 24.

Temperature

Water temperatures measured at a depth of 1 m followed trends in ambient air temperatures, with diel fluctuations having a lag time of several hours. The diel trends of water temperature are similar among the four reservoirs, with maximum temperatures occurring during mid- to late afternoon (1500-1800 hours) and minimum temperatures occurring during mid-morning (0900-1000 hours). Temperature fluctuation throughout the day was small, averaging 2.1 C° for the four reservoirs. The maximum temperature measured at a depth of 1 m was 22.9°C in reservoir 1.

Late August was chosen for the diel sampling under the assumption that limnological conditions would be most stressful for aquatic biota. Proposed reservoir uses other than fish production, in general, would not be expected to be affected by diel changes in water quality. Reservoir temperatures at a depth of 1 m did not exceed 23°C, which is less than the maximum range of 23.9°-33.9°C recommended for successful spawning and growth of fish species such as largemouth bass, catfish, buffalo, bluegill, and perch (National Technical Advisory Committee to the Secretary of the Interior, 1968). However, temperatures greater than 20°C for prolonged periods can be lethal to species such as trout and northern pike.

Dissolved oxygen

Calculated dissolved oxygen at saturation showed little change throughout the day in each reservoir, in contrast to measured concentrations of dissolved oxygen, which showed pronounced fluctuations during 24 hours. Maximum dissolved-oxygen concentrations measured in the reservoirs generally occurred late in the evening (1800-2000 hours) just prior to sunset. A gradual decrease in dissolved oxygen continued through the night and generally reached a minimum in reservoirs 9, 19, and 24 from 0700 to 0800 hours. Problems with the continuous recorder at reservoir 1 limited dissolved-oxygen data to measurements about every 3 hours with the multiparameter instrument. The lowest dissolved-oxygen measurement at this site was obtained at 0200 hours.

Although dissolved-oxygen trends at a depth of 1 m were similar among the reservoirs, concentrations varied considerably. Reservoirs 1 and 9 had oxygen maxima at concentrations considerably greater than saturation. Measured dissolvedoxygen concentrations at a depth of 1 m ranged from 5.0 to 9.5 mg/L in reservoir 1 and from 6.0 to 10.2 mg/L in reservoir 9. Of all the reservoirs, reservoir 9 had the largest diel oxygen maximum and the most symmetrical pattern relative to saturation. Reservoir 19 was unique among the four reservoirs in that measured values of dissolved oxygen were always less than saturation, ranging from 3.2 to 6.8 mg/L. Reservoir 24 had the smallest diel dissolved-oxygen minimum; however, its

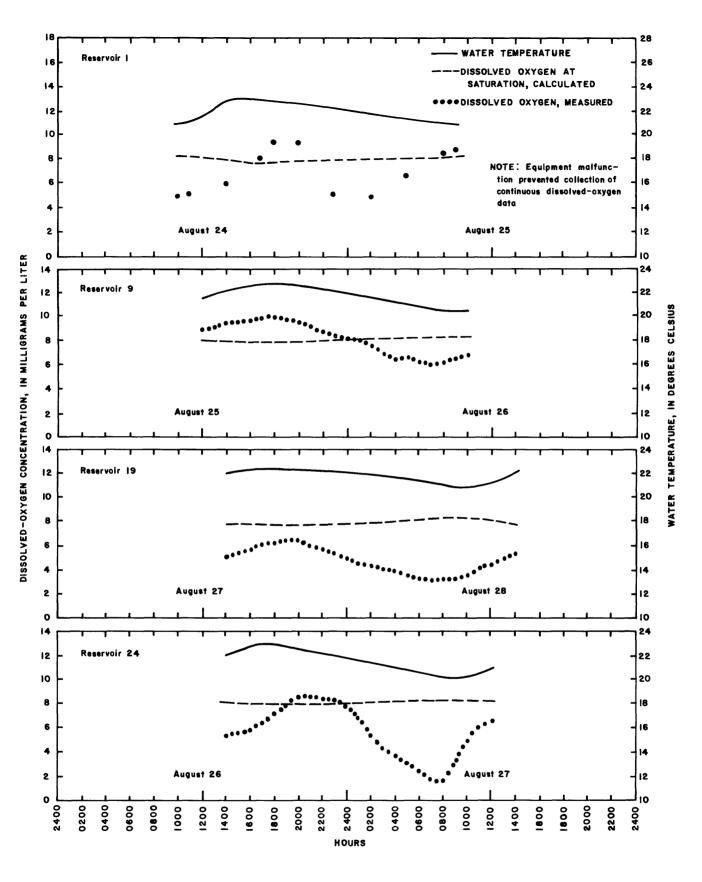


Figure 23.--Variation of water temperature and dissolved-oxygen concentration at a depth of 1 meter in reservoirs 1, 9, 19, and 24 during 24 hours in late August.

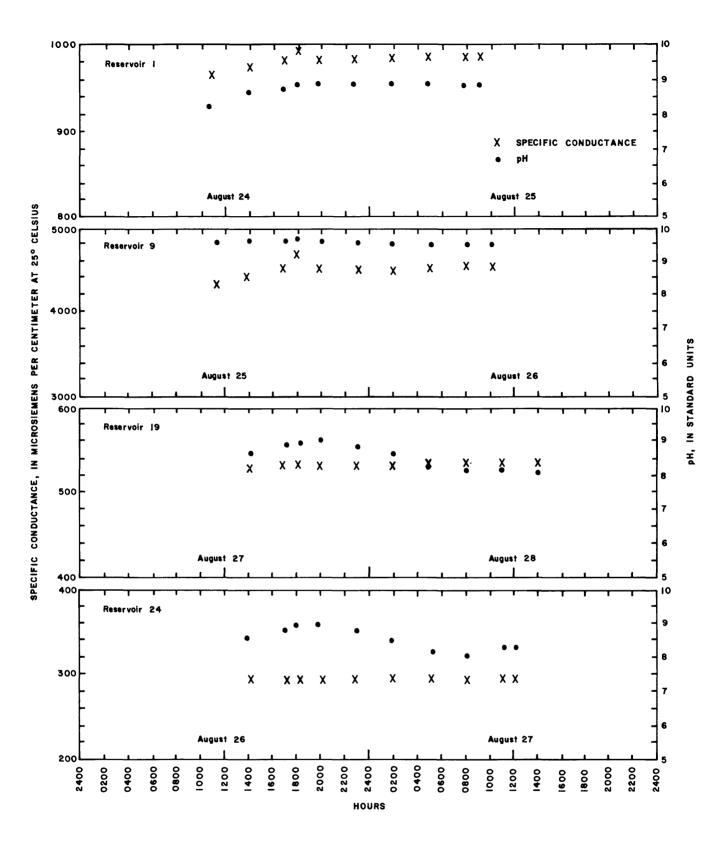


Figure 24.--Variation of pH and specific conductance at a depth of 1 meter in reservoirs 1, 9, 19, and 24 during 24 hours in late August.

dissolved-oxygen maximum exceeded saturation, producing the largest diel fluctuation (1.8-8.9 mg/L).

Diel patterns at the 1.0-m depth result from dissolved oxygen produced by photosynthesis during daylight hours surpassing the oxygen demand of respiration and decomposition, and accumulating to a maximum between 1800 and 2000 hours. As sunlight decreases, the rate of photosynthetic oxygen production decreases and finally stops, beginning a trend toward oxygen depletion. Cellular respiration and bacterial decomposition of organic matter consume oxygen, creating an oxygen minimum between 0700 and 0800 hours in each reservoir except reservoir 1. In reservoir 1, oxygen concentrations began to increase at 0200 hours, presumably in response to strong winds that caused mixing of aerated surface water with water at the 1.0-m depth.

The undersaturation of oxygen maxima in reservoir 19 may have been due to greater rates of decomposition assumed to occur in the thick bottom muds of this reservoir. Reservoirs 1 and 9 had larger dissolved-oxygen concentrations during the daylight hours than reservoirs 19 and 24, possibly because of additional production by the dense growths of aquatic macrophytes along the shore and reservoir bottom. Reservoir 24, with the largest range of diel fluctuation, was very turbid and had a noticeable humic-brown (organic) coloration. The presence of a large phytoplankton population in reservoir 24, coupled with the presence of large quantities of organic matter in the bottom sediment, could explain the considerable variations in dissolved-oxygen concentrations.

Small dissolved-oxygen concentrations affect fish production more than the other proposed reservoir uses. Within existing ranges of the study reservoirs, dissolved-oxygen availability imposes greater restrictions on aquatic biota than temperature. In fact, a major problem with increased temperature in water is that it decreases the solubility of dissolved oxygen while concomitantly leading to increased rates of oxygen consumption through accelerated metabolic demands and bacterial decomposition. Although the increased rate of metabolism may result in a greater production of oxygen through photosynthesis during the day, it may also result in greater deficits at night, and prolonged oxygen deficits substantially less than saturation during the night can severely stress desirable forms of aquatic organisms. A minimum concentration of 5.0 mg/L has been recommended by the U.S. Environmental Protection Agency (1978) to protect freshwater aquatic life and to support a diverse fish population. Reservoirs 1 and 9 sustained oxygen levels equal to or greater than 5.0 mg/L throughout the 24 hours at the 1.0-m depth, which would indicate suitable conditions for fish propagation. Dissolved-oxygen concentrations in reservoir 19 decreased to less than 5.0 mg/L for an extended time. Reservoir 24, although maintaining oxygen concentrations in excess of 5.0 mg/L for much of the day, decreased to 1.8 mg/L during the early morning hours. Dissolved-oxygen concentrations measured at the 1.0-m depth in reservoirs 19 and 24 probably would be considered marginal for supporting viable populations of fish and other oxygen-sensitive aquatic organisms.

pН

Diel fluctuations of pH at the 1.0-m depth were less than 1.0 standard unit in all four reservoirs (fig. 24). Maximum pH values occurred in late afternoon and generally corresponded to the time of maximum dissolved-oxygen concentrations. Reservoirs 1 and 9 had the least change in pH throughout the day, with ranges of 8.2 to 8.8 in reservoir 1 and 9.4 to 9.7 in reservoir 9. The pH changed from 8.2 to 9.1 in reservoir 19 and from 8.0 to 8.9 in reservoir 24. Changes in pH are related to the uptake and release of CO_2 through primary production by phytoplankton and respiration and decomposition of all aquatic organisms.

Diel values of pH measured at the 1.0-m depth generally were within limits suitable for most freshwater aquatic life, waterfowl, and livestock, except during late afternoon. Although aquatic organisms can become acclimated to temporary exposure outside protective ranges, fishery production generally deteriorates as pH values diverge from the recommended range. In all the reservoirs, the maximum pH values in late afternoon either were near or exceeded the criteria protective of fish, waterfowl, and livestock. At the 1.0-m depth, reservoir 9 continuously exceeded and reservoir 19 temporarily exceeded the pH criterion of 9.0, whereas reservoirs 1 and 24 were very close to the limit. The pH in all reservoirs exceeded the criterion protective of recreational swimming.

Specific conductance

Diel fluctuations of specific conductance at the 1.0-m depth (fig. 24) in the reservoirs were minimal, with no evident trends. Reservoir 9 had the largest diel fluctuation and also the largest values, ranging from 4,300 to 4,620 μ S/cm. The smallest values and smallest fluctuation, 290 to 294 μ S/cm, occurred at reservoir 24. Specific-conductance values fluctuated from 965 to 991 μ S/cm at reservoir 1 and from 530 to 542 μ S/cm at reservoir 19. The relatively small variation in pH values at the 1.0-m depth limits diel changes in dissolution-precipitation reactions that might otherwise affect specific conductance.

Profiles

Depth-profile measurements of water temperature, dissolved oxygen, pH, and specific conductance were made at 1800 (presunset) and 0500 (predawn) hours. These measurements were made to assist in the detection of diel water-quality changes.

Temperature

Profiles of water temperature in each reservoir (figs. 25-28) indicate lower temperatures in the near-surface water at 0500 hours compared to 1800 hours. Near-surface water temperatures at 0500 hours ranged from 20.7 °C in reservoir 24 to 21.9 °C in reservoir 19. Near-surface water temperatures at 1800 hours ranged from 22.5 °C in reservoir 19 to 24.0 °C in reservoir 9. Generally, there was little difference in the temperature of near-bottom water of each reservoir between the two times.

Temperature gradations from near-surface to near-bottom water in each reservoir were not maintained between the two profile times. Temperature differences between the near-surface and near-bottom waters at 1800 hours ranged from 2.1C° at reservoir 24 to 2.8C° at reservoirs 1 and 9. In reservoirs 9 and 24 following night-long heat loss from the water surface, the near-surface water had cooled to about the temperature of the near-bottom water.

During periods of solar radiation, water temperatures in reservoirs 9 and 24 increased throughout the entire depth. In reservoirs 1 and 19, which are deeper

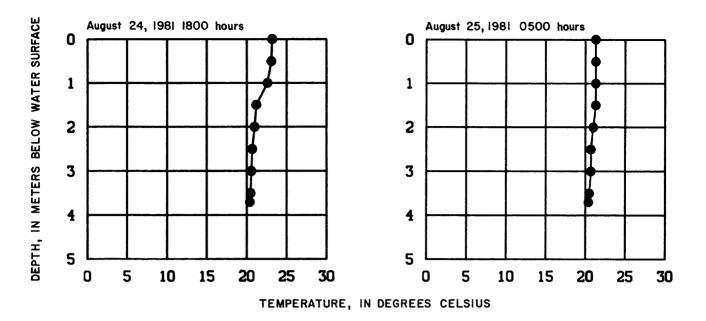


Figure 25.--Evening and morning depth profiles of temperature in late August in reservoir 1.

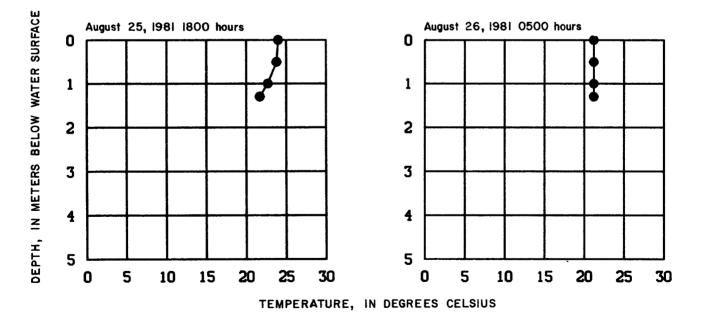


Figure 26.--Evening and morning depth profiles of temperature in late August in reservoir 9.

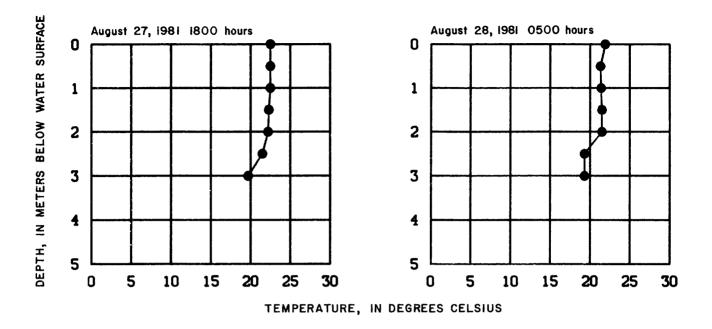


Figure 27.--Evening and morning depth profiles of temperature in late August in reservoir 19.

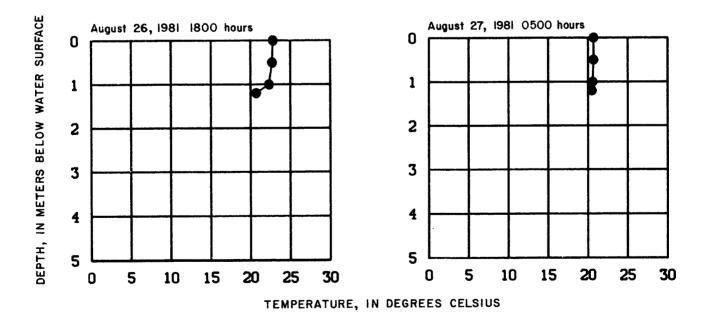


Figure 28.--Evening and morning depth profiles of temperature in late August in reservoir 24.

than reservoirs 9 and 24, warming and cooling also occurred but were restricted primarily to the upper 2.0 m. Because of the greater heat capacity per square unit of surface area in the deeper reservoirs, the available solar radiation was not enough to cause a temperature change in the near-bottom water of the deeper reservoirs. Consequently, reservoirs 1 and 19 maintained constant near-bottom temperatures between 0500 and 1800 hours.

Dissolved oxygen

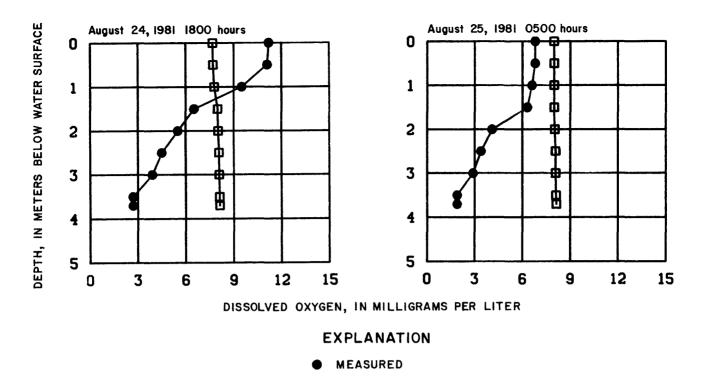
Depth profiles of dissolved oxygen (figs. 29-32) illustrate larger concentrations of dissolved oxygen throughout the water column at 1800 hours than at 0500 hours. Except in reservoir 9, the oxygen maxima in the reservoirs occurred near the surface and the minima occurred near the bottom. In reservoir 9, the oxygen maximum measured at 1800 hours occurred near middepth at 1.0 m.

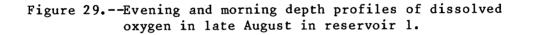
The largest differences between maximum and minimum dissolved-oxygen concentrations within the water column of each reservoir occurred at 1800 hours. Of the four reservoirs, reservoir 1 had the largest difference at 1800 hours, with dissolved-oxygen concentrations ranging from 11.2 mg/L near the surface to 2.7 mg/L near the bottom. The smallest difference at 1800 hours occurred in reservoir 9 with dissolved-oxygen concentrations ranging from 10.1 to 8.6 mg/L. Although differences between maximum and minimum dissolved-oxygen concentrations at 1800 hours were similar in reservoirs 19 and 24, concentrations in reservoir 19 (6.8-0.3 mg/L) were considerably smaller than in reservoir 24 (9.9-3.5 mg/L).

At 0500 hours, reservoir 1 again had the largest difference in dissolved-oxygen concentrations, with concentrations varying from 6.8 mg/L near the surface to 1.9 mg/L near the bottom. The smallest difference at 0500 hours was measured in reservoir 24 (2.7-2.3 mg/L). Reservoir 9 maintained the largest dissolved-oxygen concentrations (7.4-6.6 mg/L) throughout the water column at 0500 hours, whereas dissolved-oxygen concentrations in reservoir 19 decreased from 3.7 mg/L near the surface to the smallest measured oxygen minimum of 0.1 mg/L near the bottom.

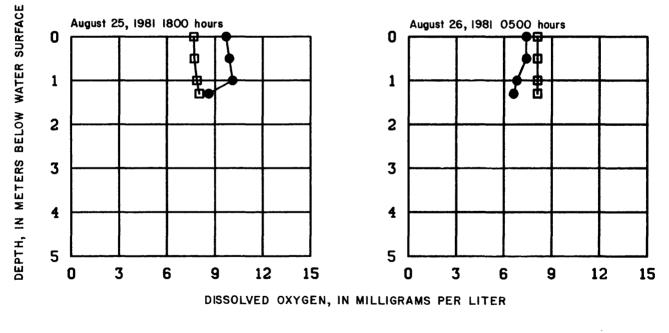
Dissolved-oxygen concentrations with respect to saturation varied considerably between 1800 and 0500 hours. All reservoirs except reservoir 19 had oxygen concentrations in excess of saturation in the near surface at 1800 hours and all had less than saturation throughout their water columns at 0500 hours. In reservoir 19, oxygen concentrations never exceeded saturation near the surface and decreased to less than 4 percent of saturation near the bottom during both sampling hours. Reservoir 9 maintained supersaturated oxygen conditions through its entire depth at 1800 hours.

Differences determined between the 1800- and 0500-hour dissolved-oxygen profiles for each reservoir are caused by oxygen production during photosynthesis and oxygen utilization during respiration and decomposition of aquatic organisms. Because phytoplankton are more numerous in the upper part of the water column, dissolved-oxygen concentrations greater than saturation will occur there during the evening (1800 hours) following day-long oxygen production. Daytime and nighttime respiration and decomposition throughout the water column virtually occur at the same rate so that the net effect is a larger range of dissolved-oxygen concentrations at 1800 hours than at 0500 hours.





AT SATURATION

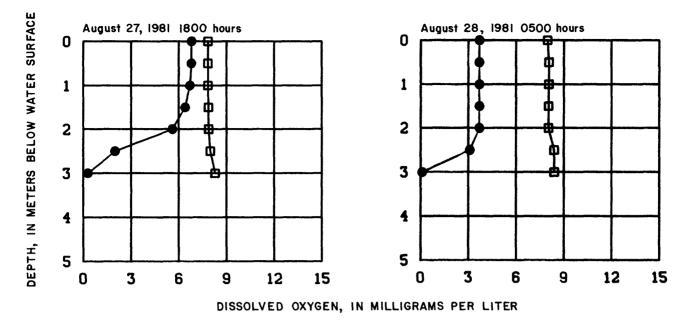


EXPLANATION

MEASURED

AT SATURATION

Figure 30.--Evening and morning depth profiles of dissolved oxygen in late August in reservoir 9.

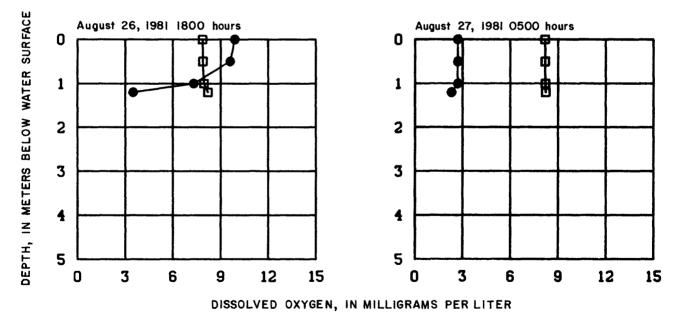


EXPLANATION

• MEASURED

AT SATURATION

Figure 31.--Evening and morning depth profiles of dissolved oxygen in late August in reservoir 19.



EXPLANATION

- MEASURED
- AT SATURATION

Figure 32.--Evening and morning depth profiles of dissolved oxygen in late August in reservoir 24.

Differences in dissolved-oxygen profiles among the reservoirs basically result from differences in abundance and distribution of aquatic plants (algae and macrophytes). Submersed macrophytes growing from the bottom of reservoir 9 to middepth could be the cause of the oxygen maximum at middepth. In addition, macrophyte growth along the bottom of reservoir 9 would produce oxygen in the lower column of water and result in the small change in dissolved-oxygen concentration with depth.

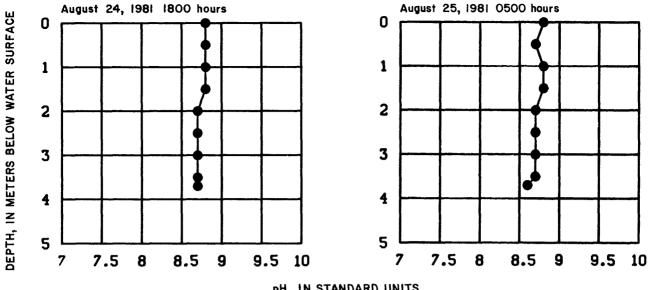
Reservoir 1 had relatively small nutrient and phytoplankton concentrations compared to the other reservoirs. However, being the second-oldest reservoir would presumably account for greater accumulations of organic sediment compared to the younger reservoirs. The large difference in dissolved-oxygen concentrations with depth in reservoir 1, in part, is due to increased decomposition rates of this accumulated organic matter and increased production rates by phytoplankton and dense macrophytes along shore.

Large respiration and decomposition rates relative to oxygen production could account for undersaturated dissolved-oxygen concentrations in reservoir 19 at 1800 and 0500 hours and the small dissolved-oxygen concentrations in the near-surface water of reservoir 24 at 0500 hours. Large production rates compared to reservoirs 1 and 9 are indicated in reservoirs 19 and 24 by their large phytoplankton concentrations. However, with these increased production rates there can be attendant large populations of consumers in higher trophic levels and dieoffs of algal cells after blooms. With large numbers of organisms there is increased respiration and increased organic matter available for decomposition. In addition, cattle wastes could be another source of decomposable organic material. Such increased respiration and accumulation of organics would produce a large oxygen demand, which could keep the dissolved-oxygen concentration at less than saturation.

Under conditions such as the large dissolved-oxygen gradients in reservoir 1, mobile organisms such as fish can migrate to upper waters where dissolved-oxygen concentrations are more favorable. However, bottom-dwelling invertebrates are restricted to the deeper oxygen-deficient waters and may become stressed. Reservoir 9, which either exceeded or was near saturation throughout its depth, had oxygen concentrations suitable for many warm-water organisms. Reservoir 19, which did not reach dissolved-oxygen saturation near the surface during either profile, approached anaerobic conditions in the near-bottom water. It is unlikely that the limited oxygen availability of this reservoir during late summer could support viable populations of oxygen-sensitive organisms. Reservoir 24 maintained relatively large oxygen concentrations within the upper 1.0 m of water at 1800 hours. However, very small concentrations persisted throughout the entire depth at 0500 hours, which would severely stress or inhibit oxygen-sensitive organisms.

pH

Depth profiles of pH during diel sampling (figs. 33-36) showed little change with depth or time. Values of pH at 1800 hours were only slightly larger than those at 0500 hours. Variation in pH with depth was less than 1.0 standard unit in each reservoir for both sampling hours. Maximum values of pH occurred near the surface and minimum values occurred near the bottom. The maximum near-surface water pH was 9.8 at 1800 hours in reservoir 9 and the minimum near-bottom water pH was 7.8 at 0500 hours in reservoir 19.



pH, IN STANDARD UNITS

Figure 33.--Evening and morning depth profiles of pH in late August in reservoir 1.

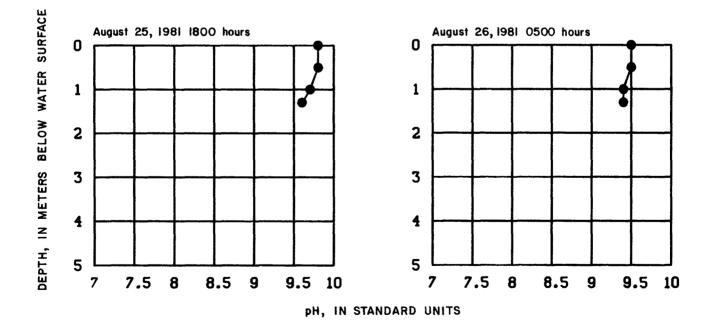


Figure 34.--Evening and morning depth profiles of pH in late August in reservoir 9.

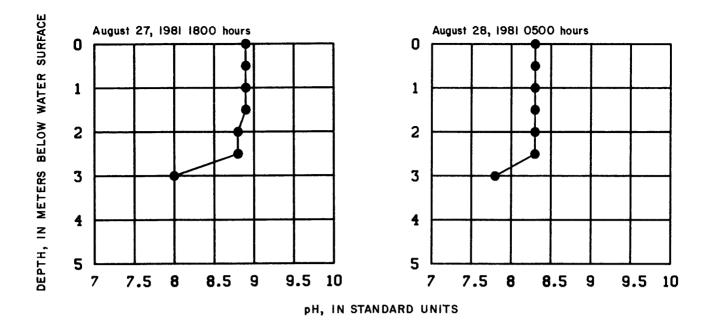


Figure 35.--Evening and morning depth profiles of pH in late August in reservoir 19.

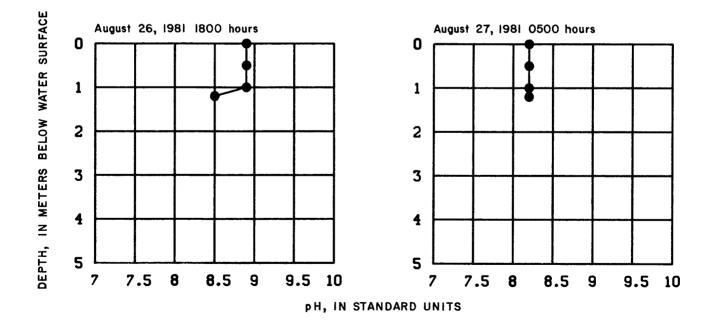


Figure 36.--Evening and morning depth profiles of pH in late August in reservoir 24.

Profiles of pH revealed no major diel fluctuations or variation with depth, indicating that the effect of photosynthesis and respiration on pH was sufficiently buffered. Values of pH remained relatively large throughout the entire depths of each reservoir and probably did not significantly increase the solubility of metal compounds in bottom sediments during the night.

Specific conductance

Specific-conductance profiles during diel sampling (figs. 37-40) showed little variation with depth or time. The difference in values from near-surface to near-bottom water was slightly larger at 1800 hours than at 0500 hours. The maximum range measured in the reservoirs was $40 \ \mu S/cm$ at 1800 hours in reservoir 9.

Chemical analyses

Major dissolved constituents

Generally, concentrations of major dissolved constituents (table 4) showed little change between the two sampling hours. In addition, only minor differences in concentrations were detected between samples collected at depths of 1.0 and 3.0 m in reservoirs 1 and 19.

The absence of any major differences between the 1800- and 0500-hour samples for major dissolved constituents, nutrients, and trace elements indicates negligible fluctuations in chemical composition on a diel basis. Major dissolved constituents would not be expected to vary significantly during 24 hours in shallow reservoirs where pH is relatively stable and wind can cause complete mixing.

Nutrients

Nutrient samples collected for analysis of nitrogen, phosphorus, and carbon concentrations (table 5) generally indicated only minor differences between the 1800- and 0500-hour sampling periods and between the depths of 1.0 and 3.0 m in reservoirs 1 and 19. Total organic carbon had the most consistent variation among the reservoirs, with increased concentrations at 0500 hours compared to 1800 hours in reservoirs 1, 19, and 24. In general, nutrient concentrations were slightly more variable than major dissolved constituents as a result of biological uptake and decay; however, differences were not consistent enough at all reservoirs to define trends.

Trace elements

Apparent diel variation in trace-element concentrations of lead and manganese occurred in the near-bottom samples of reservoirs 1 and 19 (table 6). In reservoir 1, increased total recoverable lead concentrations, from 5 μ g/L (micrograms per liter) at 1800 hours to 39 μ g/L at 0500 hours, may have been caused by bottom sediments entering the sample. Bottom-sediment contamination is probable because the dissolved-lead concentration did not increase. In reservoir 19, both dissolved and total recoverable manganese concentrations in the near-bottom samples were larger at 1800 hours than at 0500 hours. As noted by visual observation, the sediments in reservoir 19 were thick and dark, presumably containing large quantities of organic

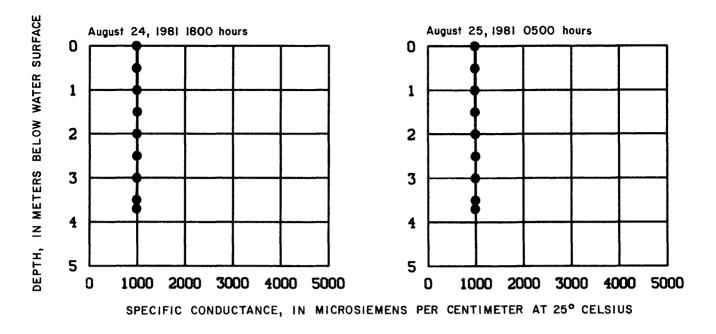


Figure 37.--Evening and morning depth profiles of specific conductance in late August in reservoir 1.

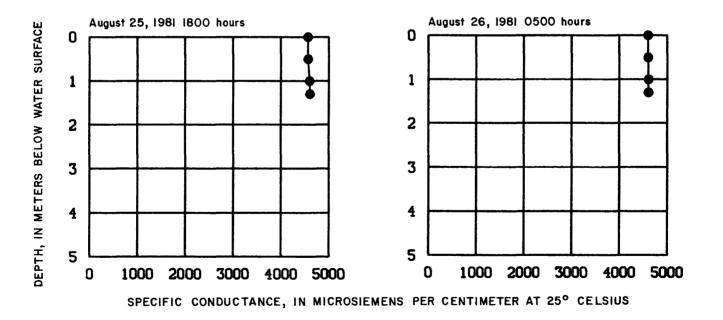


Figure 38.--Evening and morning depth profiles of specific conductance in late August in reservoir 9.

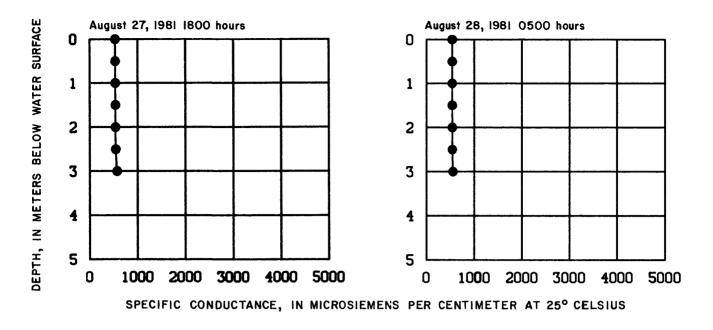


Figure 39.--Evening and morning depth profiles of specific conductance in late August in reservoir 19.

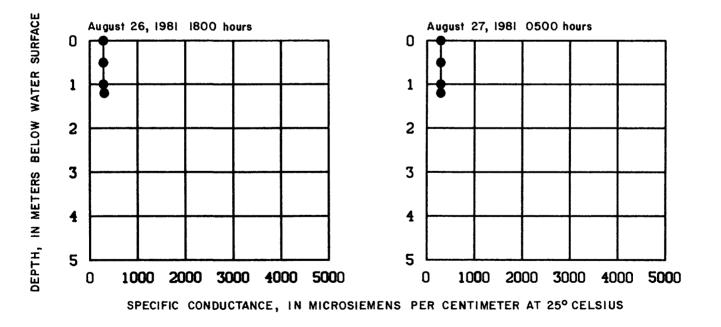


Figure 40.--Evening and morning depth profiles of specific conductance in late August in reservoir 24.

matter. Such organic matter commonly is enriched in manganese, which can be released through reduction and bacterial action as dissolved oxygen and pH decrease during the night (figs. 23 and 24) (Hem, 1970). Because larger manganese concentrations occurred at 1800 hours rather than at 0500 hours, sediment contamination in the samples from reservoir 19 also is probable.

Biological analyses

Concentrations of chlorophyll a and b (table 3) showed moderate and inconsistent fluctuations between the 1800- and 0500-hour sampling periods and between the 1.0- and 3.0-m sampling depths in reservoirs 1 and 19. Only one analysis was available for reservoir 9, which precludes comparison between sampling times. Chlorophyll a and b concentrations were significantly larger in reservoirs 19 and 24 than in reservoirs 1 and 9. Chlorophyll b concentrations, although much smaller than chlorophyll a, generally followed the same patterns in each reservoir.

Reservoir	Date	Time (24 - hour)	Sampling depth (meters)	Concentration, in the Chlorophyll a	micrograms per liter Chlorophyll b
1	08-24-81	1802	1.0	15.5	1.41
1	08-24-81	1806	3.0	1.87	.305
1	08-25-81	0502	1.0	6.08	.526
1	08-25-81	0506	3.0	2.26	.270
9	08-25-81	1802	1.0	6.62	.217
19	08-27-81	1802	1.0	42.1	10.7
19	08-27-81	1806	3.0	80.5	16.0
19	08-28-81	0502	1.0	75.9	15.4
19	08-28-81	0506	3.0	52.8	11.9
24	08-26-81	1802	1.0	272	46.4
24	08-27-81	0502	1.0	150	27.4

Table 3.--Concentrations of chlorophyll a and b collected during the diel sampling

Because phytoplankton are passive and move with water currents, their distribution in a shallow reservoir can be extremely variable. However, many viable phytoplankton cells would be expected to occur in the upper water where sunlight conditions are more favorable for photosynthesis. This upper water occurrence may account for the generally larger chlorophyll concentrations measured in the nearsurface samples of reservoirs 1 and 19. The differences in chlorophyll concentrations between 1800 and 0500 hours are variable and may occur because of uneven mixing due to temperature changes.

Chlorophyll a concentration can be considered as one manifestation of nutrient enrichment in the process of eutrophication. Taylor and others (1980) list a criterion of greater than 10.0 μ g/L for chlorophyll a as indicative of eutrophic conditions in lakes not dominated by macrophytes. With the exception of reservoir

9, which contained abundant macrophytes, the study reservoirs all contained at least one sample exceeding 10.0 μ g/L. Chlorophyll a concentration in reservoirs 19 and 24 greatly exceeded 10 μ g/L in all samples, whereas reservoir 1 had concentrations much less than 10 μ g/L in all samples except 1. Therefore, chlorophyll a concentrations indicate that reservoirs 19 and 24 are probably in more advanced stages of eutrophication than reservoir 1.

CONCLUSIONS

Water quality of the study reservoirs is affected by regional and local mineralogy, physical characteristics, and land use. Reservoirs 19 and 24 have drainages underlain entirely by the Bearpaw Shale and therefore are affected by the same regional mineralogy. These two reservoirs have larger manganese concentrations, smaller dissolved-solids concentrations, greater turbidity, and larger nitrogen (total nitrogen and total ammonia) concentrations than reservoirs 1 and 9. The larger nutrient concentrations of reservoirs 19 and 24 result in larger phytoplankton and chlorophyll a and b concentrations compared to reservoirs 1 and 9. However, the reservoirs do not show the same paired comparisons when grouped by water type, presumably as a result of differences in local mineralogy. Reservoir 19 has sodium bicarbonate sulfate water and reservoir 24, like reservoir 1, has sodium bicarbonate water. Reservoir 9 has sodium sulfate water.

Differences in water quality caused by mineralogy and land use mask differences caused by reservoir age. Virtually closed reservoirs, such as the study reservoirs, generally would attain large concentrations of water-quality constituents through years of continued water loss by evapotranspiration. However in the study area, dissolved-solids, phosphorus, and total organic carbon concentrations in the younger reservoirs (reservoirs 9 and 24) are both smaller and larger than in the older reservoirs (reservoirs 1 and 19). Consequently, no clear relationship between age and stage of eutrophication can be shown.

Trends from May to August that are similar among all the reservoirs include relatively uniform water temperature increases at all depths, increased differences between near-surface and near-bottom water dissolved-oxygen concentrations and pH, and an increase in concentration of dissolved solids. The uniform increase in water temperature at all depths is possible because the shallowness of the reservoirs allows frequent mixing. The progressive increase in the difference between nearsurface and near-bottom water for dissolved-oxygen concentrations and pH results primarily from differing ratios of photosynthesis, respiration, and decomposition. The seasonal increase in the concentration of dissolved solids most likely is due to cumulative water losses by evapotranspiration.

The study reservoirs are considered to be enriched with nutrients. Consequently, they have the potential to support large concentrations of phytoplankton. Generally, with large concentrations of phytoplankton, daytime dissolved-oxygen concentrations and pH values are large. Among all the reservoirs, inconsistent monthly fluctuations in composition and concentration of phytoplankton result in variable dissolved-oxygen concentrations and pH values. However, reservoir 1, which had one of the smaller phytoplankton and chlorophyll a and b concentrations, had one of the larger dissolved-oxygen concentrations and pH values; conversely, reservoir 24, which had the largest phytoplankton and chlorophyll a and b concentrations did not have the largest dissolved-oxygen concentrations and pH values. This dichotomy demonstrates that phytoplankton production is not the sole biological factor affecting dissolved oxygen and pH, but works in conjunction with other factors-principally respiration by all aquatic organisms and decomposition.

Although all reservoirs contained similar types of planktonic organisms, they were present in different proportions. Phytoplankton-cell concentrations were largest in reservoir 24 and smallest in reservoirs 1 and 9. However, reservoirs 1 and 9 supported large growths of macrophytes. Monthly changes in zooplankton concentrations varied among the reservoirs. There was no consistent correlation between the concentration of zooplankton and the concentration of phytoplankton-their food source.

Benthic invertebrates in bottom sediments of each reservoir were composed of large numbers of Oligochaeta and Chironomus sp. These organisms feed on large food supplies of bacteria and other micro-organisms colonized on the sediment. Hyallela azteca was common along the shore, particularly in reservoir 24. Bacteria, algae, and particulate detritus provide abundant food for these organisms. Although the total number of organisms were fewer in reservoir 9 than in reservoirs 1, 19, and 24, reservoir 9 had a stable, diverse benthic community.

There were no consistent monthly trends in fecal bacteria concentration among the study reservoirs, nor were there consistent differences between shore and midpoint bacterial concentrations. Most of the fecal bacteria existing in the reservoirs presumably originated from livestock and waterfowl waste, which can be a variable nonpoint source of contamination.

The reservoirs are subject to frequent mixing by moderate winds because of their shallow depths and diminutive thermal stratification. Mixing prevents a sustained decrease of dissolved-oxygen concentrations in the near-bottom water. However, mixing also recirculates nutrients from the bottom sediments, enhancing growth of phytoplankton and production of organic matter for decomposition.

Major water-quality changes in the study reservoirs that could affect reservoir use were: an increase in selected trace-element concentrations from May to August, a decrease in dissolved-oxygen concentration during the night, and an increase in pH during the day. Fish propagation is more affected by these changes than are the other proposed reservoir uses of waterfowl habitat, livestock watering, and recreational swimming. Because of the concentrating effect of water loss through evaporation, August concentrations of total recoverable lead in reservoirs 19 and 24 exceeded by the greatest concentration the criterion protective of fish. Owing to large oxygen consumption by respiration and decomposition, nighttime dissolved-oxygen concentrations in reservoirs 19 and 24 were less than the criterion protective of fish. In contrast, large photosynthetic rates during the day resulted in all reservoirs approaching or exceeding the maximum limit of pH protective of all proposed reservoir uses in the late afternoon.

Because similar small reservoirs in eastern Montana evaluated during an earlier study have large nutrient concentrations, they also have the potential to support large phytoplankton concentrations, which could result in small dissolved-oxygen concentrations at night. Diel sampling in other reservoirs considered for fish propagation would delineate reservoirs in which water quality during the day is protective of fish but during the night attains stressful dissolved-oxygen concentrations.

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SUPPLEMENTAL INFORMATION

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Table 4.--Major dissolved chemical constituents

[Analyses by U.S. Geological Survey. Abbreviations: m, meter; mg/L, milligram per liter]

Date	Time (24- hour)	Sam- pling depth (m)	Hard- ness (mg/L as CaCO ₃)	bonate (mg/L as	solve (mg/L	dis- d solved (mg/L	, Sodium dis- d solved (mg/L	-	
			RESE	RVOIR 1	AIR BAS	SE POND			
May, 1981 20 20	0832 0836	1.0 3.0	230 230	0.00	33 33	35 35	140 140	57 57	4.0 4.0
June 15 15	1503 1507	1.0 3.0	220 220	.00 .00	27 27	36 37	140 130	58 56	4.2 3.8
July 14 14	1547 1551	1.0 3.0	200 200	.00 .00	22 21	35 35	150 150	61 62	4.6 4.7
Aug 04 24 24 25 25	1603 1607 1802 1806 0502 0506	1.0 3.0 1.0 3.0 1.0 3.0	200 200 200 210 210 200	.00 .00 .00 .00 .00	20 21 21 22 21 21	36 36 37 38 37	150 150 150 160 160 160	61 61 62 62 62	5.5 5.5 5.7 5.7 5.8
Date	Sa pli dep (m	m- di: ng solv th (mg	as- li um, la s- t ved (m /L	bora- ory	Sulfate, dis- solved (mg/L as S04)	Chlo- ride, dis- solved (mg/L as Cl)	ride, dis-		Solids, sum of constit- uents, dis- solved (mg/L)
May, 1981 20 20	1. 3.			330 330	160 170	12 12	0.3	2.0 2.0	586 597
June 15 15	1. 3.			310 300	180 180	19 19	.3 .3	2.7 2.8	597 582
July 14 14	1. 3.			300 300	180 190	21 21	•3 •3	2.1 2.3	596 605
Aug 04 24 24 25 25	1. 3. 1. 3. 1. 3.	0 5.1 0 6.2 0 6.0 0 6.1	5 2) 5	360 360 350 350 310 350	160 160 140 150 	6.3 7.1 20 20 20 23	.5 .5 .4 .4 .4 .4	2.3 2.9 2.9 3.3 3.0 3.6	597 600 587 609 623

Date	Time (24– hour)	Sam- pling depth (m)	Hard- ness (mg/L as CaCO3	bonat (mg) as	s, ar- Calci te dis /L solv s (mg/2	- dis ed solve L (mg/]	m, Sodi - dis ed solv L (mg/	s- ved /L Perce	
				RESERVO	DIR 9GA	Y RESERVO	DIR		
May, 1981 20	1403	1.0	1,000	92	20 200	130	50	00 51	6.8
June 16	0903	1.0	950	87	70 200	110	51	0 53	7.2
July 15	0903	1.0	980	94	40 180	130	62	20 57	8.6
Aug 05 25 26	0932 1802 0502	1.0 1.0 1.0	1,100 1,300 1,300	1,00 1,20 1,30	0 240	140 160 160	64 70 71	0 54	8.4 8.6 8.6
Date	Sam- pling depth (m)	si di g sol n (m	as- 1 um, 1 .s- .ved ng/L	Alka- inity, abora- tory (mg/L as CaCO ₃)	Sulfate, dis- solved (mg/L as S04)	Chlo- ride, dis- solved (mg/L as Cl)	Fluo- ride, dis- solved (mg/L as F)	Silica, dis- solved (mg/L as SiO ₂)	Solids, sum of constit- uents, dis- solved (mg/L)
May, 1981 20	1.0		17	110	2,000	28	0.7	0.8	2,940
June 16	1.0		20	82	2,000	12	.7	.8	2,900
July 15	1.0		20	46	2,200	12	.8	•5	3,190
Aug 05 25 26	1.0 1.0 1.0		18 23 23	60 49 32	2,400 2,600 2,600	9.0 13 28	.7 1.0 1.0	3.4 .1 .7	3,460 3,770 3,790

Date	Time (24- hour)	Sam- pling depth (m)	(mg/L	bonate (mg/L as	- Calcium dis- solved (mg/L as Ca)	Magne , sium, dis- solved (mg/L as Mg)	- Sodium dis- solved (mg/L as Na)	Percent	Sodium ad- sorp- t tion ratio
			RESE	RVOIR 19-	KING RE	SERVOIR			
May, 1981 21 21	0732 0736	1.0 3.0	140 95	18 .00	34 35	13 18	55 54	44 52	2.0 2.4
June 16 16	1803 1807	1.0 3.0	140 150	21 28	35 36	13 14	61 57	47 44	2.2 2.0
July 15 15	1432 1436	1.0 3.0	150 150	25 25	35 35	14 14	65 63	47 47	2.3 2.3
Aug 05 27 27 28 28	1802 1806 1802 1806 0502 0506	1.0 3.0 1.0 3.0 1.0 3.0	120 110 110 110 110 110	3.0 .00 1.0 .00 10 .00	28 26 23 23 23 24	13 11 13 12 12 12	61 53 66 63 65 66	49 54 54 54 54 54	2.6 2.4 3.0 2.9 3.0 3.0
Date		Sam-		Alka- linity, labora- tory (mg/L as CaCO3)	Sulfate, dis- solved (mg/L as SO4)	ride, dis-	Fluo- ride, dis- solved (mg/L as F)	Silica, s dis- c solved (mg/L as s	Solids, sum of constit- uents, dis- solved (mg/L)
May, 1981 21 21		1.0 3.0	9.2 9.1	120 120	130 110	2.8 2.4	0.2	6.7 6.8	323 292
June 16 16		1.0 3.0	8.6 9.1	120 120	140 140	2.9 3.0	•2 •2	8.0 8.2	341 340
July 15 15		1.0 3.0	9.8 9.9	120 120	170 160	3.1 3.2	•2 •2	11 12	380 370
Aug 05 05 27 27 28 28		1.0 3.0 1.0 3.0 1.0 3.0	9.9 8.6 10 9.7 10 10	120 120 110 130 97 110	1 30 140 1 20 1 30 1 50 1 50	2.7 3.0 3.0 2.8 3.2 3.2 3.2	.5 .5 .2 .2 .2 .2 .0	12 11 14 14 14 14	329 326 315 333 336 346

Date	Time (24- hour)	Sam- pling depth (m)	Hard- ness (mg/L as CaCO3	bonate (mg/L as	Calcium, dis- solved (mg/L as Ca)	Magne- sium, dis- solved (mg/L as Mg)	Sodium dis- solvec (mg/L as Na)	l Percen	
			RESER	VOIR 24	EMPIRE RE	ESERVOIR			
May, 1981 20	1932	1.0	46	0.00	13	3.2	21	47	1.4
June 16	1433	1.0	55	.00	15	4.3	25	47	1.5
July 15	1802	1.0	66	.00	18	5.1	26	43	1.4
Aug 05 26 27	1431 1802 0502	1.0 1.0 1.0	57 56 56	.00 .00 .00	15 14 14	4.8 5.1 5.1	29 34 35	49 53 54	1.8 2.1 2.2
Date	Sam- pling depth (m)	- di g sol n (m	um, s-	Alka- linity, labora- tory (mg/L as CaCO ₃)	Sulfate, dis- solved (mg/L as S04)	Chlo- ride, dis- solved (mg/L as Cl)	Fluo- ride, dis- solved (mg/L as F)	Silica, s dis- solved (mg/L as	Solids, sum of constit- uents, dis- solved (mg/L)
May, 1981 20	1.0	5	.0	62	37	2.5	0.1	3.1	123
June 16	1.0	5	•6	83	23	2.6	.1	4.7	131
July 15	1.0	6	.0	100	6.0	17	.1	9.8	148
Aug 05 26 27	1.0 1.0 1.0	7	.6 .0 .2	120 140 130	2.0 1.0 2.0	3.3 3.8 2.9	.6 .2 .2	14 13 13	148 162 158

Table 5.--Selected plant nutrients

[Analyses by U.S. Geological Survey. Abbreviations and symbol: m, meter; mg/L, milligram per liter; <, less than]

Dat	e	Time (24- hour)	Sam- pling depth (m)	Nitro- gen, total (mg/L as N)	Nitro- gen, NO2+NO3 dis- solved (mg/L as N)	Nitro- gen, NO2+NO3 total (mg/L as N)	Nitro- gen, ammonia total (mg/L as N)	Nitro- gen, organic total (mg/L as N)	Phos- phorus, total (mg/L as P)	Phos- phorus, ortho, dis- solved (mg/L as P)	Carbon, organic, total (mg/L as C)
					RESERV	OIR 1AI	R BASE PO	ND	<u></u>		
		0832 0836	1.0 3.0	1.7 1.4	0.01	0.01	0.11 .11	1.6 1.3	0.08	0.01	24 17
		1503 1507	1.0 3.0	1.5 1.6	.01	.01	.07 .11	1.4 1.5	.06 .08	.04 .04	21 25
		1547 1551	1.0 3.0	1.8 1.9	.01 .02	.02 .03	.05 .13	1.8 1.8	.11 .12	.05 .05	12 13
04 24 24 25	••• ••• •••	1603 1607 1802 1806 0502 0506	1.0 3.0 1.0 3.0 1.0 3.0	2.0 1.9 1.7 1.7 	<.01 <.01 <.01 <.10 <.10	<.01 <.01 .01 <.10 <.10	.17 .32 .11 .20 .12 .19	1.8 1.6 1.5 1.8 1.6	.15 .23 .23 .23 .23 .23 .25	.09 .14 .12 .11 .02 .01	15 14 15 16 23 17
					RESERV	OIR 9GA	Y RESERVO	IR			
Мау, 20	1981	1403	1.0	1.8	.01	.01	.12	1.7	.10	<.01	17
June 16	•••	0903	1.0	1.4	.02	.01	.11	1.3	.06	.03	13
July 15	•••	0903	1.0	1.4	.02	.04	.11	1.3	.05	.05	8
25		0932 1802 0502	1.0 1.0 1.0	2.4 1.4	<.01 .10 <.10	<.01 <.10 <.10	.16 .10 .11	2.2 1.5 1.3	<.01 .04 .06	.01 .01 <.01	23 14 14

Date	Time (24- hour)	Sam- pling depth (m)	Nitro- gen, total (mg/L as N)	Nitro- gen, NO2+NO3 dis- solved (mg/L as N)	Nitro- gen, NO2+NO3 total (mg/L as N)	Nitro- gen, ammonia total (mg/L as N)	Nitro- gen, organic total (mg/L as N)	Phos- phorus, total (mg/L as P)	Phos- phorus, ortho, dis- solved (mg/L as P)	Carbon, organic, total (mg/L as C)
				RESERVO	IR 19KI	NG RESERV	OIR			
May, 21 21	1981 ••• 0732 ••• 0736	1.0 3.0	3.4 3.6	0.02	0.01	0.15	3.3 3.4	0.14	0.01	31 31
	1803 1807	1.0 3.0	3.4 3.6	.02 .02	<.01 <.01	.07 .08	3.3 3.5	.17 .17	.04 .03	37 39
	1432 1436	1.0 3.0	4.0 4.3	.02 .02	.03 .02	.20 .42	3.8 3.9	.16 .17	.02 .01	23 22
05 27 27 28	1802 1806 1802 1806 0502 0506	1.0 3.0 1.0 3.0 1.0 3.0	4.7 4.3 5.1 	<.01 <.01 <.10 <.10 .11 .12	<.01 <.01 <.10 .12 <.10 <.10	.23 .27 .15 .47 .19 .43	4.5 4.0 3.8 4.5 3.1 3.8	.01 .01 .17 .24 .17 .21	.01 .01 .02 .04 .01 .03	22 30 24 22 25 27
M	1091			RESERVOI	R 24EMP	IRE RESER	VOIR			
May, 20	1932	1.0	5.0	<.01	.02	.46	4.5	.22	.02	43
June 16	1433	1.0	4.7	.02	<.01	.06	4.6	.27	.04	35
July 15	1802	1.0	7.5	.02	•06	.13	7.3	.34	•04	30
26	1431 1802 0502	1.0 1.0 1.0	6.7 	<.01 <.10 <.10	<.01 <.10 <.10	.30 .17 .25	6.4 7.6 7.0	<.01 .32 .27	.02 .02 .02	31 36 40

	[Analyses		Geologic µg/L, mi						1,
Date	Time (24- hour)	Sam- pling depth (m)	Copper, dis- solved (µg/L as Cu)	Copper, total recov- erable (μg/L as Cu)	Iron, dis- solved (μg/L as Fe)	Lead, total recov- erable (µg/L as Pb)	Lead, dis- solved (µg/L as Pb)	Manga- nese, total recov- erable (μg/L as Mn)	Manga- nese, dis- solved (µg/L as Mn)
			RESERVO	DIR 1AI	R BASE F	POND	<u> </u>		
May, 198	1								
20		1.0		4	10	<1	~~~	50	10
20	0836	3.0		3	10	<1		50	20
June									
15	1503	1.0		4	30	12	~	90	7
15	1507	3.0		3	20	12		100	8
July									
14	1547	1.0		4	20	2	~-	50	10
14	1551	3.0		3	30	2	~-	70	20
Aug									
04	1603	1.0		6	15	<1		60	10
04	1607	3.0		5	16	<1		130	75
24	1802	1.0	1	4	11	6	4	80	21
24	1806	3.0	1	4	31	5	3	150	58
25	0502	1.0	1	4	20	3	<1	70	14
25	0506	3.0	2	4	25	39	<1	130	52
May, 198	1		RESERVO	DIR 9GA	Y RESERV	OIR			
20		1.0	~~	5	50	<1		760	660
June									
16	0903	1.0		3	70	15		190	120
July									
15	0903	1.0		4	50	<1		160	50
Aug									
05	0932	1.0		6	20	2		80	30
25		1.0	2	4	60	3	<1	40	20
26			2	5	60	6	4		
20	0502	1.0	2	Э	00	b	4	40	20

Table 6.--Trace elements

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Date	Time (24- hour)	Sam- pling depth (m)	Copper, dis- solved (µg/L as Cu)	Copper, total recov- erable (µg/L as Cu)	Iron, dis- solved (µg/L	Lead, total recov- erable (µg/L as Pb)	(µg/L	Manga- nese, total recov- erable (µg/L as Mn)	(µg/L
			RESERVO)IR 19K	ING RESE	ERVOIR			
May, 1981 21 21	0732 0736	1.0 3.0		4 3	10 20	1 1		310 350	3 9
June 16 16	1803 1807	1.0 3.0		4 3	40 <10	15		420 430	2 2
July 15 15	1432 1436	1.0 3.0		4 4	<10 10	<1 2		440 600	4 170
Aug 05 05 27 27 28 28	1802 1806 1802 1806 0502 0506	1.0 3.0 1.0 3.0 1.0 3.0	 2 2 2 2	11 7 5 5 5 5	11 12 24 42 24 30	1 55 55 55 60	 2 3 <1 <1	500 750 390 700 390 470	10 460 110 470 72 200
			RESERVOIR	₹ 24ЕМР	IRE RESE	RVOIR			
May, 1981 20	1932	1.0		8	280	1		200	40
June 16	1433	1.0		4	330	14		400	40
July 15	1802	1.0		5	110	2		930	<1
Aug 05 26 27	1431 1802 0502	1.0 1.0 1.0	 2 2	7 5 5	60 58 55	2 50 60	 3 3	780 780 830	6 7 9

[m, meter; mL, milliliter]

			RESI	ERVOIR 1-	-AIR BASE	POND		
DATE: TIME (24-HOUR): DEPTH:	0	y 20 832 .0 m	May 083 3.0		1:	ne 15 503 .0 m	June 150 3.0	7
	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent
BACILLARIOPHYTA		1.9		2.2		0.57		1.2
Bacillariophyceae (diatoms) Amphipleura paludosa	11	50					4	.13
Cyclotella spp. Cymbella minuta	11	.50	3	.12	0	0.5	4	.13
Epithemia sorex Gomphonema spp.	2	.09			8	.25	4 4	.13 .13
Navicula spp. Nitzschia acicularis	3	.14	3	.12	5 5	.16 .16	22	.69
N spp.	10		3	.12				•••
Synedra nana S. ulna	18 7	.82 .32	34 13	1.3 .50				
CHLOROPHYTA (green algae) Chlorophyceae		77		87		83		90
Actinastrum Hantzschii Ankistrodesmus falcatus	210	9.6	3 520	.12 20	290	9.1	260	8.1
Chodatella wratislavensis Coelastrum microsporum	48	2.2	6	.23			52	
Cosmarium spp.			38	1.5	64 3	2.0 .09		1.6
Crucigenia apiculata C. tetrapedia	8 120	.36 5.5	22 130	.85 5.0	59 790	1.8 25	61 1,000	1.9 31
Dictyosphaerium pulchellum Franceia ovalis	2	.09			21		.,	
Gloeocystic gigas	89	4.1			84	.66 2.6		
Oocystis crassa O. lacustris	10 5	.45 .23	9	.35	92	2.9		
0. parva	240	11	220	8.5	240	7.5	340	11
Pediastrum Boryanum P. duplex	38 23	1.7	13 13	.50 .50	49 64	1.5 2.0	78 48	2.4 1.5
P. tetras Scenedesmus abundans	300	14	460	18	150	4.7	4 340	.13
S. bijuga	30	1.4	53	2.0	160	5.0	160	5.0
S. dimorphus S. opoliensis	23	1.1	34 3	1.3	5	.16	13	.41
S. protuberans S. quadricauda	31 210	1.4 9.6	88 330	3.4 13	33 170	1.0 5.3	270	8.4
Staurastrum spp.			3	.12	3	.09	270	0.4
Tetraedron caudatum T. minimum	5 250	.23 11	190	7.3	59	1.8	70	2.1
T. pentaedricum Tetrastrum staurogeniaeform	2	.09	9 110	.35	3 320	.09 10	9 170	.28 5.3
-	ie 30		110		520		170	
CRYPTOPHYTA (cryptomonads) Cryptophyceae		5.1		4.3		16		7.2
Chroomonas sp. Crypt monas sp.	110 3	5.0 .14	110 3	4.2	190 330	5.9 10	230	7.2
CYANOPHYTA (blue-green algae)		4.9		8.4		.47		.94
Cyanophyceae Aphanizomenon flos-aquae	13	.59	3	.12	10		20	
Cyanarcus hamiformis	95	4.3	220	8.3	10 5	.31 .16	30	.94
EUGLENOPHYTA (euglenoids) Euglenophyceae		.32		.12				
Euglena spp.	7	.32	3	.12				
PYRRHOPHYTA (fire algae) Dinophyceae (dinoflagellates) Gymnodinium spp.	250	11 11						
Total number of cells ¹ Total number of taxa	2,200		2,600 30		3,200		3,200	

DATE: TIME (24-HOUR): DEPTH:	1	y 14 547) m	July 15 3.0	51		ist 4 503) m	Augus 160 3.0 r	7
	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent
BACILLARIOPHYTA Bacillariophyceae (diatoms)		.21		14		.55		7.2
Cyclotella spp. Gomphonema spp. Melosira granulata Navicula spp.	3 3 3	.07 .07 .07	14 3 69 3 3	1.5 .32 7.3 .32	13	.45	3 1	1.2
Nitzschia acicularis N. filformis N. spp. Rhoicosphemia curvata Synedra ulna	J	.07	3 34 3	.32 .32 3.6 .32	3	.10	13 1	.40 5.2 .40
CHLOROPHYTA (green algae) Chlorophyceae		92		79		54		52
Ankistrodesmus falcatus Chlamydomonas spp. Closteriopsis longissima Coelastrum microsporum	2,900	71	7 7 10	.74 .74 1.1	1,400	48	35 1	14 .40
Cosmarium spp. Crucigenia tetrapedia	36	.07			3	.10	1	.40
Oocystis crassa O. parva Pediastrum Boryanum	150 120	3.7 2.9	240 130	26 14			8	3.2 .40
P. duplex Quadrigula lacustris Scenedesmus abundans S. bijuga	9 3	.22 .07	3 3 28	.32 .32 3.0	5	.17	1	.40 .40
S. dimorphus S. guadricauda Schroederia setigera Sphaerocystis Schroeteri	16 520	.39 13	7 58 230 3	.74 6.2 24 .32	5 160	.17 5.5	1 13 66	.40 5.2 26
Tetraedron minimum CRYPTOPHYTA (cryptomonads)		4.2	17	1.8 7.3	3	.10 38	1	.40 38
Cryptophyceae Chroomonas sp. Cryptomonas sp.	22 150	.54 3.7	69	7.3	610 490	21 17	18 77	7.2 31
CYANOPHYTA (blue-green algae) Cyanophyceae		4.5				6.6		2.4
Anabaenae circinalis Aphanizomenon flos-aquae Oscillatoria spp. Spirulina spp.	3 180	.07 4.4			190	6.6	4 1 1	1.6 .40 .40
EUGLENOPHYTA (euglenoids) <i>Buglena</i> spp.							2	.80 .80
Total number of cells ¹ Total number of taxa	4,100 16		940 22		2,900		250 21	

RESERVOIR 1--AIR BASE POND--Continued

	RESERVO	IR 1AIR	BASE PON	DCont.	RESERVOIR 9GAY RESERVOIR				
DATE: TIME (24-HOUR): DEPTH:	Ŭ18	st 24 302 .0 m	Augus 18 3.		17	y 20 403 .0 m	June 0903 1.0	3	
	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	
BACILLARIOPHYTA Bacillariophyceae (diatoms)		3.1		23		0.13		1.1	
Cocconies placentula Gyrosigma macrum	2	.15	3	1.2			2	.10	
G. spp. Melosira granulata Navicula spp. Vitanabia arian baia	33	2,54	2 42	.80 17			1 19	.05	
Nitzschia acicularis N. spp.	5	.38	10	4.0	3	.13	19	• • • •	
CHLOROPHYTA (green algae) Chlorophyceae		45		17		21		39	
Actinastrum Hantzschii Ankistrodesmus falcatus					12	.52	440 45	22 2.2	
Carteria spp. Chlamydomnas spp.	400	31	7	2.8	470	20	290	15	
Cosmarium spp. Oocystis spp. Quadrigula lacustris Scenedesmus abundans	2	.15 .31	3 3 5	1.2 1.2 2.0					
S. dimorphous S. quadricauda	2 4	.15 .31	10	4.0					
S. quadificatua Schroederia setigera Sphaerocystis Schroeteri Tetraedron minimum	170	13	10 10 5	4.0 2.0					
CRYPTOPHYTA (cryptomonads) Cryptophyceae		33		59		1.0			
Chroomonas Sp. Cryptomonas Sp.	200 240	15 18	1 2 0 2 8	48 11	23	1.0			
CYANOPHYTA (blue-green algae) Cyanophyceae		14		2.0		54		45	
Anabaena Augustumalis	5	.38			440	19	28	1.4	
A. circinalis A. spiroides			2	1 0	800	35	5	.25	
Aphanizomenon flos-aquae Coelosphaerium Kuetzingianu Gomphosphaeria lacustris	170 m	13	3 2	1.2 .80			64	3.2	
Lyngbya Sp. Microcystis aeruginosa	5	.38			9	.39	01	5.1	
Oscillatoria spp. Pseudoanabaena spp.	2	.15					810	40	
EUGLENOPHYTA (euglenoids)						23		4.8	
Euglenophyceae Euglena acus Lepocinclis spp. Phacus spp.					100 410 6	4.4 18 .26	95	4.8	
PYRRHOPHYTA (fire algae) Dinophyceae (dinoflagellates)		.69				1.4		9.8	
Ceratium hirudinella Gymnodinium spp.	2	.15			32	1.4	190	9.5	
Peridinium spp.	7	.54					7	.35	
Total number of cells ¹ Total number of taxa	1,300 17		250 15		2,300		2,000		

Table 7.--Taxa, percentage composition, and number of phytoplankton collected during 1981--Continued

DATE: TIME (24-HOUR): DEPTH:	09	у 15 03 0 m	Ì	gust 5 0932 1.0 m	Augus 180 1.0		
	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	
BACILLARIOPHYTA		65		1.2		2.9	
Bacillariophyceae (diatoms) Amphipleura paludosa Chaetoceros muelleri Cyclotella spp. Gyrosigma macrum	14 10 13 17	.67 .48 .62 .81			16 170	.23 2.4	
Navicula Spp. Nitzschia acicularis	1,300	62	150	1.2	16	.23	
CHLOROPHYTA (green algae) Chlorophyceae		18		20		1.7	
Actinastrum Hantzschii Ankistrodesmus falcatus Carteria SPP. Dictyosphaerium pulchellum	6 140 130 10	.29 6.7 6.2 .48	400 2,100	3.1 16	16 47	.23 .66	
Scenedesmus opoliensis Tetraedron minimum Treubaria setigerum	10 55 34	.48 2.6 1.6	62 53 9	.48 .41 .07	40 16	.56 .23	
CRYPTOPHYTA (cryptomonads) Cryptophyceae		3.6		40		30	
Chroomonas Sp. Cryptomonas Sp.	58 17	2.8 .81	2,100 3,100	16 24	900 1,200	13 17	
CYANOPHYTA (blue-green algae) Cyanophyceae		2.0		33		55	
Anabaena spiroides Gomphosphaeria lacustris Merismopedia tenuissima	7 27 7	.33 1.3 .33	1,800 9 18	14 .07 .14	190 32	2.7 .45	
Pseudoanabaena Spp.	,	•33	2,500	19	3,700	5 2	
EUGLENOPHYTA (euglenoids) Euglenophyceae		1.4		.96		.11	
Euglena acus Phacus Spp.	27 3	1.3 .14	71 53	•55 •41	8	•11	
PYRRHOPHYTA (fire algae) Dinophyceae (dinoflagellates)		10		2.2	1.5	10	
Glenodinium spp. Gymnodinium spp.	130	6.2			16	.23	
Peridinium spp.	79	3.8	280	2.2	740	10	
Total number of cells ¹ Total number of taxa	2,100 21		13,000 15		7,100 15		

RESERVOIR 9--GAY RESERVOIR--Continued

		KESEKVOIK 19KING KESEKVOIK									
DATE: TIME (24-HOUR): DEPTH:	0	у 21 732 .0 ш	May 07 3.		1	ne 16 803 .0 m	June 16 1807 3.0 m				
	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent			
BACILLARIOPHYTA		6.4		13		14		15			
Bacillariophyceae (diatoms) Amphipleura paludosa			30	.52							
Nitzschia dissipata	51	.13	50	• 52							
N. palea spp.	2,400	6.0	650	11	3,700	10	1,000	10			
Rhizosolenia longiseta Stephanodiscus Spp.	100	.25	59	1.0	76 1,400	.21 3.9	450	4.6			
CHLOROPHYTA (green algae) Chlorophyceae		20		41		68		63			
Actinastrum Hantzschii Ankistrodesmus falcatus	2,300 1,900	5.8 4.8	1,000 510	17.24 8.8	4,800	13	700 28	7.1			
Chodatella wratislavensis Coelastrum microsporum Dictyosphaerium pulchellum Dimorphococcus lunatus Elakatothrix gelatinosa Franceia ovalis	610 556	1.5 1.4			2,400 11,000 300 300 210	6.7 31 .83 .83 .58	450 3,100 450	4.6 32 4.6			
Pediastrum tetras Scenedesmus abundans S. dimorphous S. opoliensis	200 1,100	.50 2.8	450	7.8	300 530 1,600	.83 1.5 4.4	140 280 250	1.4 2.9 2.6			
S. quadricauda S. spp.	560	1.4	300	5.2	1,300	3.6	56	.57			
Selenastrum minutum Tetraedron minimum Tetrastrum staurogeniaefor	610 mae 250	1.5 .63	89 30	1.5 .52	150 1,600 76	.42 4.4 .21	340 220 170	3.5 2.2 1.7			
CRYPTOPHYTA (cryptomonads)		3.0		11		3.9		4.6			
Cryptophyceae Cryptomonas Sp.	1,200	3.0	620	11	1,400	3.9	450	4.6			
CYANOPHYTA (blue-green algae) Cyanophyceae		69		34		12		17			
Anabaena spiroides A. spp.	100	.25	180	3.1	76	.21					
Aphanocapsa elachista	760	1.9					56	.57			
Merismopedia tenuissima Oscillatoria Spp. Pseudoanabaena Spp.	2,000 25,000	5.0 62	540 1,300	9.3 22	4,300	12	1,600	16			
EUGLENOPHYTA (euglenoids) Euglenophyceae		.13		.52		.63		.57			
Euglena Spp.					76	.21					
Phacus Spp. Trachelomonas hispida T. Spp.	51	.13	30	.52	76 76	.21 .21	56	.57			
Total number of cells ¹ Total number of taxa	40,000 18		5,800 14		36,000		9,800 18				

RESERVOIR 19--KING RESERVOIR

DATE: TIME (24-HOUR): DEPTH:	14	ly 15 432 .0 m	14	у 15 36 0 m	ĭ	ust 5 802 .0 m	Augus 180 3.0	5
	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	Célls per mL	Per- cent
BACILLARIOPHYTA Bacillariophyceae (diatoms)				1.4		1.0		1.4
Nitzschia Spp.							300	.64
Stephanodiscus spp.			570	1.4	430	1.0	380	.81
CHLOROPHYTA (green algae) Chlorophyceae		63		84		76		7 9
Actinastrum Hantzschii	63	.15						
Ankistrodesmus falcatus	9,600	22	14,000	34	7,100	17	13,000	28
Chodatella wratislavensis					1,000	2.3	1,400	3.0
Coelastrum microsporum	1,000	2.3	760	1.8	6,500	15	4,800	10
Crucigenia apiculata	250	.58	95 950	.23 2.3	160	.37	380	.81
C. spp. Dictyosphaerium pulchellum	250	.58	1,200	2.3	830	1.9		
Elakatothrix gelatinosa	63	.15	1,200	2.5	000	1.5	150	.32
Golenkinea radiata	5,000	12	5,600	14	1,100	2.6	160	.34
Scenedesmus abundans	1,300	3.0	2,000	4.9	2,800	6.5	2,300	4.9
S. dimorphous	3,900	9.1	3,100	7.6	8,400	20	7,800	17
S. opoliensis	250	.58	570	1.4	490	1.1	830	1.8
S. quadricauda					100 330	.23	300	.64
S. spp. Selenastrum minutum	2,000	4.7	3,300	8.1	1,000	.77 2.3	76 2,100	.16 4.5
Tetraedron minimum	2,800	6.5	1,900	4.6	1,900	4.4	2,300	4.9
T. trigonum	_,		.,		.,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		2,000	.16
Tetrastrum staurogeniaefor.		1.6	760	1.8	810	1.9	1,200	2.6
Treubaria setigerum	63	.15	190	.46			150	.32
CRYPTOPHYTA (cryptomonads) Cryptophyceae		30		11		18		4.2
Chroomonas sp.	13,000	30	4,700	11	6,400	15	1,600	3.4
Cryptomonas sp.	130	.30			1,500	3.5	380	.81
CYANOPHYTA (blue-green algae) Cyanophyceae		4.7		3.0		5.6		16
Aphanothece nidulans					380	.88	76	.16
Chroococcus Prescottii					430	1.0		
Gomphosphaeria spp.			95	.23			230	.49
Merismopedia tenuissima	250	•58	280	.68			6,600	14
Microcystis aeruginosa Oscillatoria SPP.	1,000	2.3	660	1.6	380 1,200	.88 2.8	76 460	.16
Pseudoanabaena spp.	760	1.8	190	.46	1,200	2.0	400	.98
EUGLENOPHYTA (euglenoids)		.30				.36		
Euglenophyceae					100			
Euglena spp. Phacus spp.	130	.30			100 54	.23		
rnacus spp.	100	.30			54	•13		
Total number of cells ¹ Total number of taxa	43,000		41,000		43,000		47,000	

RESERVOIR 19--KING RESERVOIR--Continued

	RESERVOIF	R 19КІ	NG RESERVC	IR-Cont.	RESERVOIR 24EMPIRE RESERVOIR				
DATE: TIME (24-HOUR): DEPTH:	August 180 1.0)2	August 180 3.0)6	1	y 20 932 .0 m	June 16 1433 1.0 m		
	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	Cells per mL	Per- cent	
BACILLARIOPHYTA		.23		1.9		46		29	
Bacillariophyceae (diatoms) Cyclotella spp. Navicula spp. Nitzschia acicularis N. palea	84	.23	510 84	1.6 .26	190 140 7,100	1.2 .88 44	350 15,000	.66 28	
CHLOROPHYTA (green algae)		86		76		9.9		24	
Chlorophyceae Actinastrum Hantzschii Ankistrodesmus falcatus Chodatella wratislavensis Coelastrum microsporum Cosmarium Spp.	11,000 670	31 1.7	8,400 2,000	26 6.2	240 810	1.5 5.1	4,200 230 120	7.9 .43 .23	
Corucigenia apiculata Dictyosphaerium pulchellum Gloeocystis spp. Kirchneriella contorta Pediastrum duplex	840 340	2.3 .94	1,000	3.1	47	.29	120 120 120	.23	
P. tetras Scenedesmus abundans S. dimorphous S. opoliensis	170 1,300 11,000 420	.47 3.6 31 1.2	340 11,000	1.1 34	140	.88	230 1,900	.43 3.5	
S. quadricauda S. spp. Selenastrum Bibraianum S. minutum Tetraedron minimum Tetrastrum staurogeniaeforu Treubaria setigerum	1,000 1,700 mae 1,800 590	2.8 4.7 5.0 1.6	170 1,000 510	.53 3.1 1.6	95 240	.59 1.5	930 120 3,400 1,000 230 230	1.8 .23 6.4 1.9 .43 .43	
CRYPTOPHYTA (cryptomonads)		6.9		3.9		15		22	
Cryptophyceae Chroomonas Cryptomonas Sp.	2,300 170	6.4 .47	1,000 250	3.1 .78	660 1,700	4.1 11	470 1 1, 000	.89 21	
CYANOPHYTA (blue-green algae)		7.8		17		26		14	
Cyanophyceae Anabaena affinis Aphanocapsa elachista	250	.69	420	1.3	760	4.8	1,600	3.0	
A. spp. Chroccccus spp. Merismopedia tenuissima Microcystis aeruginosa	1,200 420	3.3 1.2	5,000	16			120 230 2,800	.23 .43 5.3	
Pseudoanabaena Spp.	930	2.6			3,300	21	2,900	5.5	
EUGLENOPHYTA (euglenoids) Euglenophyceae		.46		1.6		2.4		11	
Euglena spp. Buglena spp. Phacus spp. Trachelomonas hispida T. spp.	84 84	.23 .23	510	1.6	140 240	.88 1.5	230 120 1,000 4,400	.43 .23 1.9 8.3	
Total number of cells ¹ Total number of taxa	36,000 21		32,000		16,000 15		53,000 26		

Table 7.--Taxa, percentage composition, and number of phytoplankton collected during 1981--Continued

Table 7Taxa,	composition, during 1981-	of	phytoplankton

per mL cent per <ml< th=""> cent <th< th=""><th></th><th>RESERVOI</th><th>K 24EP</th><th>IFIKE KESEI</th><th>KVUIRCO</th><th>ntinued</th><th></th></th<></ml<>		RESERVOI	K 24EP	IFIKE KESEI	KVUIRCO	ntinued	
per mL cent per mL cent per mL cent per mL cent BACILLARIOPHYTA BacIllariophyceae (diatoms) Cyclotella striata 810 1.4 220 .28 76 .27 Mitszchia acigularis 3.600 6.4 10,000 13 14,000 2.21 N. palea 3.600 6.4 10,000 13 14,000 2.21 Rhizosolenia longiseta 76 51 36 2.00 2.6 Chlorophyceae Ankistrodesmus falcatus 9,400 17 11,000 14 11000 17 Cocalastrum microsporum 2,500 4.5 5,000 6.4 460 .21 Castrum microsporum 2,100 38 1,600 1.5 1,100 1.6 Castrum microsporum 2,000 4.5 3,000 4.5 1,000 1.6 Scandesus abundans 50 5.7 1.00 1.7 3.00 1.6 1.5 S. struatus 200 .36 3.50	TIME (24-HOUR):	18	302		Í431	180	02
Bacillariophyceae (diatoms) 7<							Per- cent
Cyclotelia striata 810 1.4 220 .28 Mitzschia acicularis 3600 6.4 10,000 13 14,000 22 N. spp. 710 1.3 330 .42 1,500 2.4 CHLOROPHYTA (green algae) 76 51 36 36 Chlorophycae Ankistrodesmus falcatus 9,400 17 11,000 14 11,000 17 Codastrum microsporus 2,500 4.5 5,300 4.2 76 5 Cosastrum microsporus 2,100 38 1,200 1.5 1,100 1.6 Diarophococcus lunatus 14,000 25 3,500 4.5 1,100 1.6 S. acumatus 2,000 3.6 10 .14 76 .1 S. acumatus 2,000 3.6 100 .2 .100 .2 .100 .2 .2 .100 .2 .2 .100 .2 .2 .100 .2 .2 .2 <td< td=""><td></td><td></td><td>9.1</td><td></td><td>14</td><td></td><td>25</td></td<>			9.1		14		25
n. palea 3,600 6.4 10,000 13 14,000 22 Rhizosolenia longiseta 710 1.3 330 .42 1,500 2.4 Schubophytra (green algae) 76 51 36 36 330 .42 76 .5 Chodrophyteae Ankistrodesmus falcatus 9,400 17 11,000 14 11,000 17 Coclastrum microsporum 2,500 4.5 5,000 6.4 460 .5 Coclastrum microsporum 2,100 3.8 1,200 1.5 1,100 1.4 Crucispen crucifera 510 .91 .16 .14 76 .1 Scenedesmus abundans 300 .54 110 .14 76 .1 S. erouatus 2,000 4.1 2,300 4.1 2,000 .2 S. erouatus 510 .91 .900 2.4 1,200 .2 S. erouatus 510 .91 .900 .2 .2 </td <td>Cyclotella striata</td> <td>810</td> <td>1.4</td> <td>220</td> <td>.28</td> <td>76</td> <td>.12</td>	Cyclotella striata	8 10	1.4	220	.28	76	.12
Rhizosolenia longiseta 150	N. palea					14,000	22
Chlorophyceae 200 17 11,000 14 11,000 17 Chodatella quadriseta 200 .36 330 .42 76 .5 Correctislavensis 330 .42 76 .5 .500 6.4 460 .5 Correctislavensis 910 1.6 1.600 2.1 1.700 2.5 Correctislavensis 510 .91 1.600 1.600 2.1 1.700 2.5 Flarkatothrix virisis 300 .54 110 .14 76 .1 Scenedesmus abundans 2.300 .54 110 .14 76 .1 S. arcuatus 2.300 .54 170 1.2 .6 .1 S. arcuatus 2.300 .51 .5 .6 .5 .6 .5 .6 .5 .5 .6 .5 .6 .5 .6 .5 .6 .6 .6 .6 .6 .6 .5 .5 .6 .5 .6 .5 .5 .6 .5 .5 .5 <		/10	1.5	330	•42		.24
Ankistrodesmus falcatus 9,400 17 11,000 14 11,000 14 Chodatella quadriseta 200 .36 220 .28 150 .2 Corratislavensis 330 .42 76 .3 .42 76 .3 Corratisma sirrosporum 2,500 4.5 5,000 6.4 460 Corratigna crucifera 510 .91 .60 2.1 1,700 2 Crucigna crucifera 2,100 3.8 1,200 1.5 Primorphococcus lunatus 14,000 25 3,500 4.5 1.00 1.6 Scenedesmus abundans			76		51		36
c. vratislavensis 330 .42 76 Ccelastrum microsporum 2,500 4.5 5,000 6.4 460 Crucigena crucifera 510 .911 1.60 1.600 2.1 1,700 2.500 Crucigena crucifera 510 .911 .901 1.6 1.600 2.1 1,700 2.500 Dimorphococcus lunatus 14,000 2.5 3,500 4.5 1,100 1.6 Pediastrum horphococus lunatus 2,000 .54 110 .14 76 .1 Scenedesmus abundans 2,000 .54 .10 .14 76 .1 S. arcusinatus 2,000 .36 .30 .42 .150 .2 S. arcusinatus 1,000 .91 .900 .42 .1,200 1.5 S. protubersans 400 .71 .330 .42 .42 .100 .2 S. protubersans 100 .18 .540 .69 1,400 .2 .2 S. gracilis 2,000 .36 .50 .69 .2	Ankistrodesmus falcatus						17 .24
Cosmarium spp. 910 1.6 1,600 2.1 1,700 2.5 Crucigena crucifera 510 .91 .91 .50 1.5 1.5 Dimorphococcus lunatus 14,000 25 3,500 4.5 1,100 1.6 Pediastrum Boryanum 300 .54 110 .14 76 .1 Scaenedesmus abundans 2,300 4.1 2,300 3.0 680 1.1 S. arcmatus 200 .36 .54 .100 .24 1,200 1.5 S. arcmatus 200 .36 .510 .91 1,900 2.4 1,200 1.5 S. producto-capitatus 510 .91 1,900 2.4 1,200 1.5 S. quadricauda 1,200 .1 1,300 1.7 230 .5 S. ercuberans 100 .18 540 .69 .400 .5 S. ercuberans 2,000 .36 220 .28 .69 .		200	• 50				.12
Crucigena crucifera 510 .91 C. spp. 2.100 3.8 1,200 1.5 Dimorphococcus lunatus 14,000 25 3,500 4.5 1,100 1.6 Elakatothrix viridis 300 .54 110 .14 76 .1 Scenedesmus abundans 300 .54 76 .1 <	Coelastrum microsporum						.73
c. spp. 2,100 3.8 1,200 1.5 Dimorphococcus lunatus 14,000 25 3,500 4.5 1,100 1.6 Pediastrum Boryanum 300 .54 110 .14 76 .5 Pediastrum Boryanum 300 .54 76 .5 .6 .5 Scenedesmus abundans 2,300 4.1 2,300 3.0 680 1.1 S. atoutinatus 2,300 .6 .6 .6 .6 .6 .6 .6 .6 .6 .7				1,600	2.1	1,700	2.1
Dimorphococcus lunatus 14,000 25 3,500 4.5 1,100 1.6 Elakatothrik viridis 300 .54 110 .14 76 .1 P. duplex 300 .54 76 .1				1,200	1.5		
Pediastrum Boryanum 110 .14 P. duplex 300 .54 76 Scenedesmus abundans 2,300 4.1 2,300 3.0 680 1.1 S. acuminatus 200 .36 .30 .30 680 1.1 S. acuminatus 200 .36 .30 .42 150 .5 S. producto-capitatus 510 .91 1,900 2.4 1,200 1.5 S. quadricauda 1,200 2.1 .970 1.2 .76 S. eprotuberans 400 .71 .330 .42 150 S. guadricauda 1,200 2.1 .970 1.2 .76 S. eprotuberans 100 .18 .540 .69			25			1,100	1.8
p. duplex 76 76 Scenedessus abundans 2,300 4.1 2,300 3.0 680 1.1 S. acuminatus 200 .36 .30 680 1.1 S. atimorphous 510 .91 .900 2.4 1,200 1.6 S. producto-capitatus 510 .91 1,900 2.4 1,200 1.5 S. producto-capitatus 510 .91 1,900 2.4 1,600 2.5 S. gradicicauda 1,200 2.1 970 1.2 76 .1 S. vertucosus 100 .18 540 .69 .400 2.5 S. gracilis 200 .36 .54 .69 .60 .60 .60 7.8 1,700 2.7 S. westii .000 .18 .76 .97 .60 1.2 .76 .1 Tetrastrum staurogeniaeformae 200 .36 .220 .28 .76 .1 Tetrastrum staurogeniaeformae 200 .36 .220 .28 .76 .1		300	.54	110	• /	150	.24
Scene desmus abundans 76 76 S. acuminatus 200 .36 S. dimorphous 510 .91 S. producto-capitatus 510 .91 S. producto-capitatus 510 .91 S. producto-capitatus 510 .91 S. quadricauda 1,200 2.1 970 S. quadricauda 1,200 .18 540 .69 S. sudricauda 1,000 .18 540 .69 .200 S. gracilis 200 .36 .69 .200 .36 S. mutum 2,600 4.6 6,100 7.8 1,700 2.7 S. westii 76 .7 .7 .7 .9 .2 .000 .36 .200 .2 .36 .2 .2 .36 .1 .3 .30 .4 .30 .2 .36 .2 .2 .2 .36 .2 .2 .36 .2 .36 .2 .36 .1 <t< td=""><td></td><td>300</td><td>5.4</td><td>110</td><td>•14</td><td>76</td><td>.12</td></t<>		300	5.4	110	•14	76	.12
s. accuminatus 2,300 4.1 2,300 3.0 680 1.1 s. arcuatus 200 .36 .30 .30 .30 .30 .30 .30 .30 .30 .30 .30 .30 .30 .30 .30 .30 .30 .50 .30 .30 .50 .30 .30 .50 .30 .30 .50 .30 .30 .42 .30 .42 .50 .50 .50 .50 .50 .24 .100 .12 .76 .7 .30 .42 .50 .50 .50 .50 .610 1.1 1.300 .1.7 .230 .30 .50 .50 .50 .600 1.60 .1.8 .500 .50 <td></td> <td>500</td> <td>•74</td> <td></td> <td></td> <td></td> <td>.12</td>		500	•74				.12
S. dimorphous 510 .91 .900 2.4 1,200 1.5 S. producto-capitatus 510 .91 1,900 2.4 1,200 1.5 S. protuberans 400 .71 330 .42 150 .5 S. protuberans 100 .18 540 .69 1,400 2.2 S. sepn. 610 1.1 1,300 1.7 230 .5 Selenastrum Bibraianum 1,000 1.8 540 .69 .60 S. westii .760 .7 .700 .7 .700 .7 Tetradoron hastatum 100 .18 .760 .97 .760 .2 T. minum 2,400 4.3 1,600 2.1 2,000 .3 Tetradoron hastatum 100 .18 .760 .28 .76 .1 Tetrastrum staurogeniaeformae 200 .36 220 .28 .76 .1 Cryptophyceae .1,300 2.3 .540 .69 1,200 1.5 Cyanophyceae .1,200<		2,300	4.1	2,300	3.0		1.1
S. producto-capitatus 510 91 1,900 2.4 1,200 1.5 S. protuberans 400 .71 330 .42 150 .2 S. quadricauda 1,200 2.1 970 1.2 76 .1 S. verrucosus 100 .18 540 .69 1,400 2.2 S. sproin 610 1.1 1,300 1.7 230 .2 S. gracilis 200 .36 .69 .2 .2 .2 S. westii 76 .1 .700 2.1 .100 .18 T. minimum 2,600 4.6 6,100 7.8 1,700 2.7 S. Westii 76 .2 .000 .36 .200 .36 .200 .36 T. trigonum 2,400 4.3 1,600 2.1 2,000 3.2 Tretrastrum staurogeniaeformae 200 .36 .20 .28 .28 .15 Cryptophyceae 1,300 2.3 .540 .69 1,200 1.5 Cryp	S. arcuatus	200					
s. protuberans 400 .71 330 .42 150 s. quadricauda 1,200 2.1 970 1.2 s. verrucosus 100 .18 540 </td <td></td> <td></td> <td></td> <td>1 000</td> <td>o (</td> <td>1 000</td> <td>• •</td>				1 000	o (1 000	• •
s. quadricauda 1,200 2.1 970 1.2 76 s. verrucosus 100 .18 540 .69 1,400 2.2 s. spp. 610 1.1 1,300 1.7 230 .5 sclenastrum Bibraianum 1,000 1.8 540 .69 .69 s. gracilis 200 .36 .69 .69 .7 .76 .7 s. westii 100 .18 76 .1 .700 3.2 T. trigonum 2,400 4.3 1,600 2.1 2,000 3.2 T.trigonum 760 .97 760 1.2 .76 .15 Tetrastrum staurogeniaeformae 200 .36 220 .28 .76 .1 Treubaria setigerum 2.3 .69 1.5 .76 .7 .76 .7 RYPTOPHYTA (cryptomonads) 2.3 .540 .69 1,200 1.5 .30 .69 .69 .20 .5 YANOPHYTA (blue-green algae) 11 29 .5 .50 .4 <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>.24</td>							.24
S. verrucosus 100 .18 540 .69 1,400 2.2 S. spp. 610 1.1 1,300 1.7 230 .5 Selenastrum Bibraianum 1,000 1.8 540 .69 .69 S. gracilis 200 .36 .36 .36 .69 .17 230 .5 S. westii 76 .36 .600 4.6 6,100 7.8 1,700 2.5 Tetraedron hastatum 100 .18 76 .1 .60 .97 760 1.2 T. mininum 2,400 4.3 1,600 2.1 2,000 3.2 Tetradoron hastatum 100 .18 76 .1 .12 Tetradoron material setigerum 220 .28 .69 1.2 .12 Cryptophyceae 1,300 2.3 .540 .69 1,200 1.5 Cyanophyceae 1,300 2.3 .540 .69 1,200 1.5 Cyanophyceae 15,000 19 5,800 .5 .6 .6							.12
s. spp. 610 1.1 1,300 1.7 230 .5 Selenastrum Bibraianum 1,000 1.8 540 .69 .69 S. gracilis 200 .36 .69 .69 .7 .7 .700 2.7 S. westii 100 .18 .600 4.6 6,100 7.8 1,700 2.7 T. trigonum 2,400 4.3 1,600 2.1 2,000 3.2 T. trigonum 760 .97 760 1.7 Tetrastrum staurogeniaeformae 200 .36 220 .28 76 .1 Cryptophyceae 220 .28 76 .1 .500 1.5 .200 1.5 Cryptophyceae 1,300 2.3 540 .69 1.200 1.5 Cyanophyceae 11 29 15 .530 .2 .30 .4 Aphanocapsa spp. 1,300 2.5 5.4 .21 .200 .5 Werismopedia tenuissima 15,000 19 5,800 .300 .4 <	-						2.2
S. gracilis 200 .36 S. minutum 2,600 4.6 6,100 7.8 1,700 2.7 S. Westii 100 .18 76 .1 Tetraedron hastatum 100 .18 76 .1 T. minimum 2,400 4.3 1,600 2.1 2,000 3.2 T. trigonum 760 .97 760 1.2 .1 1.2 .1 1.2 .1					1.7		.37
s. minutum 2,600 4.6 6,100 7.8 1,700 2.5 s. Westii 76 76 76 76 76 Tetradron hastatum 100 .18 76 76 76 76 T. minimum 2,400 4.3 1,600 2.1 2,000 3.2 Tetrastrum staurogeniaeformae 200 .36 220 .28 76 .1 Tetrastrum staurogeniaeformae 200 .36 220 .28 76 .1 Cryptophyceae .1,300 2.3 .69 1.5 .1 .200 1.5 Cyanophyceae .1,300 2.3 540 .69 1.200 1.5 Cyanophyceae .1,300 2.3 540 .69 1.200 1.5 Coyanophyceae .1,300 2.3 .540 .69 1.200 1.5 Coyanophyceae .1,200 .1.5 .200 .5 .6 .20 .20 .20 Colosphaerium spp. .5,900 11 6,800 8.7 1,900 .6 </td <td></td> <td></td> <td></td> <td>540</td> <td>.69</td> <td></td> <td></td>				540	.69		
S. Westii 100 .18 76 .1 Tetraedron hastatum 2,400 4.3 1,600 2.1 2,000 3.2 T. trigonum 760 .97 760 1.2 Tetrastrum staurogeniaeformae 200 .36 220 .28 76 .1 Terubaria setigerum 220 .28 76 .1 .1 .20 .28 .1 ERYPTOPHYTA (cryptomonads) 2.3 .69 1.200 1.5 .200 1.5 Cryptophyceae 1,300 2.3 540 .69 1.200 1.5 YANOPHYTA (blue-green algae) 11 29 15 .500 .69 .200 1.5 Cyanophyceae 11 29 .5 .69 .200 .5 .69 .200 .5 Anabaena affinis 1,200 .5 .54 .21 .200 .5 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 .20 <td< td=""><td></td><td></td><td></td><td>6 100</td><td>7 0</td><td>1 700</td><td>0 7</td></td<>				6 100	7 0	1 700	0 7
Tetraedron hastatum 100 .18 T. minimum 2,400 4.3 1,600 2.1 2,000 3.2 T. trigonum 760 97 760 1.2 Tetrastrum staurogeniaeformae 200 .36 220 .28 76 .1 Treubaria setigerum 220 .28 76 .1 .100 .18 CRYPTOPHYTA (cryptomonads) 2.3 .69 1.5 .200 1.5 Cryptophyceae 0 11 29 15 .530 .6 Cyanophyceae 11 29 15 .530 .6 .530 .6 XANOPHYTA (blue-green algae) 11 29 15 .530 .6 .2 .6 .7 .6 .7 .6 .7 .7 .6 .7 .6 .7		2,000	4.0	0,100	/.0		.12
T. minimum 2,400 4.3 1,600 2.1 2,000 3.2 T. trigonum 760 .97 760 1.2 Tetrastrum staurogeniaeformae 200 .36 220 .28 76 .1 Treubaria setigerum 220 .28 76 .1 .1 20 .28 76 .1 Cryptophyceae 220 .28 .69 1.200 1.5 Cryptophyceae 1,300 2.3 .69 1.200 1.5 Cyanophyceae 11 29 15 .530 .6 Anabaena affinis 1,200 1.5 .530 .6 Aphanocapsa Spp. 870 1.1 230 .6 Coelosphaerium Spp. 870 1.1 230 .6 Merismopedia tenuissima 15,000 19 5,800 .6 Oscillatoria spp. 5,900 11 6,800 8.7 1,900 .6 Cuglenophyceae 510 .91 1,200 1.5 1,200 .6 Euglenophyceae 650		100	.18			,.	•12
Tetrastrum staurogeniaeformae 200 .36 220 .28 76 .1 Treubaria setigerum 220 .28 76 .1 Cryptophyceae 220 .28 76 .1 Cryptophyceae 1,300 2.3 .69 1.9 Cryptophyceae 1,300 2.3 540 .69 1,200 1.5 Cyanophyceae 11 29 15 530 .6 .69 .200 1.5 XANOPHYTA (blue-green algae) 11 29 15 530 .6 .69 .200 1.5 XANOPHYTA (blue-green algae) 11 29 15 .5 .6 .69 .200 1.5 Anabaena affinis 1,200 1.5 .200 .23 .6 .5 .6 .6 .6 .23 .6 .2 .6 .2 <td></td> <td>2,400</td> <td></td> <td></td> <td>2.1</td> <td></td> <td>3.2</td>		2,400			2.1		3.2
Treubaria setigerum 220 .28 CryptoPHYTA (cryptomonads) 2.3 .69 1.5 Cryptophyceae 1,300 2.3 540 .69 1.200 1.5 Cryptomonas sp. 1,300 2.3 540 .69 1.200 1.5 Cyanophyceae 11 29 15 .530 .6 .69 .6							1.2
Cryptophyceae 1,300 2.3 540 .69 1,200 1.5 CYANOPHYTA (blue-green algae) 11 29 15 Cyanophyceae 11 29 15 Ana baena affinis 1,200 1.5 A. Spp. 1,200 1.5 Aphanocapsa Spp. 76 .530 .6 Coelosphaerium Spp. 870 1.1 230 .5 Merismopedia tenuissima 15,000 19 5,800 9.2 Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.6 Euglenophyceae 2.5 5.4 21 21 200 .36 .300 .4 Phacus Spp. 510 .91 1,200 1.5 1,200 1.5 Phacus Spp. 510 .91 1,200 1.5 1,200 1.6 Phacus Spp. 910 1.6 2,400 3.1 11,000 17 Pracus Spp. 910 1.6 2,400 3.1 11,000 17 Prexidinium spp. 200 .36 </td <td></td> <td>mae 200</td> <td>.36</td> <td></td> <td></td> <td>76</td> <td>.12</td>		mae 200	.36			76	.12
Cryptomonas sp. 1,300 2.3 540 .69 1,200 1.5 CYANOPHYTA (blue-green algae) 11 29 15 Cyanophyceae 11 29 15 Anabaena affinis 1,200 1.5 A. spp. 30 .6 Aphanocapsa Spp. 76 .6 Coelosphaerium spp. 870 1.1 230 .6 Merismopedia tenuissima 15,000 19 5,800 9.2 Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.0 EUGLENOPHYTA (euglenoids) 2.5 5.4 21 21 Euglenophyceae 300 .2 .300 .2 .2 .2 .20 .36 .28 .300 .2 <td></td> <td></td> <td>2.3</td> <td></td> <td>.69</td> <td></td> <td>1.9</td>			2.3		.69		1.9
Cyanophyceae 1,200 1.5 Ana baena affinis 1,200 1.6 A. spp. 300 .6 Aphanocapsa Spp. 76 .76 Coelosphaerium spp. 15,000 19 5,800 9.2 Merismopedia tenuissima 15,000 19 5,800 9.2 Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.0 SUGLENOPHYTA (euglenoids) 2.5 5.4 21 Euglenophyceae 300 .4 300 .4 Fhacus spp. 510 .91 1,200 1.5 1,200 1.5 Phacus spp. 510 .91 1,200 1.5 1,200 1.5 T. spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .3 .3 Dinophyceae .36 .28 .3 .3 (dinoflagellates) .200 .36 220 .28 .3 Peridinium spp. 200 .36 220 .28		1,300	2.3	540	.69	1,200	1.9
A. spp. 530 .6 Aphanocapsa Spp. 76 .76 Coelosphaerium Spp. 15,000 19 5,800 9. Merismopedia tenuissima 15,000 19 5,800 9. Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.0 EUGLENOPHYTA (euglenoids) 2.5 5.4 21 Euglenophyceae .300 .4 Fhacus spp. 510 .91 1,200 1.5 1,200 1.6 Phacus spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .5 .5 .5 .5 Dinophyceae .36 .28 .5 .5 .5 .5 .5 VYRRHOPHYTA (fire algae) .36 .28 .5 .5 .5 .5 .5 Peridinium spp. 200 .36 .20 .28 .5 .5 Cotal number of cells ¹ 56,000 78,000 63,000 .5 .5			11		29		
Aphanocapsa Spp. 76 Coelosphaerium spp. 870 1.1 230 Merismopedia tenuissima 15,000 19 5,800 9.2 Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.0 Euglenophyceae 2.5 5.4 21 Euglena spp. 510 .91 1,200 1.5 1,200 1.5 Phacus Spp. 510 .91 1,200 1.5 1,200 1.6 Phacus spp. 510 .91 1,200 1.5 1,200 1.6 Phacus spp. 910 1.6 2,400 3.1 11,000 17 PytRRHOPHYTA (fire algae) .36 .28 .1 .1 .1 .1 Dinophyceae (dinoflagellates) .200 .36 .220 .28 .2 Peridinium spp. 200 .36 220 .28 .2 .2 Cotal number of cells ¹ 56,000 78,000 63,000 .3 .3 .3							1.9
Coelosphaerium spp. 870 1.1 230 230 Merismopedia tenuissima 15,000 19 5,800 92 Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.0 SUGLENOPHYTA (euglenoids) 2.5 5.4 21 Euglenophyceae 510 .91 1,200 1.5 1,200 1.5 Phacus spp. 510 .91 1,200 1.5 1,200 1.5 Trachelomonas hispida 650 .83 990 1.6 T. spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .3 Dinophyceae .36 .28 .3 (dinoflagellates) 200 .36 220 .28 .3 Peridinium spp. 200 .36 220 .28 .3 Cotal number of cells ¹ 56,000 78,000 63,000 .3							.84
Merismopedia tenuissima Oscillatoria spp. 15,000 19 5,800 9.2 Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.6 EUGLENOPHYTA (euglenoids) 2.5 5.4 21 Euglenophyceae 510 .91 1,200 1.5 1,200 1.5 Phacus spp. 510 .91 1,200 1.5 1,200 1.5 Trachelomonas hispida 650 .83 990 1.6 T. spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .30 .30 .30 Dinophyceae .36 .28 .30 .30 .30 (dinoflagellates) .200 .36 .220 .28 .30 Peridinium spp. 200 .36 220 .28 .30 Gotal number of cells ¹ 56,000 78,000 63,000 63,000				870	1 1		.12 .37
Oscillatoria spp. 5,900 11 6,800 8.7 1,900 3.0 EUGLENOPHYTA (euglenoids) 2.5 5.4 21 Euglenophyceae 510 .91 1,200 1.5 1,200 1.9 Phacus spp. 510 .91 1,200 1.5 1,200 1.6 Trachelomonas hispida 650 .83 990 1.6 T. spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .36 Dinophyceae .36 .28 .36 (dinoflagellates) 200 .36 220 .28 .36 Fotal number of cells ¹ 56,000 78,000 63,000 .300							9.2
Euglenophyceae 510 .91 1,200 1.5 1,200 1.5 Phacus spp. 300 .4 Trachelomonas hispida 650 .83 990 1.6 T. spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .36 Dinophyceae .36 .28 .36 Peridinium spp. 200 .36 220 .28 .36 Cotal number of cells ¹ 56,000 78,000 63,000 63,000		5,900	11		8.7		3.0
Euglena spp. 510 .91 1,200 1.5 1,200 1.5 Phacus spp. 300 .200 .300 .200 .300 .200 Trachelomonas hispida 650 .83 990 1.6 .400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .36 .28 .36 Dinophyceae (dinoflagellates) .200 .36 .20 .28 .36 Peridinium spp. 200 .36 220 .28 .36 .36 Cotal number of cells ¹ 56,000 78,000 63,000 .300 .300 .300			2.5		5.4		21
Phacus spp. 300 Trachelomonas hispida 650 .83 990 1.6 T. spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .36 Dinophyceae .36 .28 .36 Peridinium spp. 200 .36 220 .28 .6 Total number of cells ¹ 56,000 78,000 63,000 63,000		510	.91	1.200	1.5	1,200	1.9
Trachelomonas hispida 650 .83 990 1.6 T. spp. 910 1.6 2,400 3.1 11,000 17 PYRRHOPHYTA (fire algae) .36 .28 .36 Dinophyceae .36 .28 .36 Peridinium spp. 200 .36 220 .28 76 .36 Total number of cells ¹ 56,000 78,000 63,000 63,000 63,000				_,			.48
Dinophyceae (dinoflagellates) 200 .36 220 .28 76 .7 Peridinium spp.		910	1.6				1.6
(dinoflagellates) 200 .36 220 .28 76 .1 Peridinium spp. Cotal number of cells ¹ 56,000 78,000 63,000			.36		.28		.12
Total number of cells ¹ 56,000 78,000 63,000	(dinoflagellates)	200	.36	220	28	76	.12
	.erraruram ohh.		• 50		•20		•12
Total number of taxa 31 31 36							
	lotal number of taxa	31		31		36	

RESERVOIR 24--EMPIRE RESERVOIR--Continued

¹Total number of cells rounded to two significant figures. Percents are calculated from unrounded values.

			Rese	rvoir	1Air Base	Pond		
Date: Time (24-hour): Depth:	083	May 20 0832 1.0 m		May 20 0836 3.0 m		15 3 m	1.	e 15 507 0 m
	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent
ROTIFERA Monogononta		3.6				2.3		
Ploima Brachionus Sp. Keratella cochlearis	2	3.6			2	2.3		
ARTHROPODA Crustacea Cladocera		96				98		100
Bosmina longirostris Daphnia ambigua Daphnia thorata Copepoda	8	14			11 33 2	12 38 2.3	18 130 55	6.2 45 19
Cyclops bicuspidatus thomasi	45	82	120	100	40	46	88	30
Total number of organisms ¹ Total number of taxa	55 3		120		88 5		290 4	

[m, meter; L, liter]

	Reservoir 1Air Base PondContinued									
Date: Time (24-hour): Depth:	July 14 1547 1.0 m		July 155 3.0	1	August 4 1603 1.0 m		August 1607 3.0 m			
	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent		
ARTHROPODA Crustacea Cladocera		100	······	100		100		100		
Bosmina longirostris Daphnia ambigua Daphnia thorata	11 9 23	17 14 35	4 28 73	2.9 20 53	10 67 40	4.6 31 19	2 44 85	.73 16 31		
Copepoda Cyclops bicuspidatus thomasi	23	35	34	24	99	46	140	52		
Total number of organisms ¹ Total number of taxa	66 4		140 4		220 4		270 4			

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	Reser		Air Base Cont.		Reservoir 9Gay Reservoir				
Date: Time (24-hour): Depth:			August 24 1806 3.0 m		May 20 1403 1.0 m		June 16 0903 1.0 m		
	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	
ARTHROPODA Crustacea Cladocera	110,	100		100		100		100	
Bosmina longirostris Daphnia ambigua Daphnia thorata Copepoda	2 2	11 11	2 43 4	2.7 59 5.5			4	100	
Cyclops bicuspidatus thomasi	15	79	24	33	22	100			
Total number of organisms ¹ Total number of taxa	19 3		73 4		22 1		4 1		

Table 8.--Taxa, percentage composition, and number of zooplankton collected during 1981--Continued

	Rese	rvoir	9Gay Rese	rvoir-	-Continued	
Dat Time (24-hour Dept): 0903		August 0932 1.0 m		August 25 1802 1.0 m	
	Organisms per L		Organisms per L	Per- cent	Organisms per L	Per- cent
ARTHROPODA Crustacea Copepoda		100		100		100
Cyclops bicuspidatus thomasi	9	100	12	100	16	100
Total number of organisms ¹ Total number of taxa	9 1		12 1		16 1	

Table 8Taxa,	percentage	composition,	and number	of
zooplankton	collected a	during 1981(Continued	

			Reservo	ir 19	-King Reser	voir		
Date: Time (24-hour): Depth:	May 21 0732 1.0 m		May 21 0736 3.0 m		June 16 1803 1.0 m)	June 16 1807 3.0 m	
	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent
ARTHROPODA Crustacea Cladocera		100		100		100		100
Bosmina longirostris Daphnia ambigua Daphnia catawba Copepoda	44 20	21 9.3	10 20 4	3.8 7.0 1.5	6 2	2.2	2 4 2	3.3 1.6
Cyclops bicuspidatus thomasi	130	63	210	81	43	46	69	57
Diaptomus Sp.	16	7.5	16	6.	1 48	52	46	38
Total number of organisms ¹ Total number of taxa	210 4		260 5		93 3		120 4	

	Reservoir 19King ReservoirContinued									
Date: Time (24-hour): Depth:	July 15 1432 1.0 m		July 15 1436 3.0 m		August 5 1802 1.0 m		August 5 1806 3.0 m			
	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent		
ARTHROPODA Crustacea Cladocera		100		100		100		100		
Bosmina longirostris Daphnia ambigua Daphnia catawba	2 4	6.2 12			2 2	3.8 3.8	3 3 4	8.3		
Copepoda Cyclops bicuspidatus thomasi	14	44	58	35	13	25	5	10		
Diaptomus sp.	12	38	106	65	36	68	39	81		
Total number of organisms ¹ Total number of taxa	32 4		170 2		53 4		48 3			

			19King rCont.		Reservoir 24Empire Reservoir				
Date: Time (24-hour): Depth:	August 27 1802 1.0 m		August 27 1806 3.0 m		May 20 1932 1.0 m		June 16 1433 1.0 m		
	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent	
ROTIFERA					·······				
Monogononta Ploima								5.5	
Keratella cochlearis							11	5.5	
ARTHROPODA Crustacea Cladocera		100		100		100		94.47	
Bosmina longirostris	2	7.7					24	12	
Daphnia ambigua	2 9						2	1.0	
Daphnia catawba Copepoda	4	35 15	2	1.5					
Cyclops bicuspidatus thomasi			8	5.9	18	100	160	80	
Diaptomus sp.	11	42	120	93			3	1.5	
± £			<u> </u>						
Total number of organisms ¹ Total number of taxa	26 4		130 3		18 1		200 5		

Reservoir 24--Empire Reservoir--Continued

Date: Time (24-hour): Depth:	July 15 1802 1.0 m		August 1431 0.5 m	5	August 26 1802 1.0 m	
	Organisms per L	Per- cent	Organisms per L	Per- cent	Organisms per L	Per- cent
ARTHROPODA Crustacea Cladocera		100		100		100
Bosmina longirostris Daphnia ambigua Copepoda	49 2	34 1.4			7 2	10 2.9
Cyclops bicuspidatus thomasi	80	56	20	50	50	74
Diaptomus sp.	12	8.4	20	50	9	13
Total number of organisms ¹ Total number of taxa	140		40 2		68 4	

¹Total number of cells are rounded to two significant figures. Percents are calculated from unrounded values.

Table 9.--Taxa, percentage composition, and number of benthic invertebrates collected during August 1981

[m ² , s	square meter]	
	RESERVOIR 1AIR B	ASE POND
Middle of	4.6 meters	At

	······		REDERVOI	IK 1AIF	THE TONE			
Sampling location:	Middle of reservoir		4.6 meters (15 feet) from shore		At shore		Average	
	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent
ANNELIDA Oligochaeta (aquatic worms)	1,900	31 31	820	14 14	87	50 50	940	23 23
ARTHROPODA Insecta		67		85		25		75
Diptera (true flies) Ceratopogomidae Chironomidae	87	1.4	87	1.5			44	1.1
Chironomus Sp. Chryptochironomus Sp. Glyptotendipes Sp.	3,400 87	56 1.4	4,000 130	68 2,2			2,500 29 43	61 .71 1.1
Procladius sp. Hemiptera (aquatic bugs)	480	7.7	780	13			420	10
Corixidae	43	.70			43	25	29	.71
MOLLUSCA Gastropoda (snails)		2.1		1.5		25		2.1
Lymnaeidae <i>Lymnaea</i> sp. Physidae			43	.73			14	.34
Physa Sp. Pelecypoda (bivalves)					43	25	14	•34
Sphaeriidae	130	2.1	43	.73			58	1.4
Total number of organisms ¹ Total number of taxa	6,100 7		5,900		170		4,100	

	RESERVOIR 9GAY RESERVOIR								
Sampling location:	Middle of reservoir		4.6 meters (15 feet) from shore		At shore		Average		
	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	
ANNELIDA Hirudinea (leeches) Oligochaeta (aquatic worms)	220	12 12	43 2,900	67 .98 66	170	9.3 9.3	14 1,100	41 .52 41	
ARTHROPODA Crustacea Amphipoda (scuds) Talitridae		23		29		80		39	
Hyallela azteca Insecta			87	1.99 26.72	173	9.29 69.78	87	3.23 35.82	
Diptera (true flies) Ceratopogonidae Chironomidae			43	.98			14	.52	
Chironomus Sp. Chryptochironomus Sp. Einfeldia Sp. Glyptotendipes Sp. Parachironomus Sp. Paratanytarsus Sp. Procladius Sp.	170 170	9.3 9.3	390 350 220	8.9 7.9 4.9	87 130 87 130 87	4.7 7.0 4.7 7.0 4.7	58 220 120 120 29 43 29	2.2 8.0 4.3 1.1 1.6 1.1	
Ephemeroptera (mayflies) Baetidae Baetis sp.	43	2.3					14	.52	
Caenidae <i>Caenis</i> s p.	+3	213			170	9.3	58	2.2	
Hemiptera (aquatic bugs) Corixidae Odonata (dragonflies)			170	4.0	220	12	130	4.8	
Coenagrionidae Ishnura sp.	43	2.3			390	21	140	5.3	
MOLLUSCA Gastropoda (snails) Physidae		65		4.0		12		20	
Physa sp. Planorbidae	390	21			130 87	7.0 4.7	170 29	6.4 1.1	
Pelecypoda (bivalves) Sphaeriidae	820	44	170	4.0			330	12	
Total number of organisms ¹ Total number of taxa	1,900		4,400		1,900		2,700		

Table 9.--Taxa, percentage composition, and number of benthic invertebrates collected during August 1981--Continued

	RESERVOIR 19KING RESERVOIR								
Sampling location:	Middle of reservoir		4.6 meters (15 feet) from shore		At shore		Average		
	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	
ANNELIDA Oligochaeta (aquatic worms)	2,300	82 82	1,500	28 28			1,300	32 32	
ARTHROPODA Crustacea Amphipoda (scuds)		18		70		98		67	
Talitridae Hyallela azteca Insecta Diptera (true flies)			43	.83	87	2.3	43	1.1	
Ceratopogonidae Chironomidae			43	.83	43	1.1	29	.74	
Chironomidae Chironomus sp Chryptochironomus Sp. Einfeldia Sp.	480	17	3,000 43 220	58 .83 4.2	2,600 740	68 20	2,000 260 72	51 6.6 1.8	
Procladius Sp. Hemiptera (aquatic bugs)	43	1.5	43	.83	130	3.4	72	1.8	
Corixidae Odonata (dragonflies) Coenagrionidae			220	4.2			72	1.8	
Ishnura Sp.					130	3.4	43	1.1	
MOLLUSCA				1.7		2.2		1.5	
Gastropoda (snails) Planorbidae					43	1.1	14	.36	
Pelecypoda (bivalves) Sphaeriidae			87	1.7 1.7	43	1.1 1.1	43	1.1 1.1	
Total number of organisms ¹ Total number of taxa	2,800		5,200		3,800 8		3,900 11		

Table 9.--Taxa, percentage composition, and number of benthic invertebrates collected during August 1981--Continued

Sampling location:	Middle of reservoir		4.6 me (15 fe from s	eet)	At		Average		
	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	Organisms per m ²	Per- cent	
ANNELIDA Hirudinea (leeches) Oligochaeta (aquatic worms)	43 1,300	31 1.0 30	87 11,000	92 .74 91	820	5.9 5.9	43 4,200	43 43	
ARTHROPODA Crustacea Amphipoda (scuds) Talitridae Hyallela azteca		69		8.5	12,000	94 85	4,000	57 40	
Insecta Diptera (true flies) Chironomidae Chironomus Sp. Chryptochironomus Sp. Einfeldia Sp. Glyptotendipes Sp.	2,900	68	820 87 43	7.0 .74	820 87	5.9 .62	1,200 29 270 29	12 .29 2.8 .29 .14	
Kiefferulus sp. Procladius sp. Hemiptera (aquatic bugs) Corixidae Notonectidae Notonecta sp.	43	1.0	43	.37 .37	350	2.5	14 14 120 14	.14 .14 1.2 .14	
Total number of organisms ¹ Total number of taxa	4,300		12,000		14,000		9,900 11		

RESERVOIR 24--EMPIRE RESERVOIR

¹Total number of organisms rounded to two significant figures. Percents are calculated from unrounded values.

Table 10.--Bacterial concentration of samples collected from May through August 1981

[Concentration in number of organisms per 100 milliliters.	$\langle . $ less than; \rangle . greater than
(·,, ·, B,

							I	Bacter	ia				
Sample information	Fecal coliform (FC)			ł	Fecal streptococci (FS)				FC/FS ratio				
	-					RESE	RVOIR	1AI	R BAS	E POND			
Date	5-20	6-15	7-14	8-4	8-24	5-20	6-15	7-14	8-4	8-24	5-20	6-15	7-14 8-4 8-24
Concentration at mid-reservoir	² 4	² 10	² 45	² 4	21	² 7	² 16	23	² 6	74	³ .57	3,63	2.0 ³ .67 .2
Concentration near shore	² 2	² 14	35	² 5	27	² 5	² 13	31	74	37	³ .40	³ i.1	1.1 ³ .068 .7
						RESERVOIR 9GAY RESERVOIR							
Date	5-20	6-16	7-15	8-5	8-25	5-20	6-16	7-15	8-5	8-25	5-20	6-16	7-15 8-5 8-2
Concentration at mid-reservoir	² 1	² 3	21	² 5	60	² <1	² 13	² 18	² 9	² 2	³ >1.0	³ .23	1.2 ³ .56 30
Concentration near shore	² 1	² 3	² 12	² 12	48	² <1	² 5	180	36	28	³ >1.0	³ .60	³ .067 ³ .33 1.7
						RESER	VOIR 1	9KII	NG RES	SERVOIR			
Date	5-21	6-16	7-15	8-5	8-27	5-21	6-16	7-15	8-5	8-27	5-21	6-16	7-15 8-5 8-2
Concentration at mid-reservoir	750	130	² 20	² 25	² 45	170	120	² 20	25	<5	4.4	1.1	³ 1.0 ³ 5.0 ³ >9.0
Concentration near shore	² 1,100	² 330	² 22	² 35	² 20	208	378	² 18	² 10	² 40	³ 5.3	³ .87	³ 1.2 ³ 3.5 ³ .5
						RESER	VOIR 2	4EMI	PIRE H	RESERVOI	R		
Date	5-20	6-16	7-15	8-5	8-26	5-20	6-16	7-15	8-5	8-26	5-20	6-16	7-15 8-5 8-2
Concentration at mid-reservoir	² 990	² 110	² 90	² 10	120	390	85	135	160	² 20	³ 2.5	³ 1.3	³ .67 ³ .063 ³ 6.0
Concentration near shore	560	100	100	² 65	160	370	78	87	² 50	² 35	1.5	1.3	1.2 ³ 1.3 ³ 4.6

 1 Fecal coliform organisms per 100 milliliters divided by fecal streptococcal organisms per 100 milliliters

 $^2\ {\rm Estimated}$ concentration based on nonideal colony count

³ FC/FS ratio based on nonideal colony counts

*GPO 691 - 922 (1985)